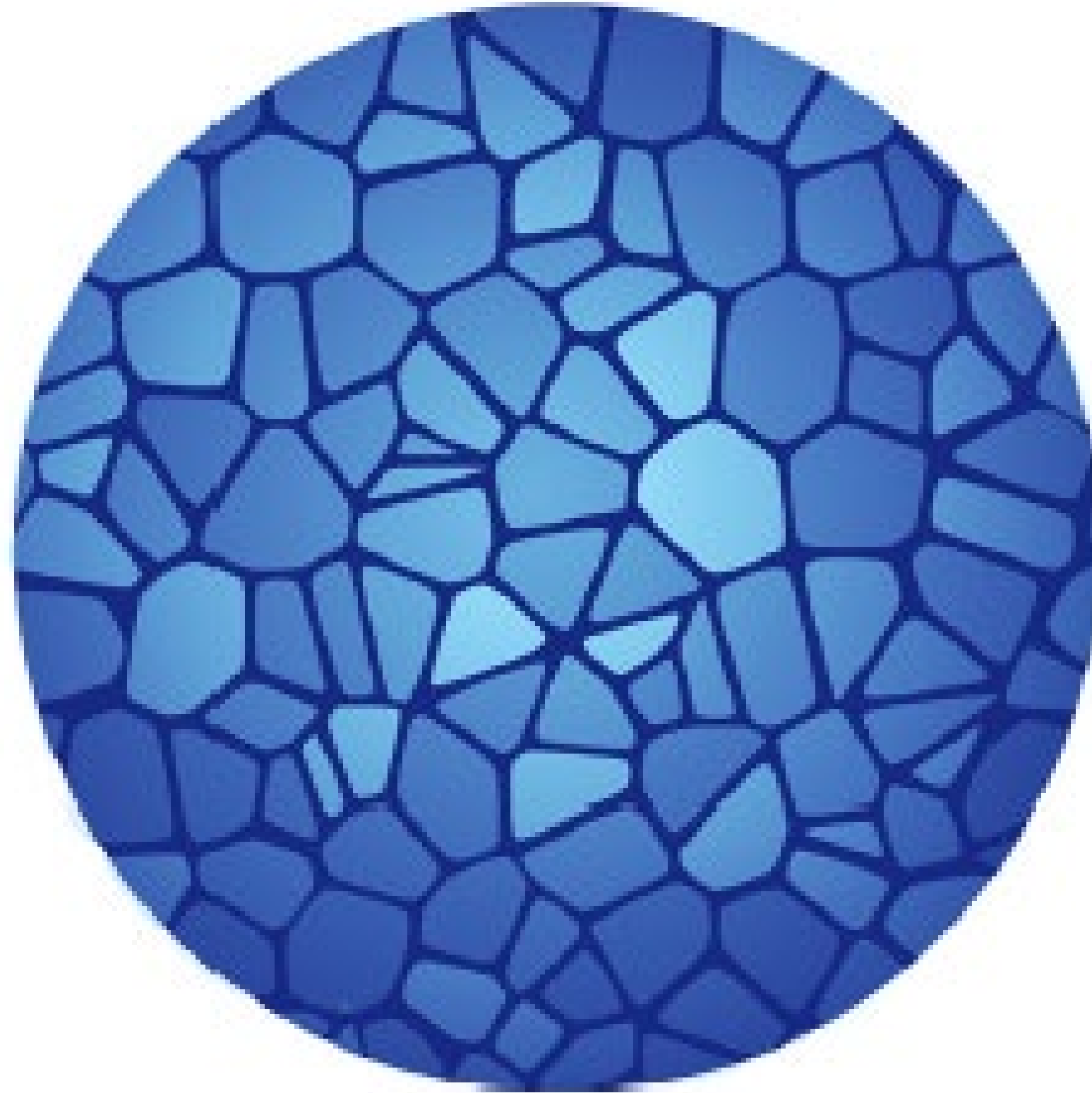


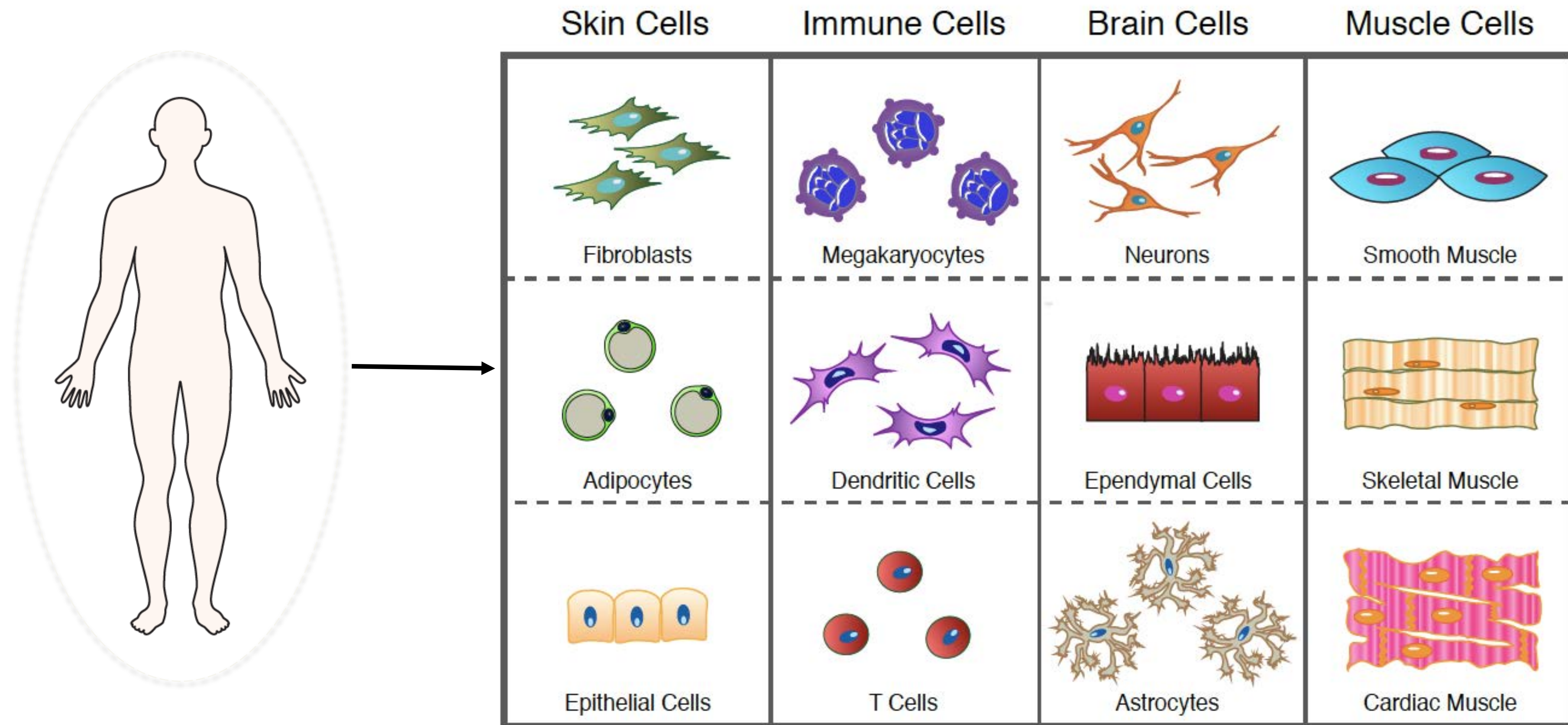
# The Human Cell Atlas



Ed Lein, Ph.D  
Investigator, Allen Institute for Brain Science  
Seattle, Washington USA  
Member, Organizing Committee, Human Cell Atlas

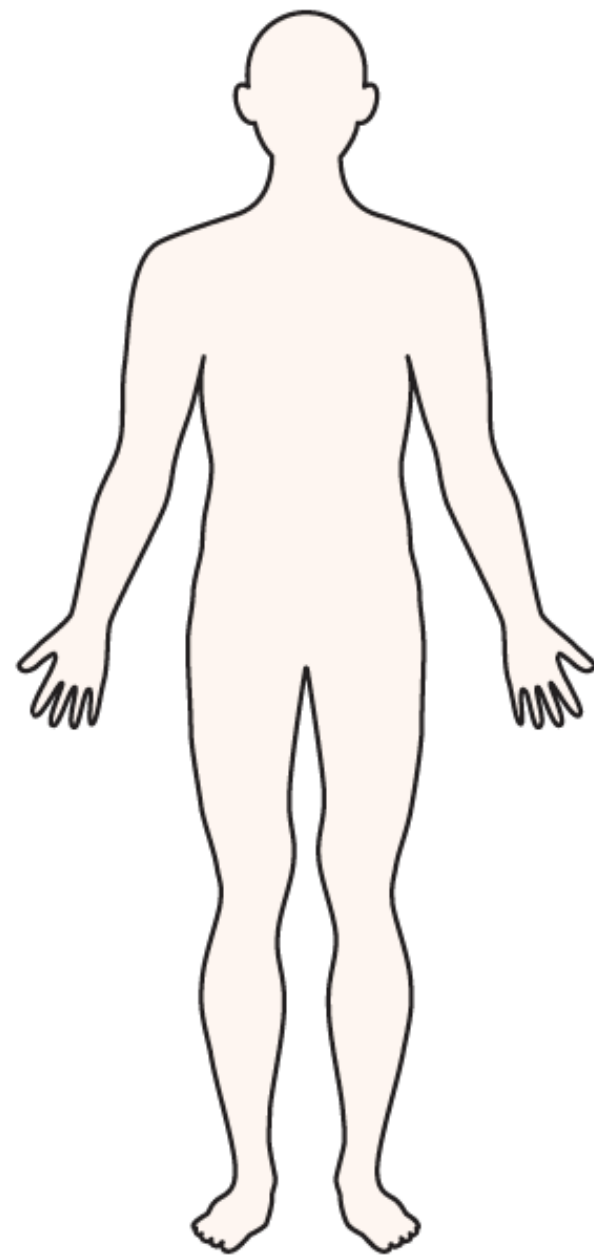
Moonshot International Symposium  
Tokyo, Japan  
December 18, 2019

# Cells are our basic units



Cells are classified by structure, location, function, molecules

# Problem: we do not really know our cells



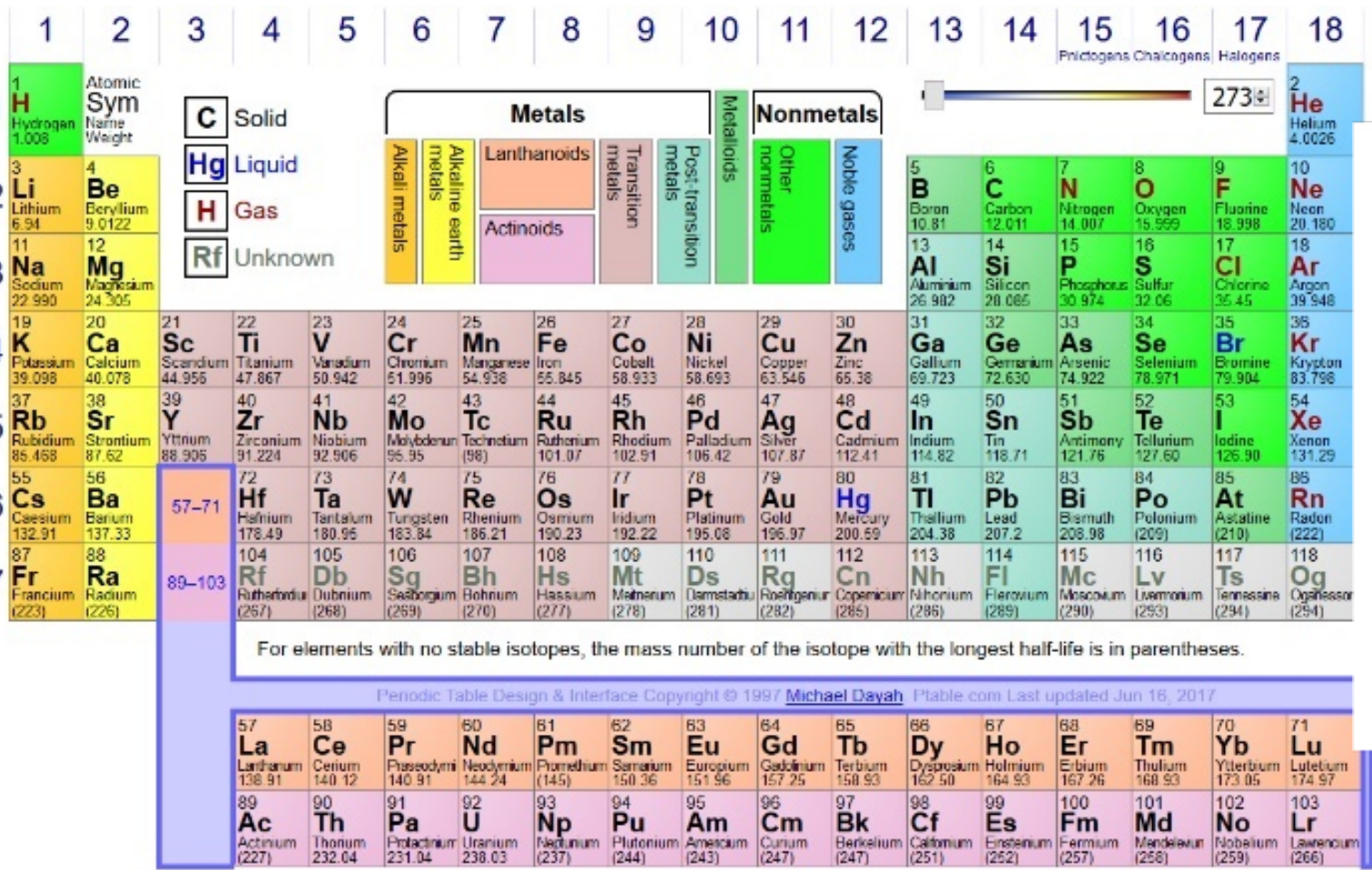
- ~37.2 trillion cells
- Text book: ~300 'major' cell types?
- Science: ~100 sub-sub-types of neurons just in the retina, or a single region of the brain's neocortex



# Human Cell Atlas: Mission

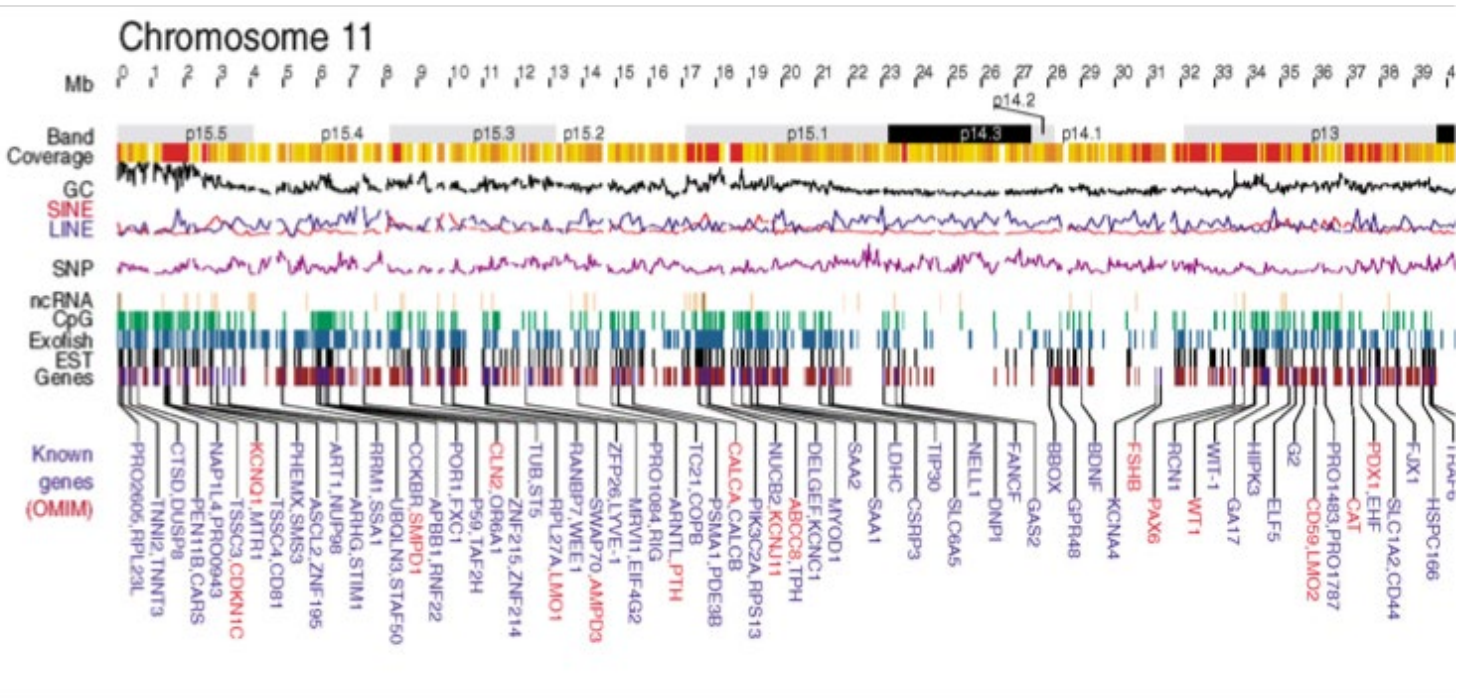
To create a comprehensive reference map of the types and properties of all human cells, the fundamental unit of life, as a basis for understanding, diagnosing, monitoring, and treating health and disease

## Chemistry



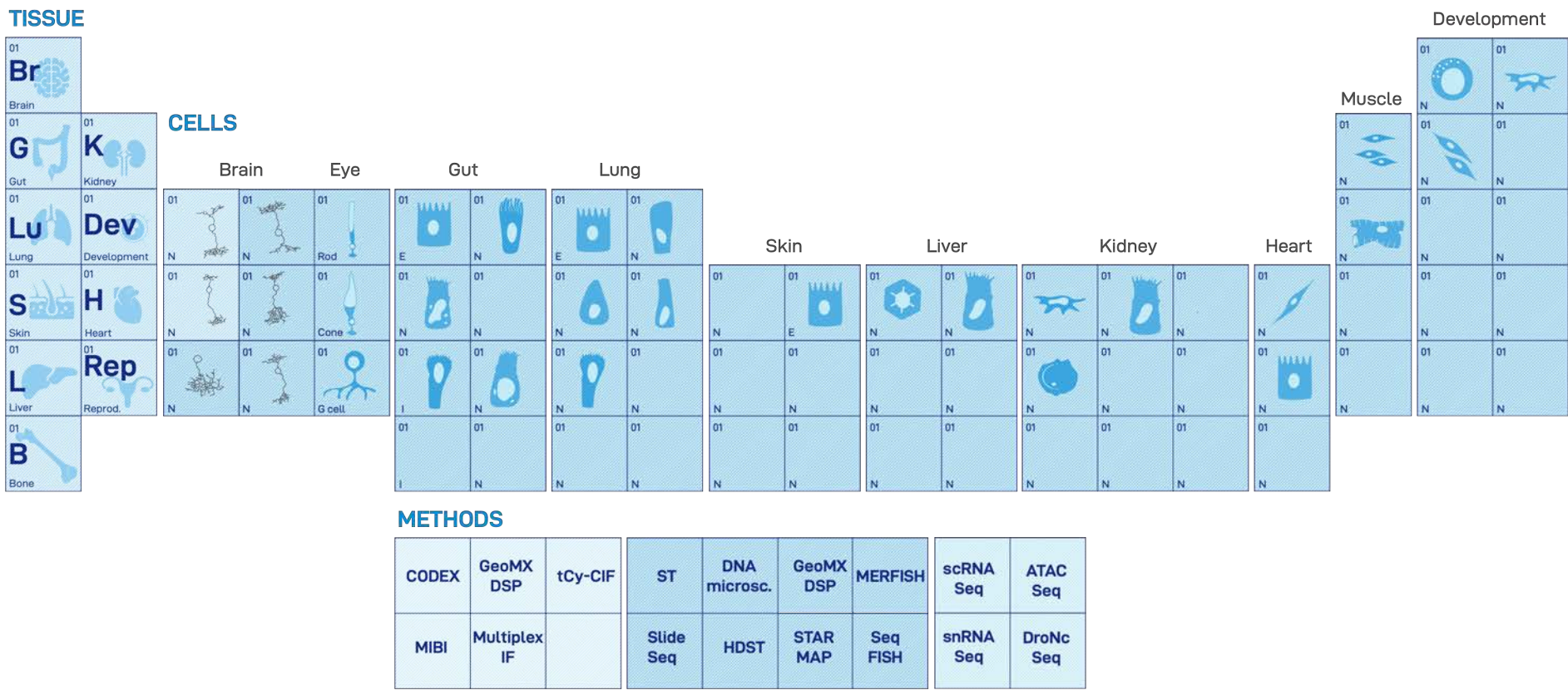
Mendeleev, 1869-1900

## Genomics



Human Genome Project, 1990-2003

## Biology and Medicine

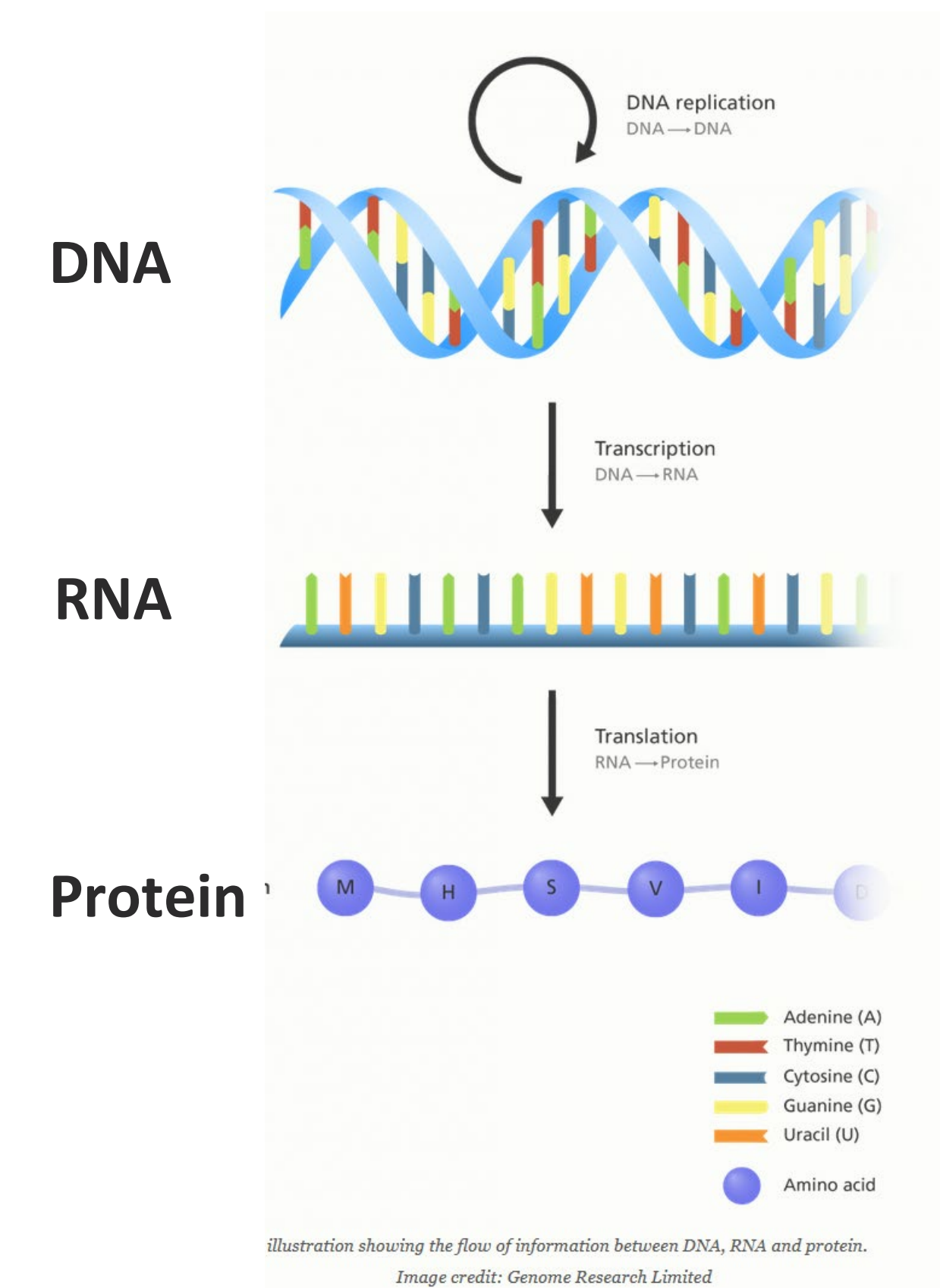




# What gives different cells different properties?

## They use different genes

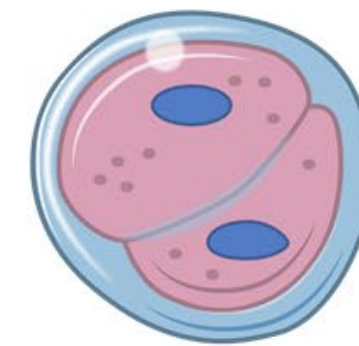
### Gene expression is a fingerprint of a cell



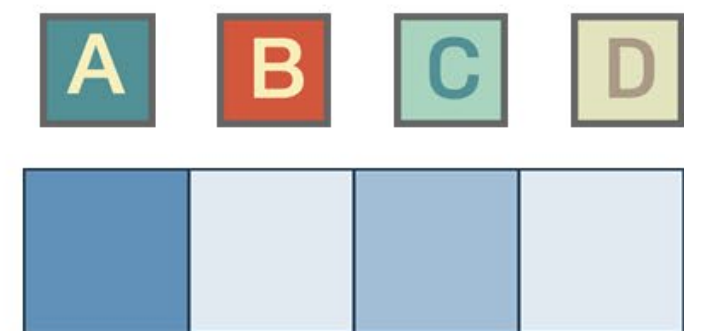
A single RNA molecule is called a transcript

The complete set of RNA molecules in a cell is a “transcriptome”

1

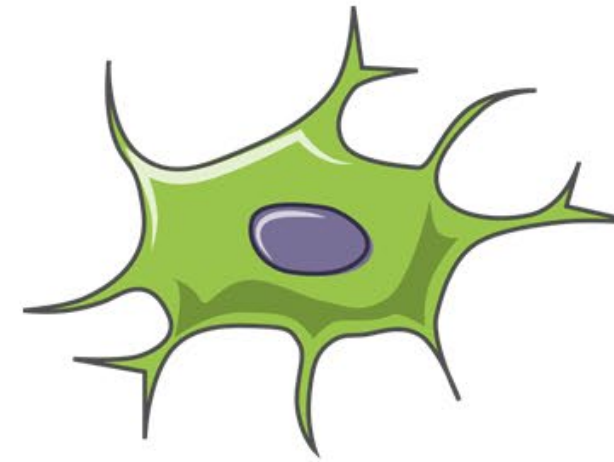


gene:

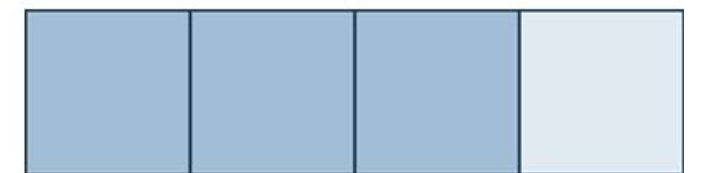


cell 1

2



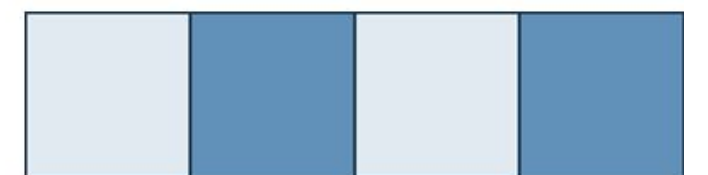
cell 2



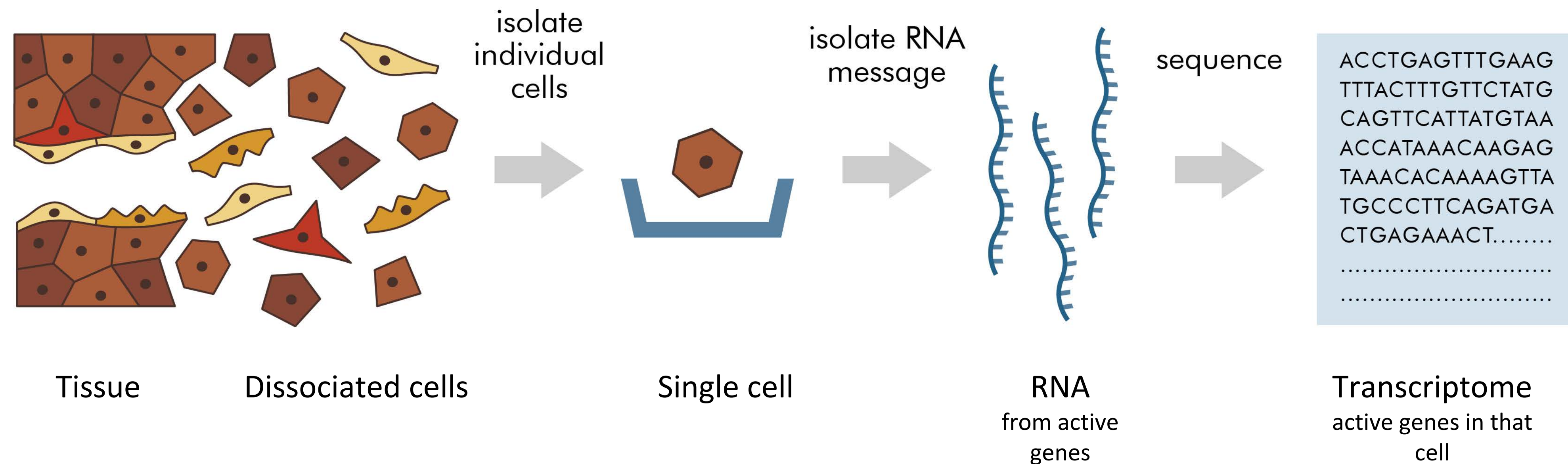
3



cell 3



# “Disruptive” technological advance: Single cell genomics



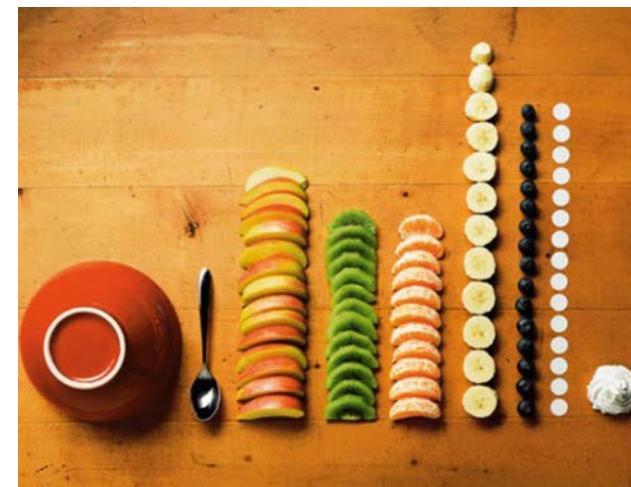
**“Single cell transcriptomics” methods can measure 4,000-12,000 genes per cell, across tens of thousands of cells, in any tissue in any species**



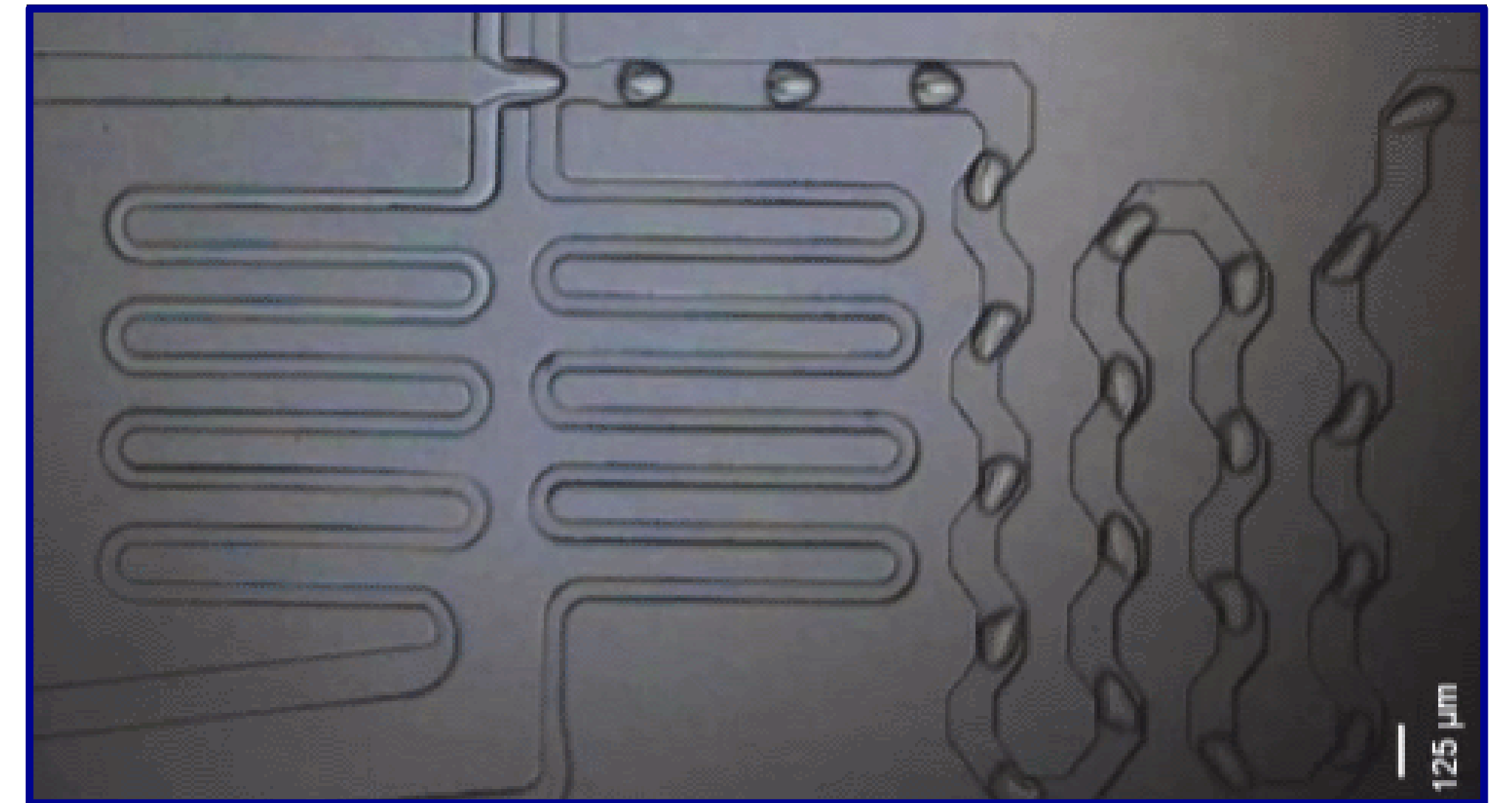
# Massively parallel methods let us profile many cells



Bulk genomics



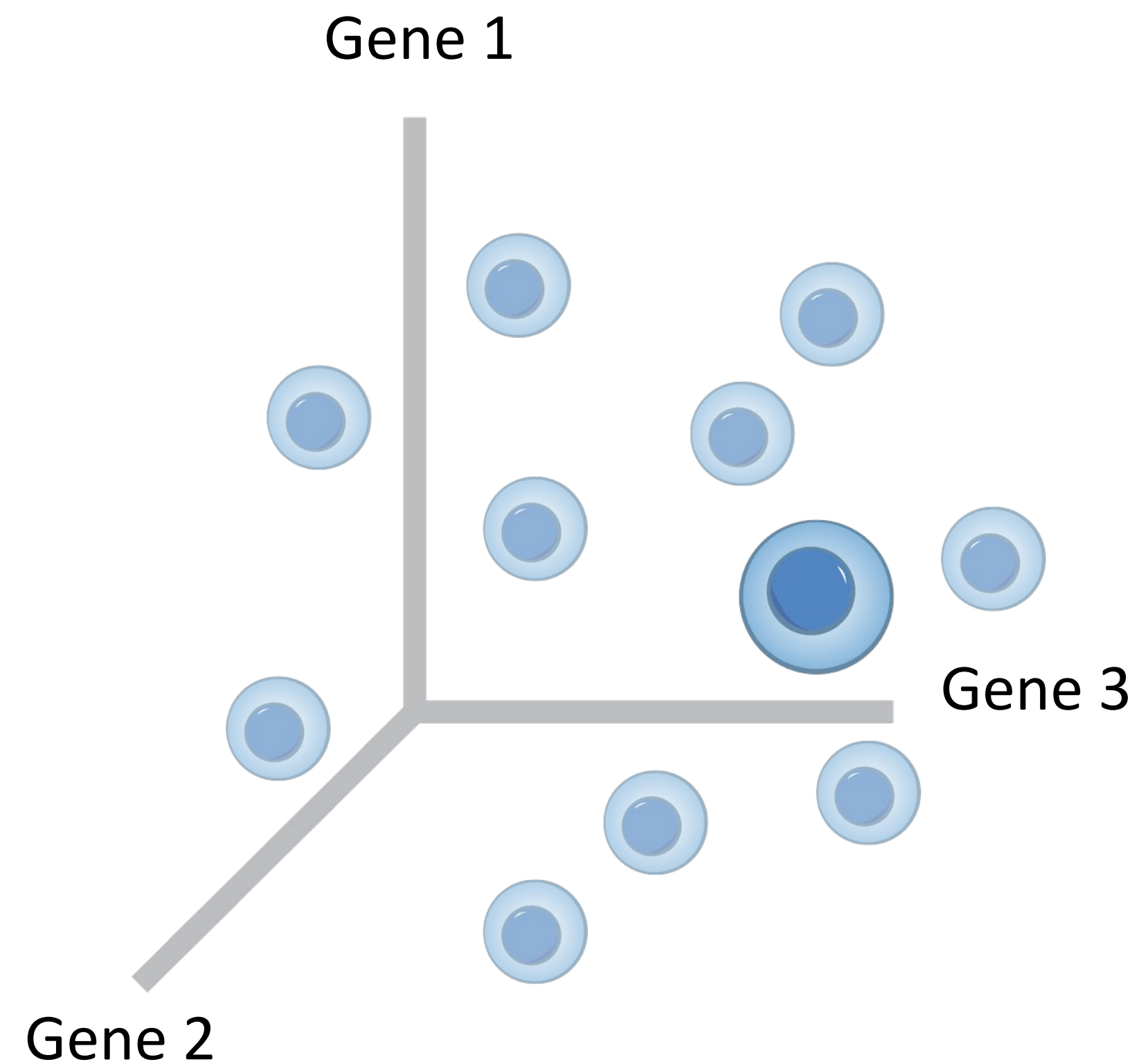
Single cell genomics



5,000 cells/second

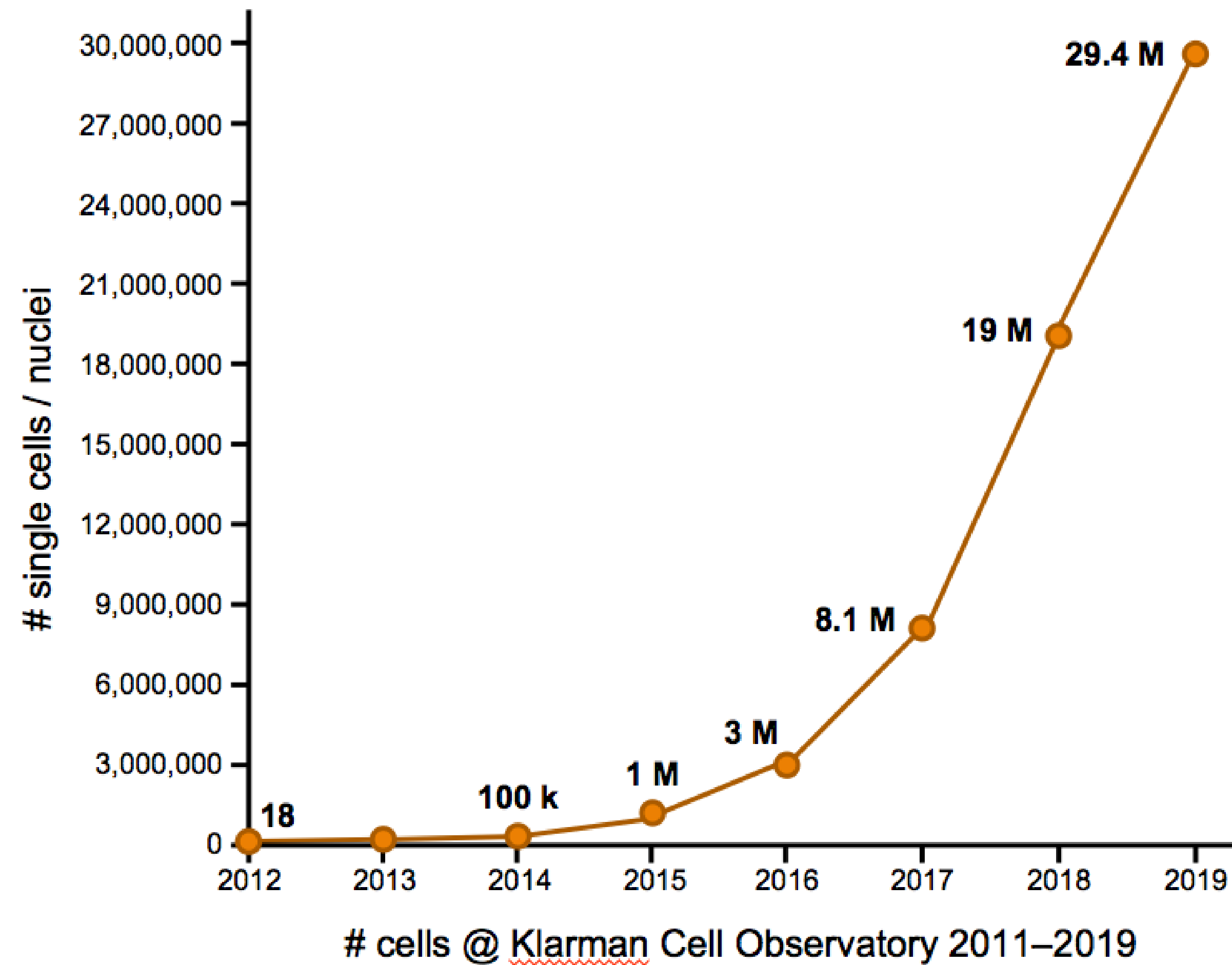
# Gene expression provide the coordinates for a map of all human cells

**Idea:** There are ~20,000 genes in the genome.  
We can define a cell as a point in 20,000- dimensional gene expression space



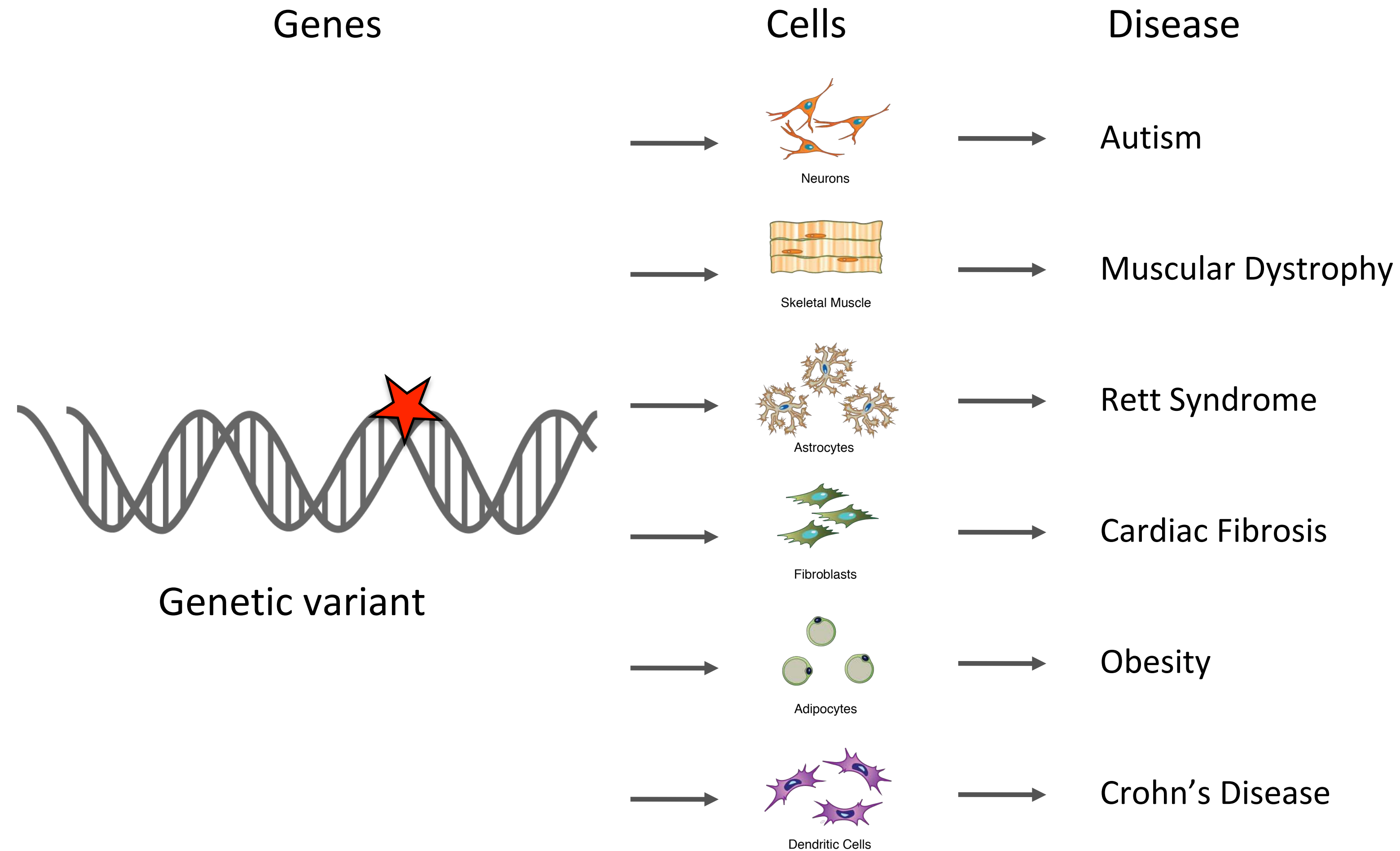


Single cell genomics is growing exponentially, and provides the necessary scale to approach an atlas of human cells



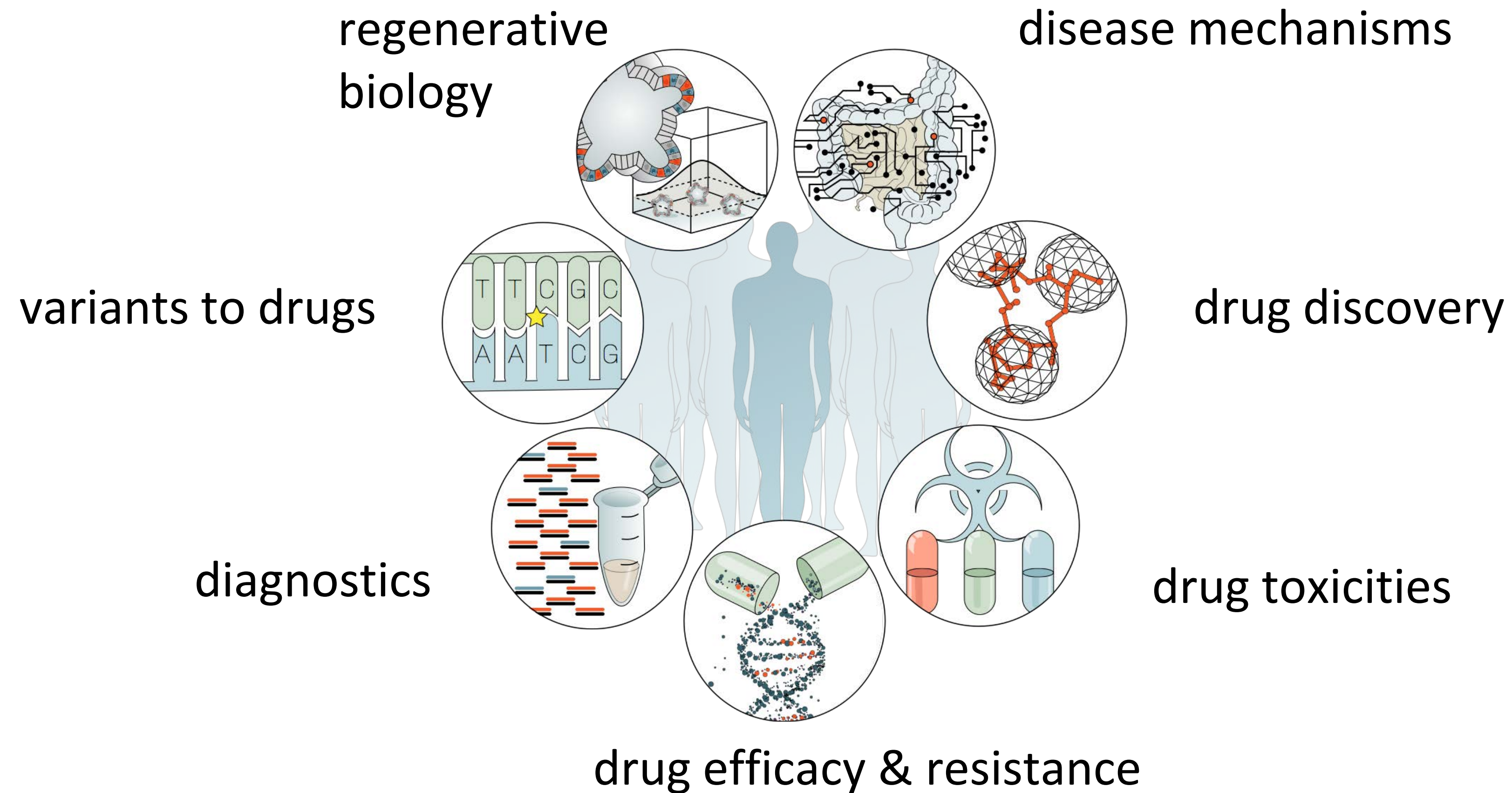
# Why a human cell atlas?

Knowing our cells is essential to understand the genes that cause disease





# Knowing our cells is essential to treating disease

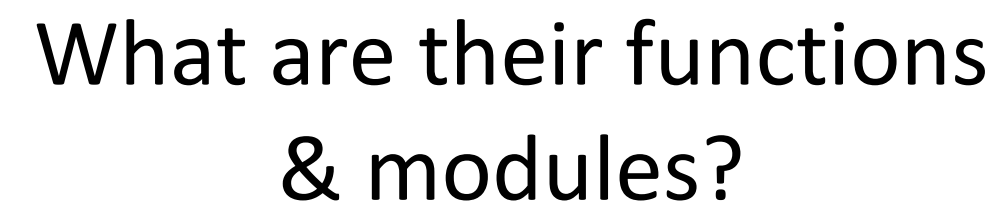
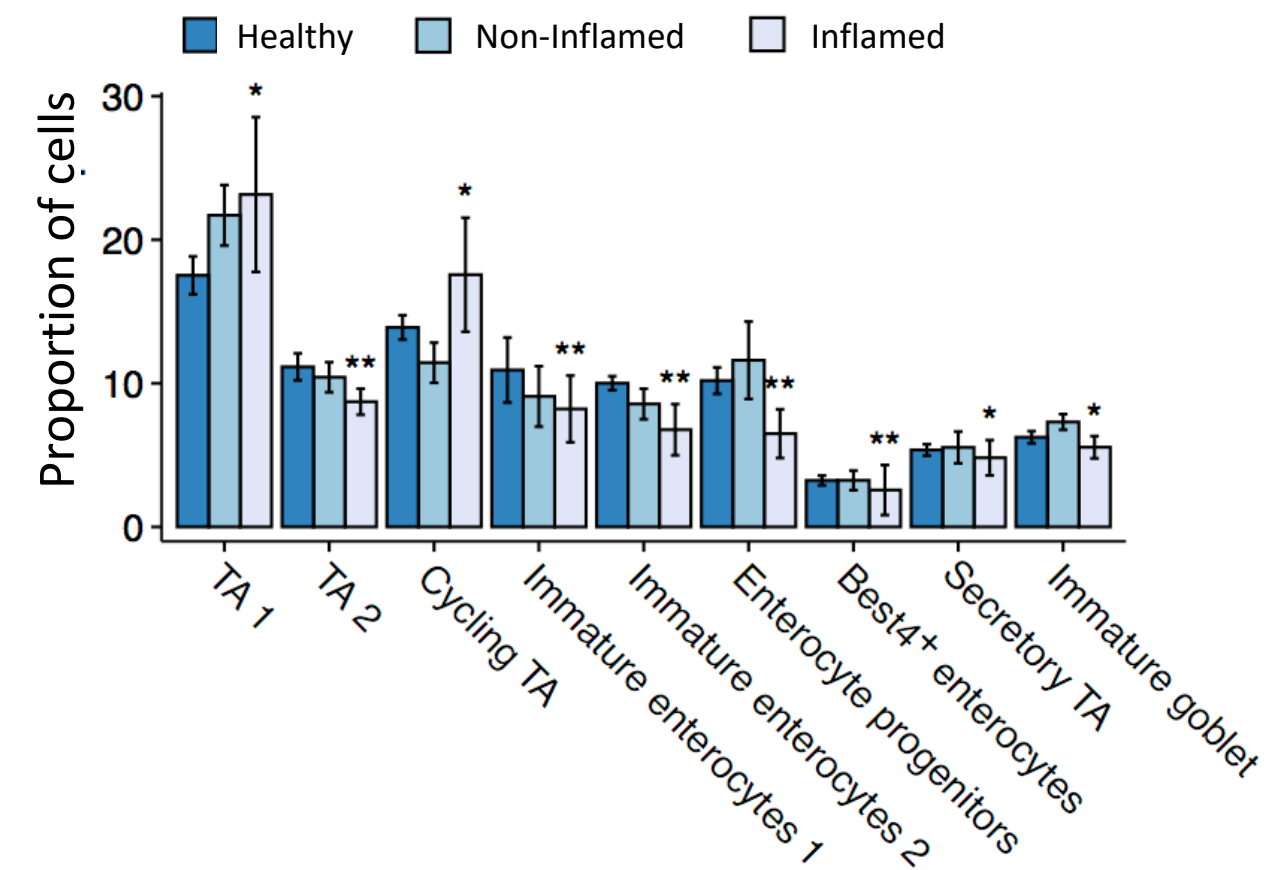






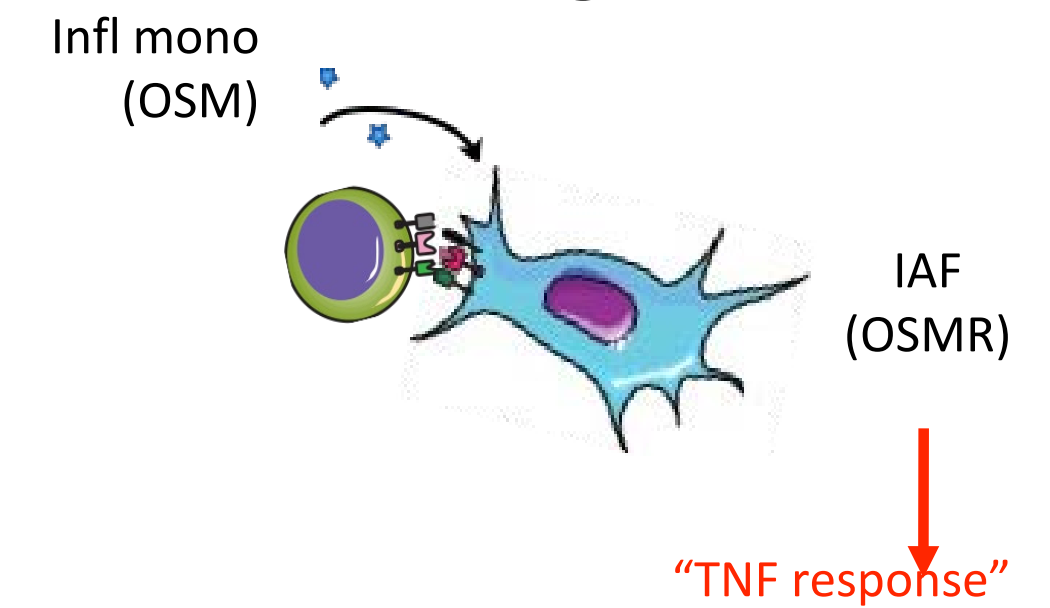
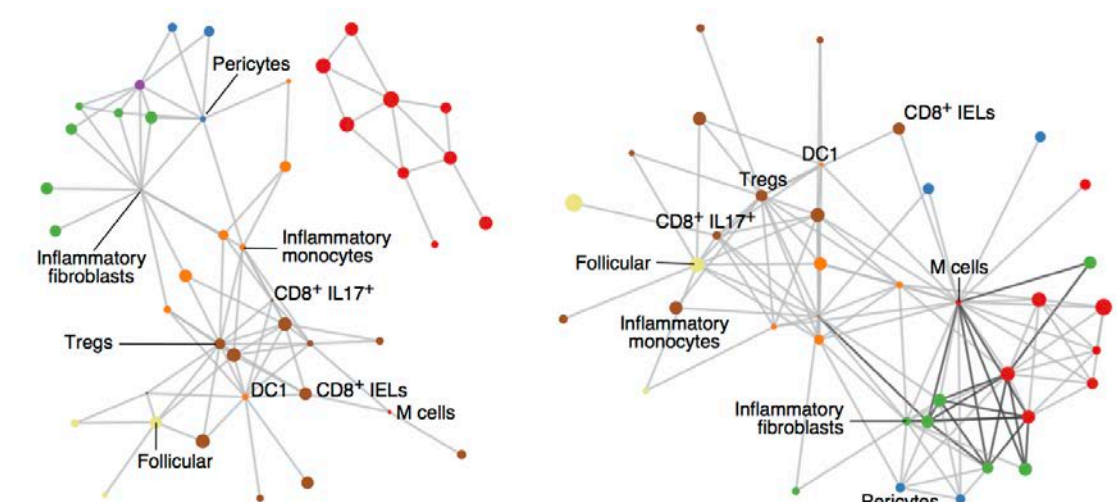
## Which cells are disrupted?

# Which cell programs are changed?



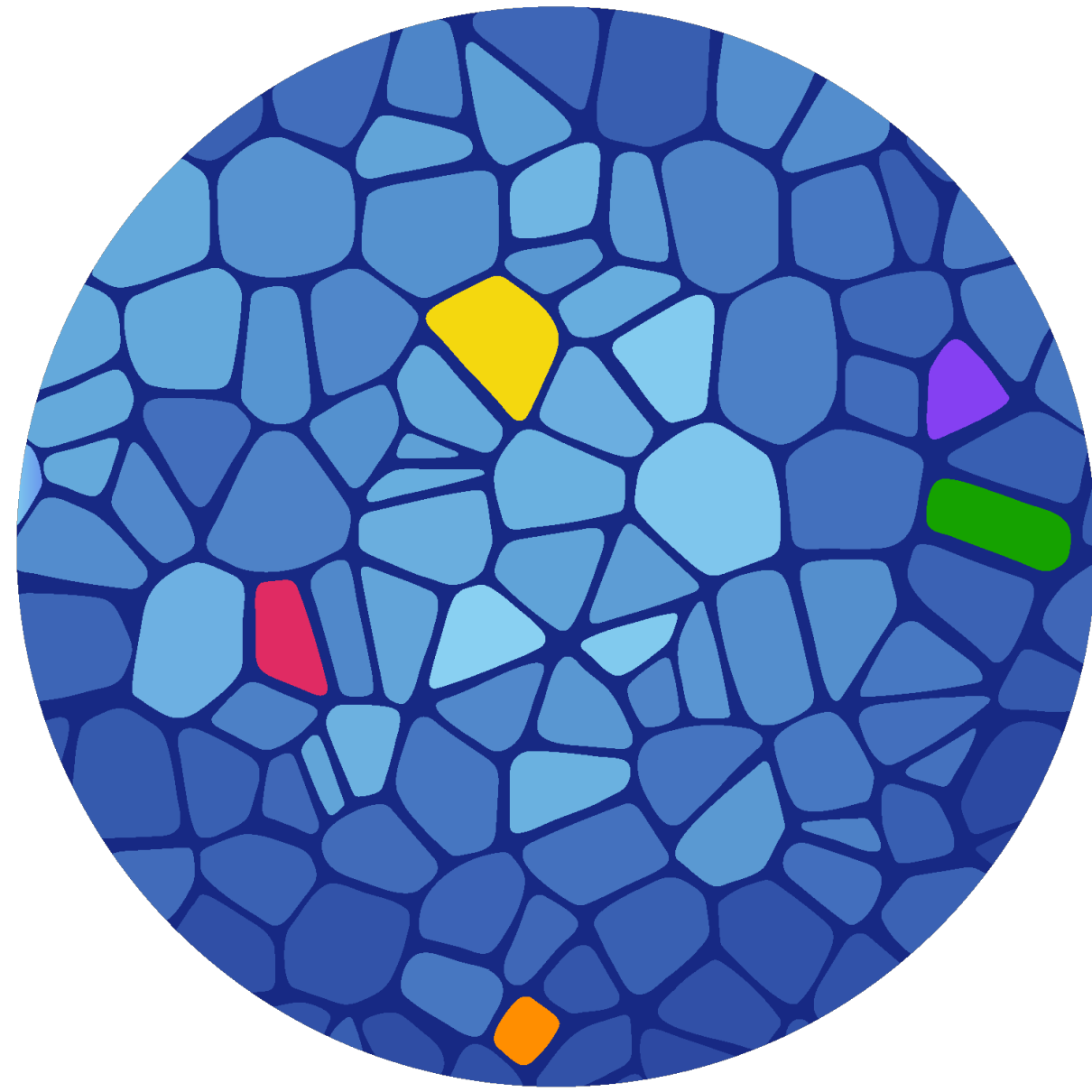
# Which communications are disrupted?

# What is the effect of drug?



Smilie et al. (2019) Cell



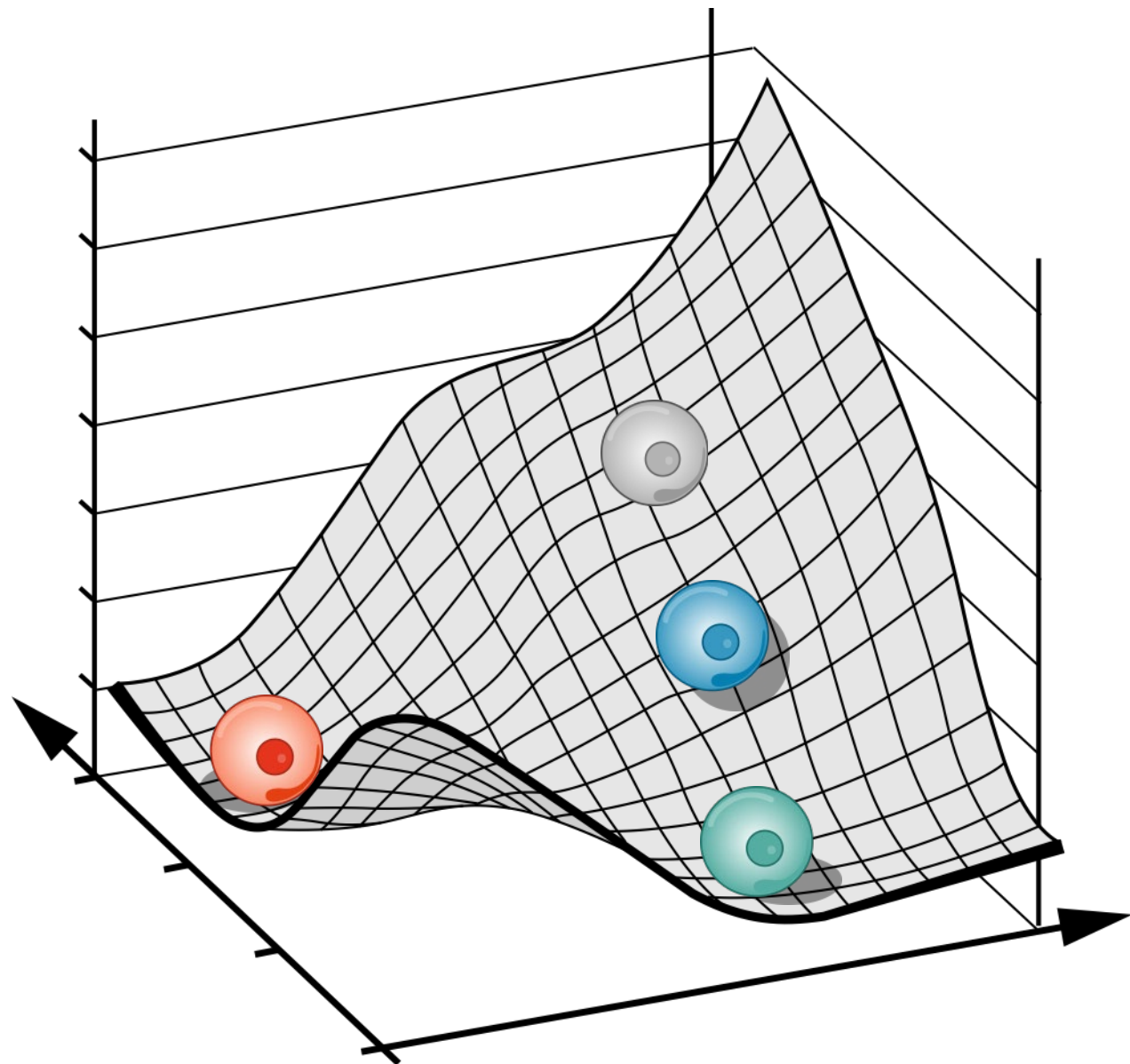


## What is the Human Cell Atlas?

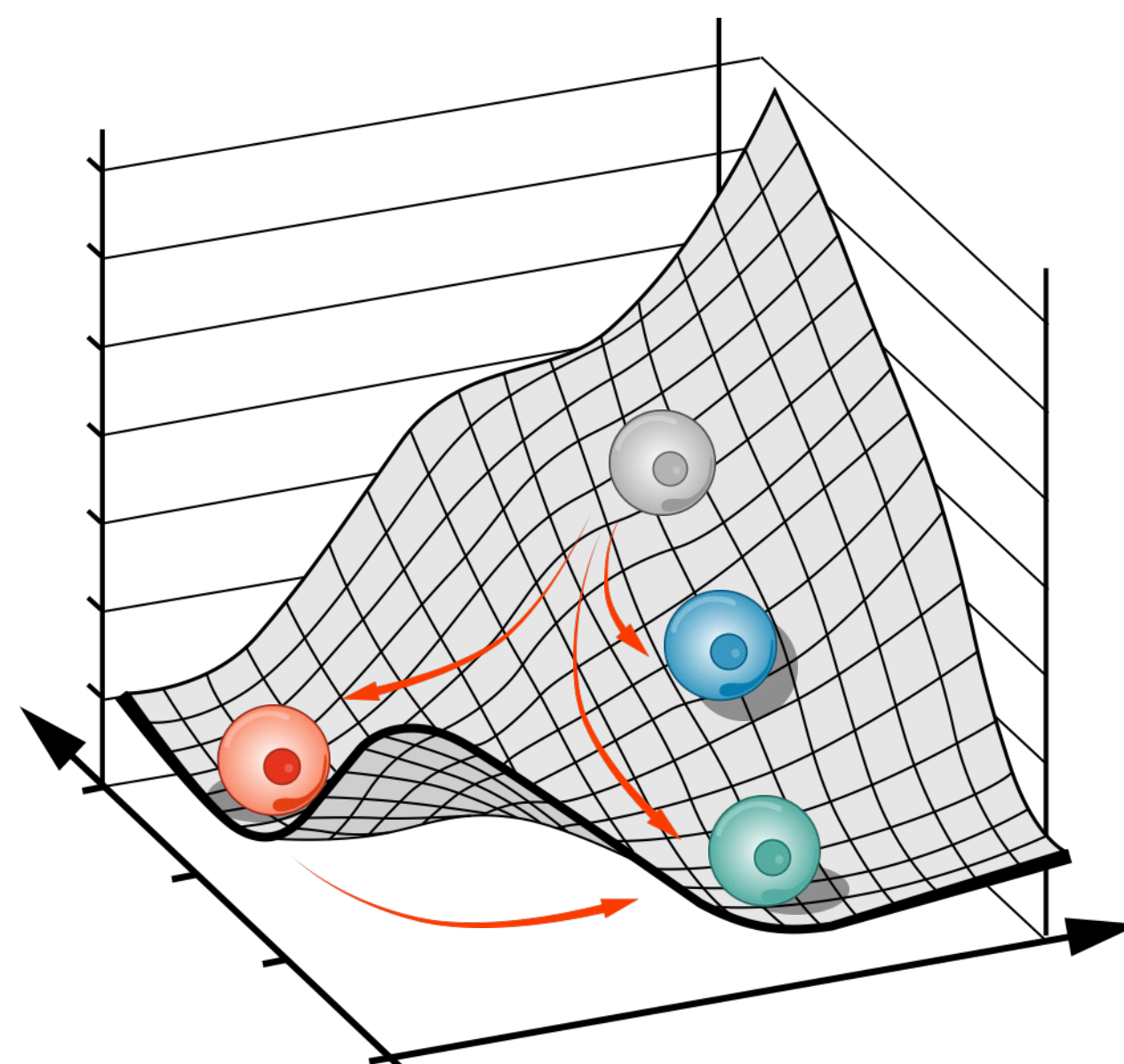
- The HCA Roadmap: concept, practice, and use
- Atlas infrastructure: Tools, data platform

# Concepts

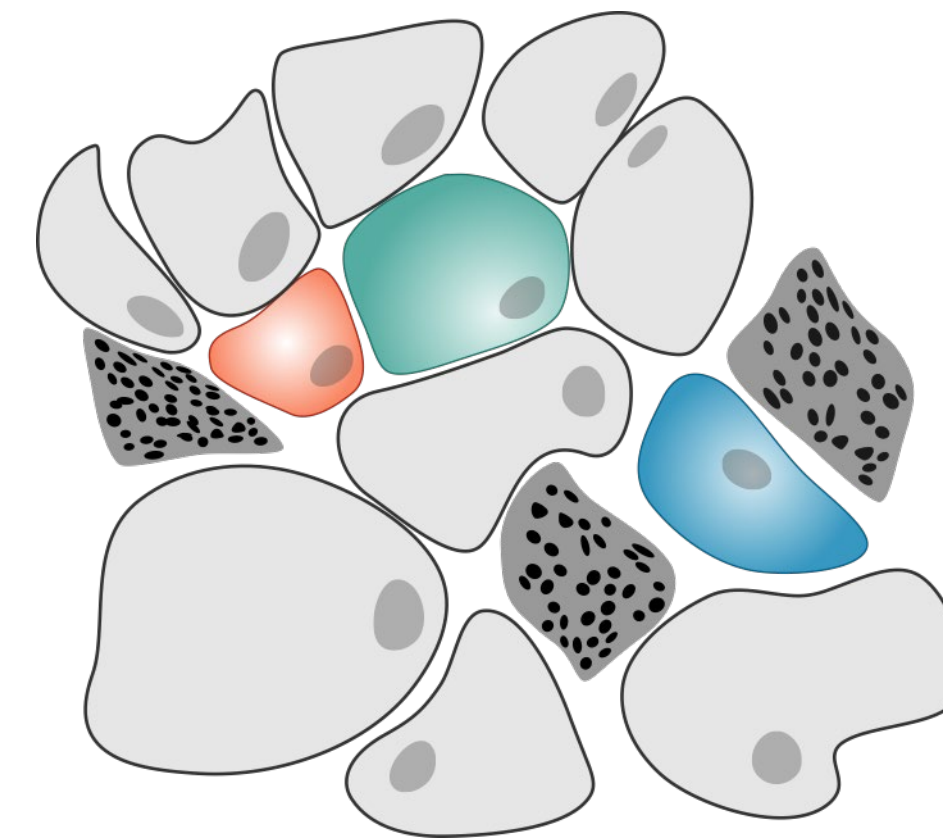
types / states



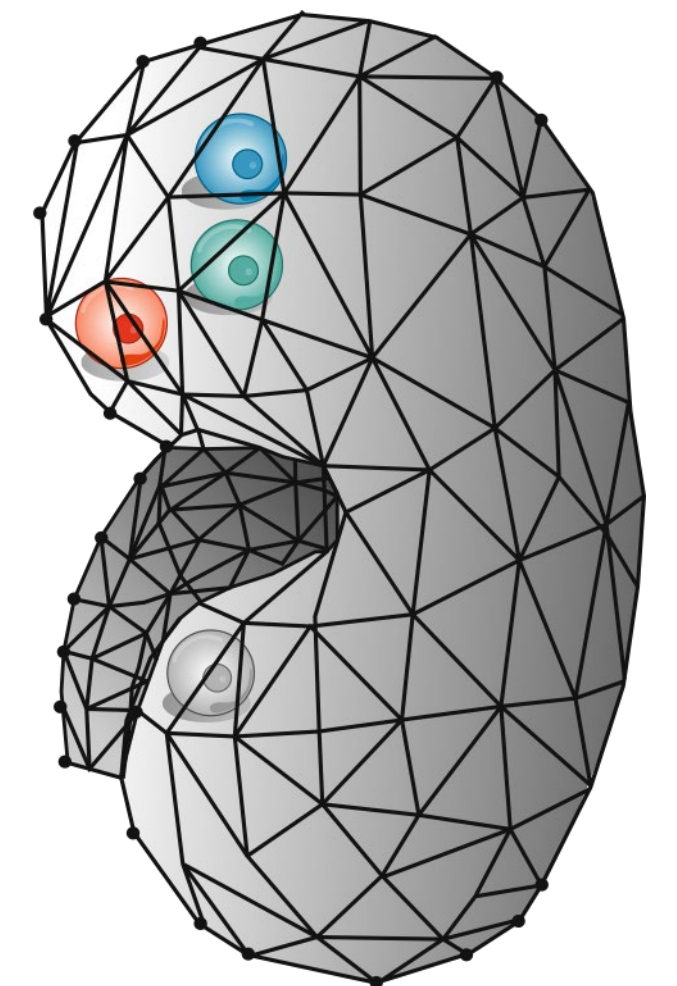
trajectories/transitions



histological modules



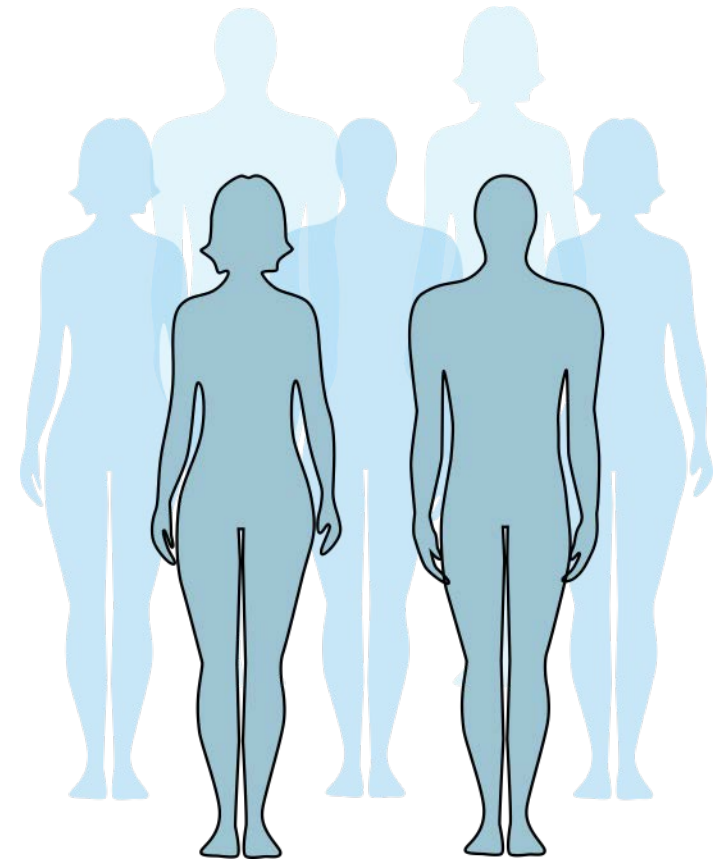
positions



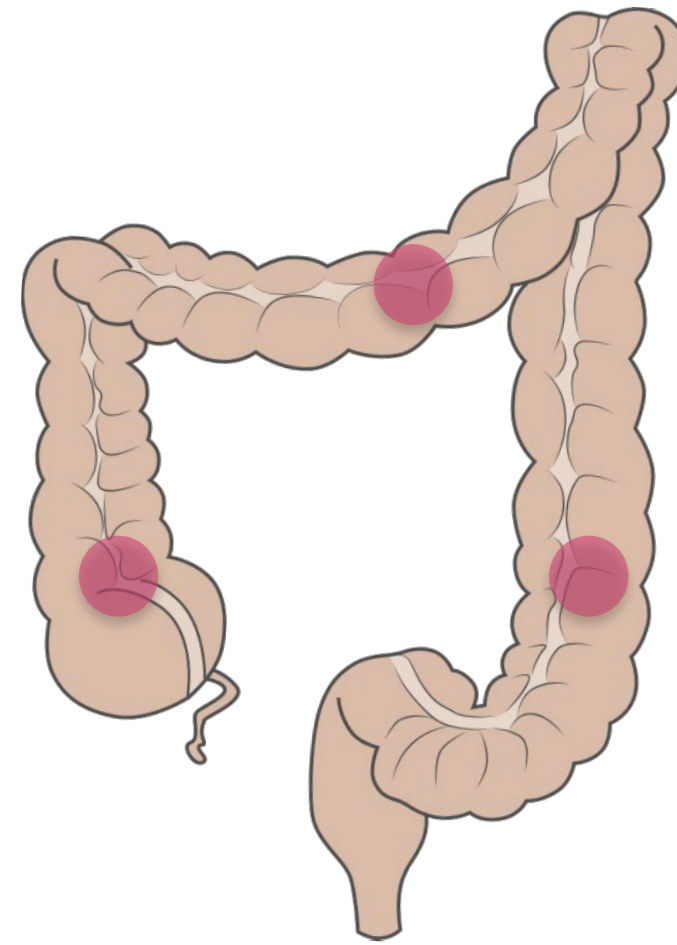


# Tissue sampling

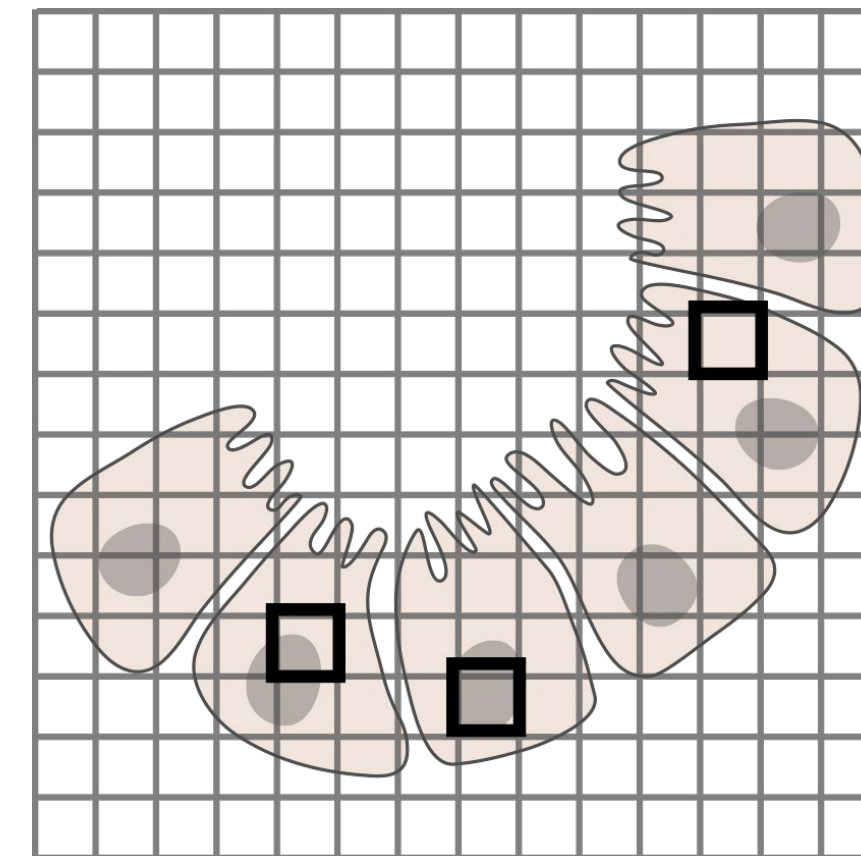
Number of individuals



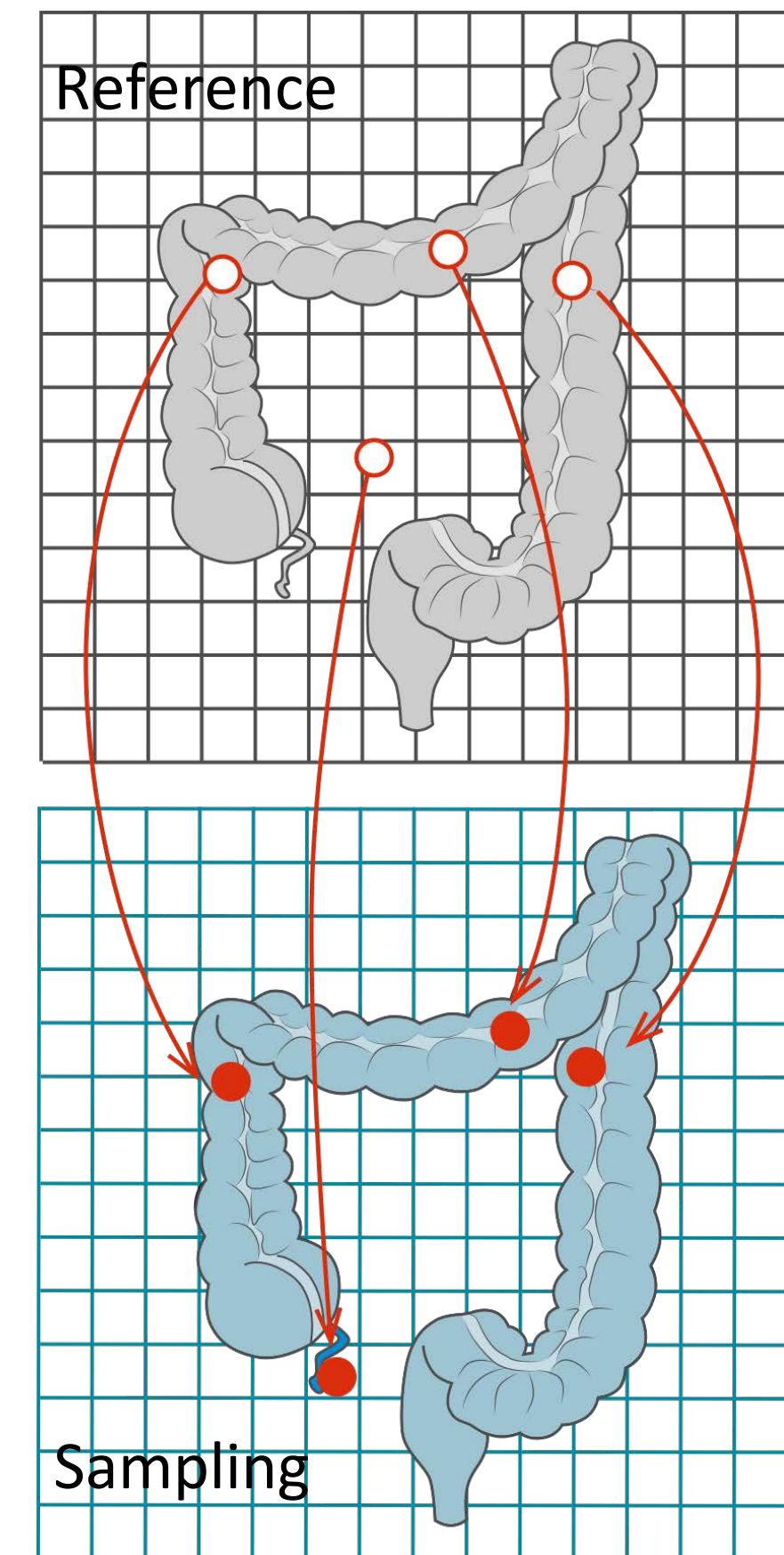
Anatomical sampling



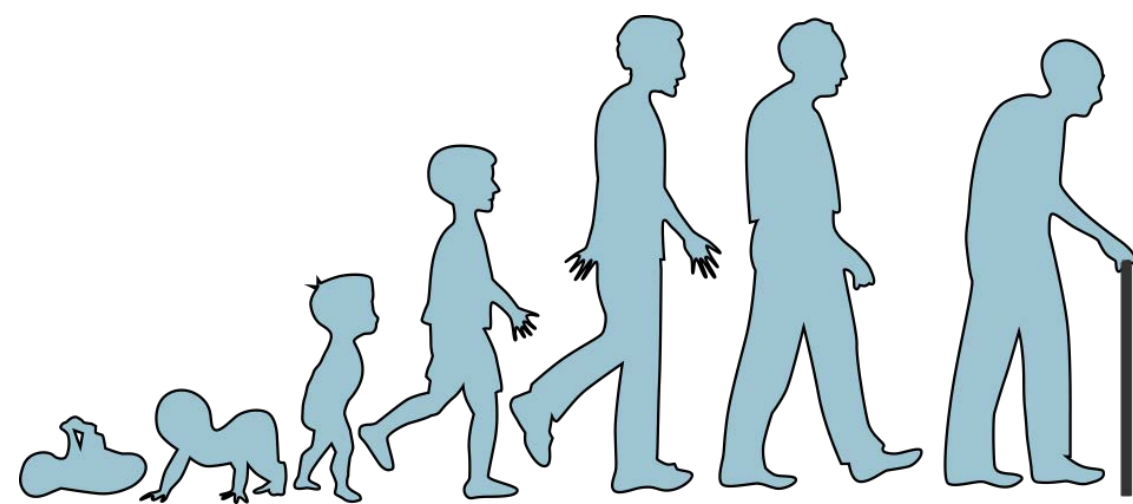
Number of regions



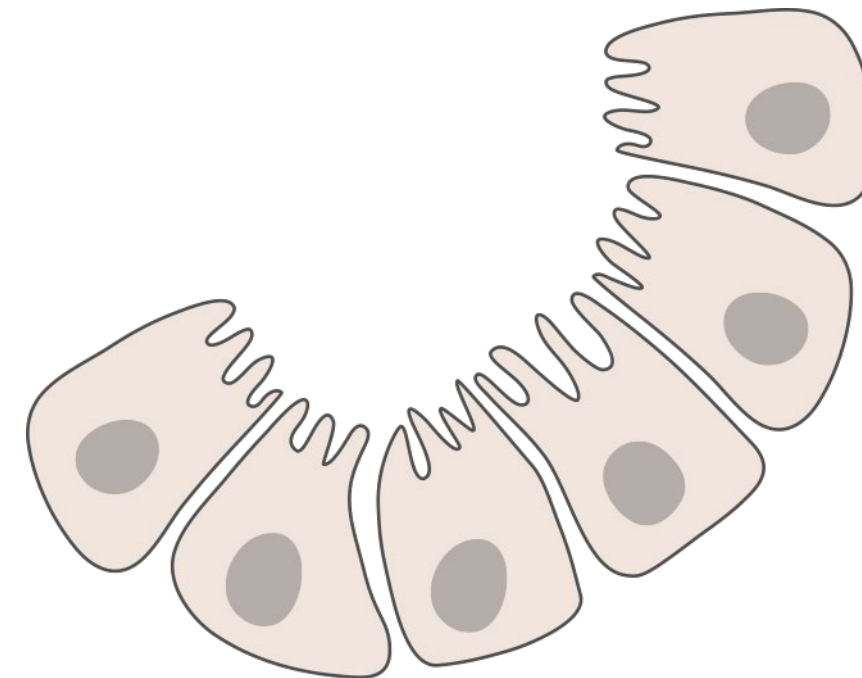
Coordinate framework



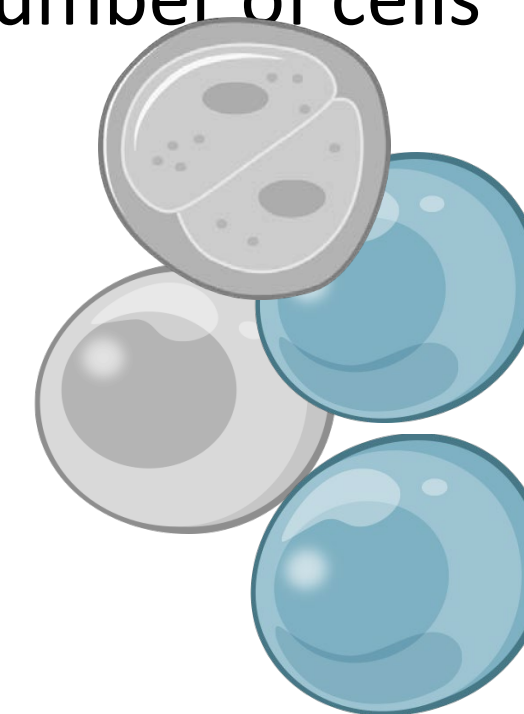
Development and aging



Histological sampling



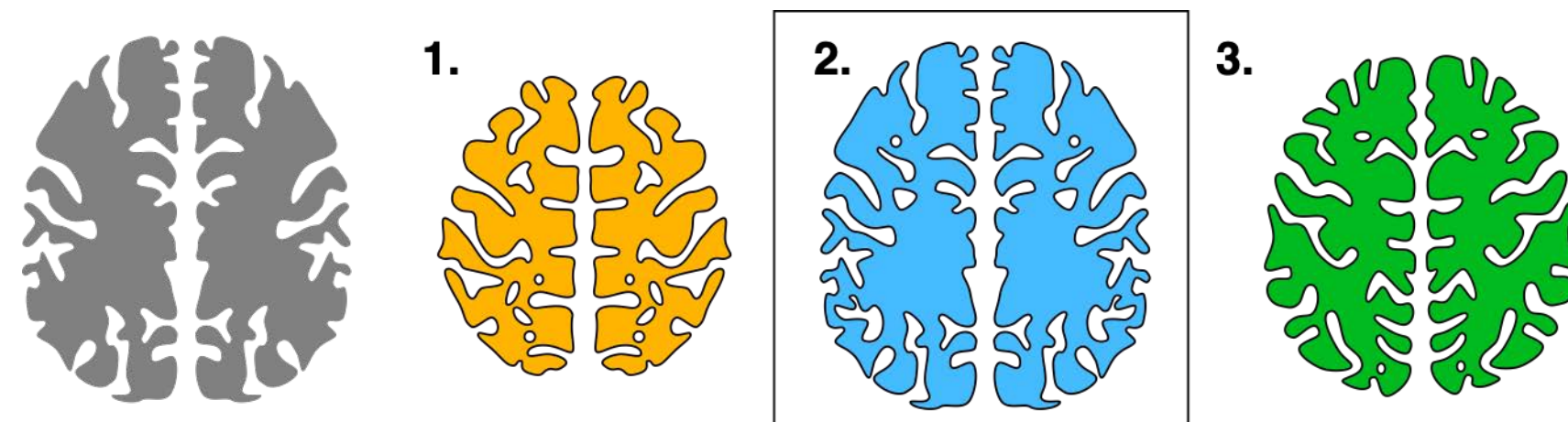
Number of cells



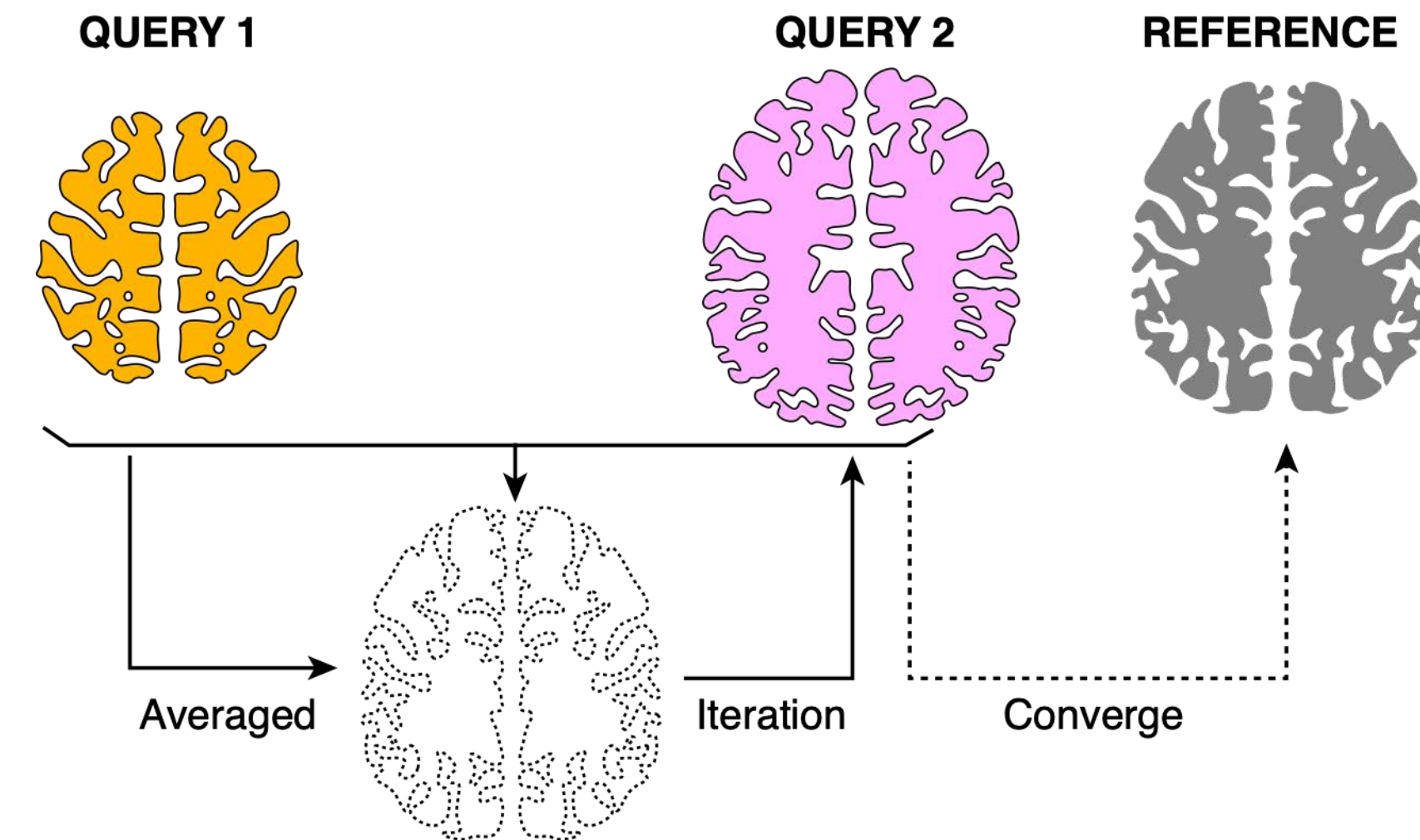


# Common Coordinate Frameworks (CCFs) to map and aggregate data

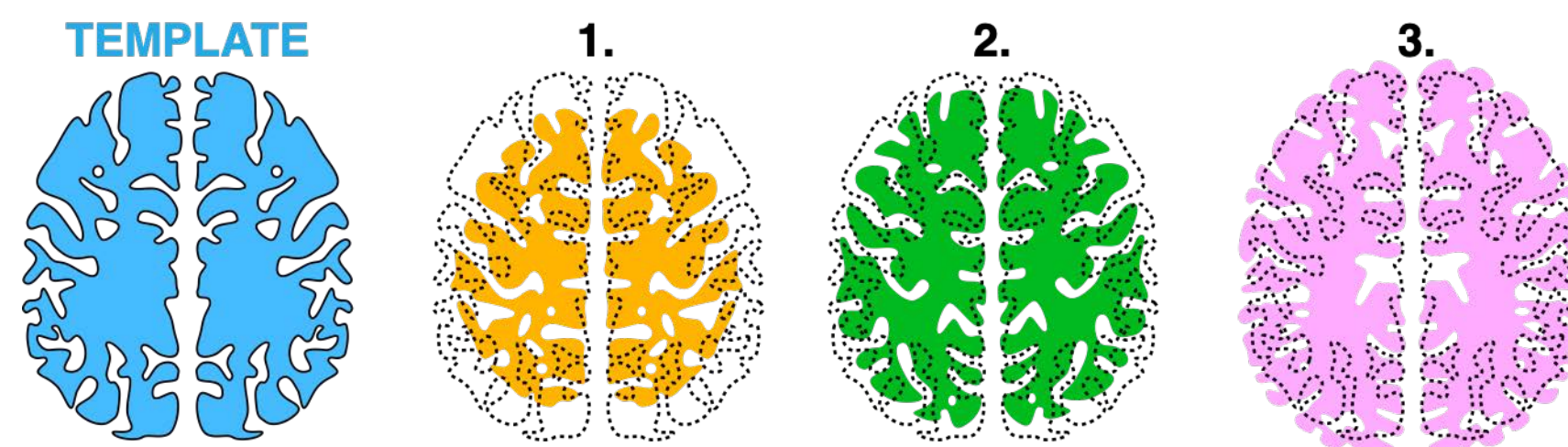
1. Calculate best template



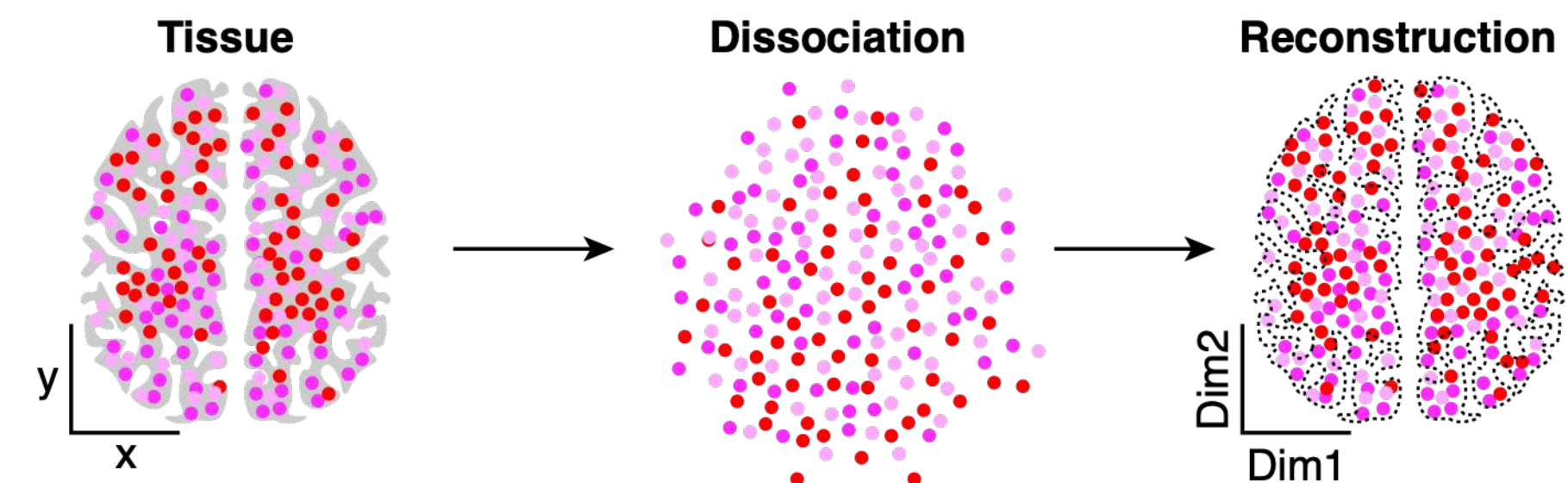
3. Approach 2: Iteratively align and average



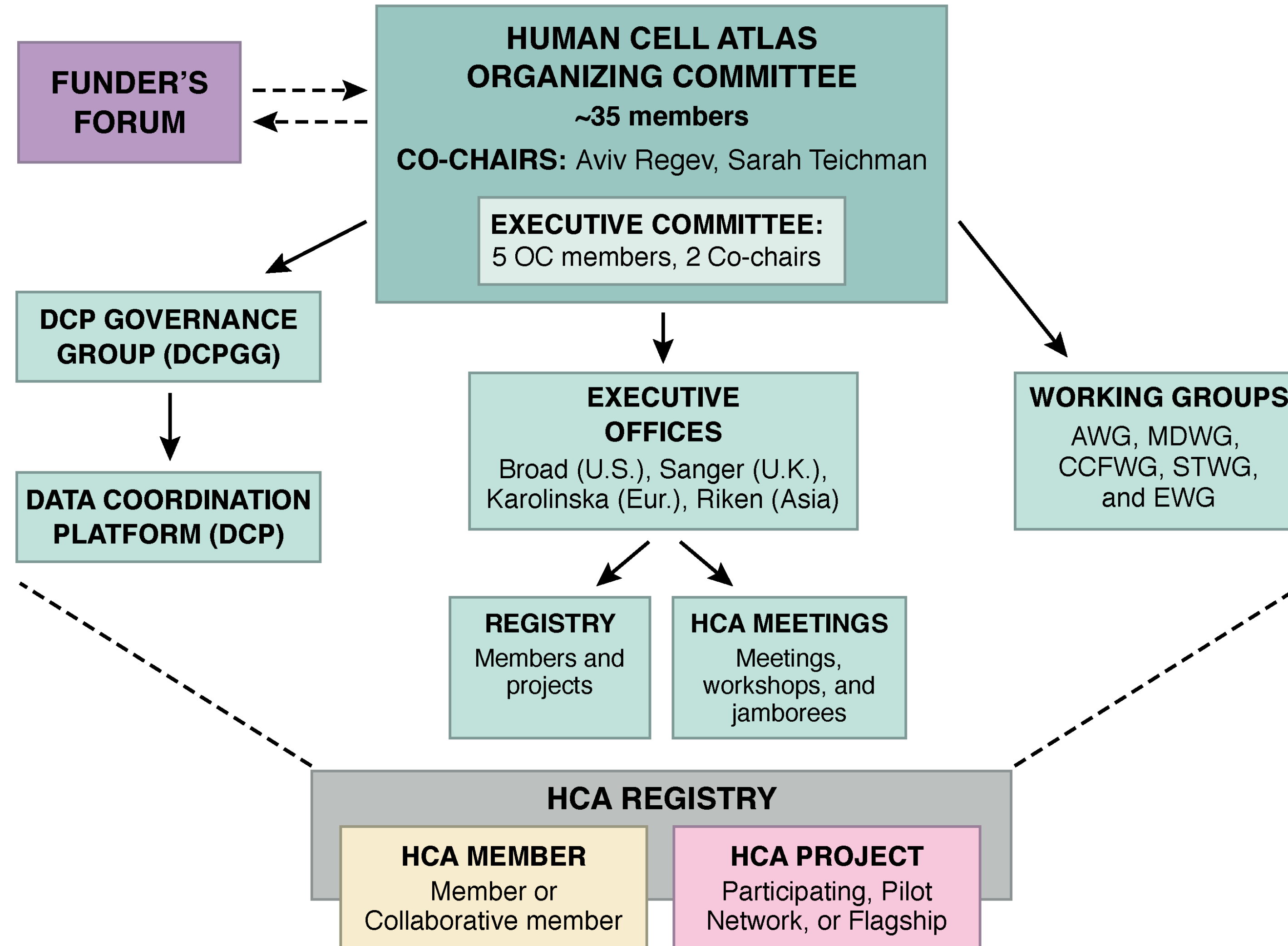
2. Approach 1: Map all to one template



4. Reconstructing an atlas from its features

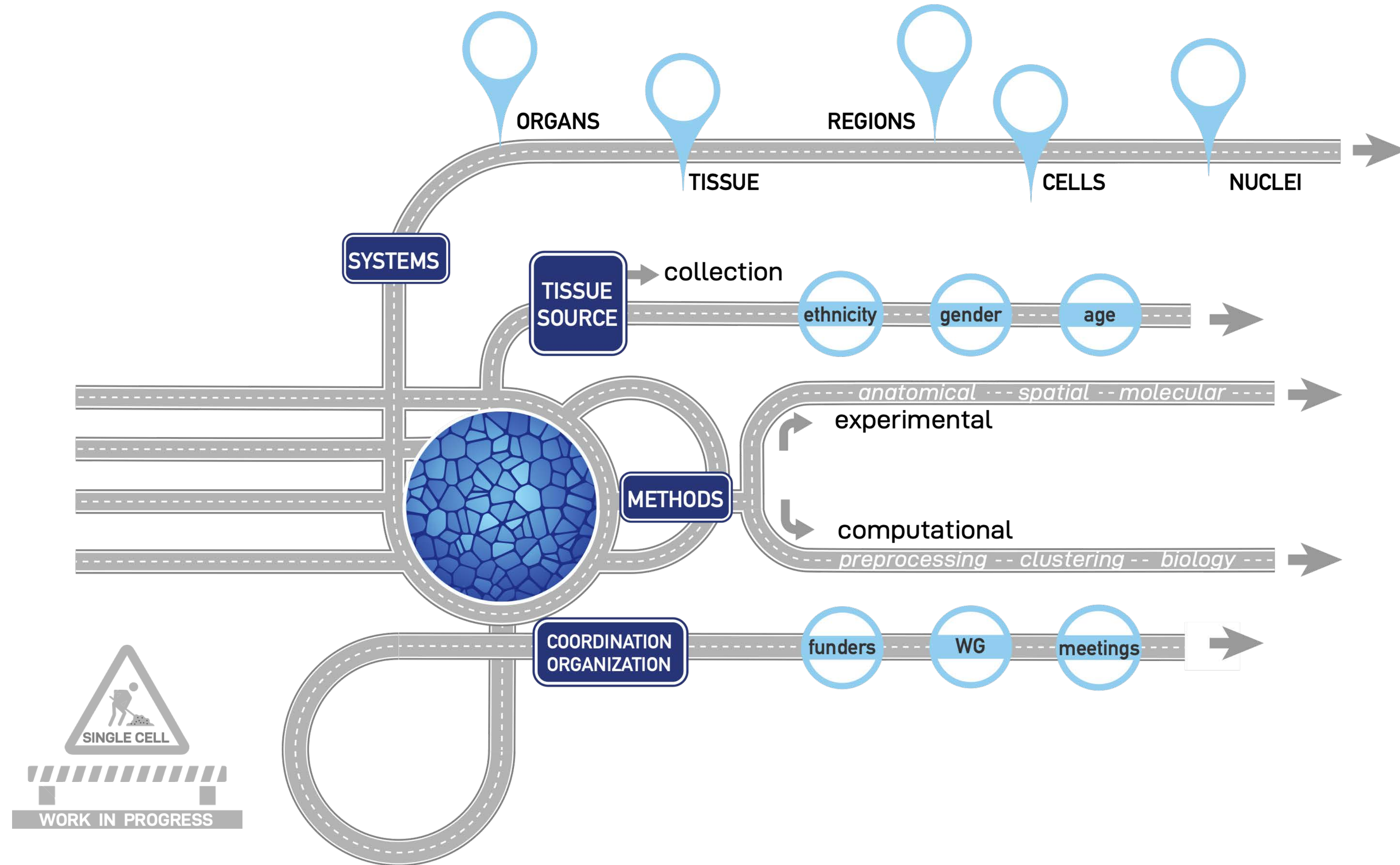


# The HCA Consortium





# The HCA Roadmap



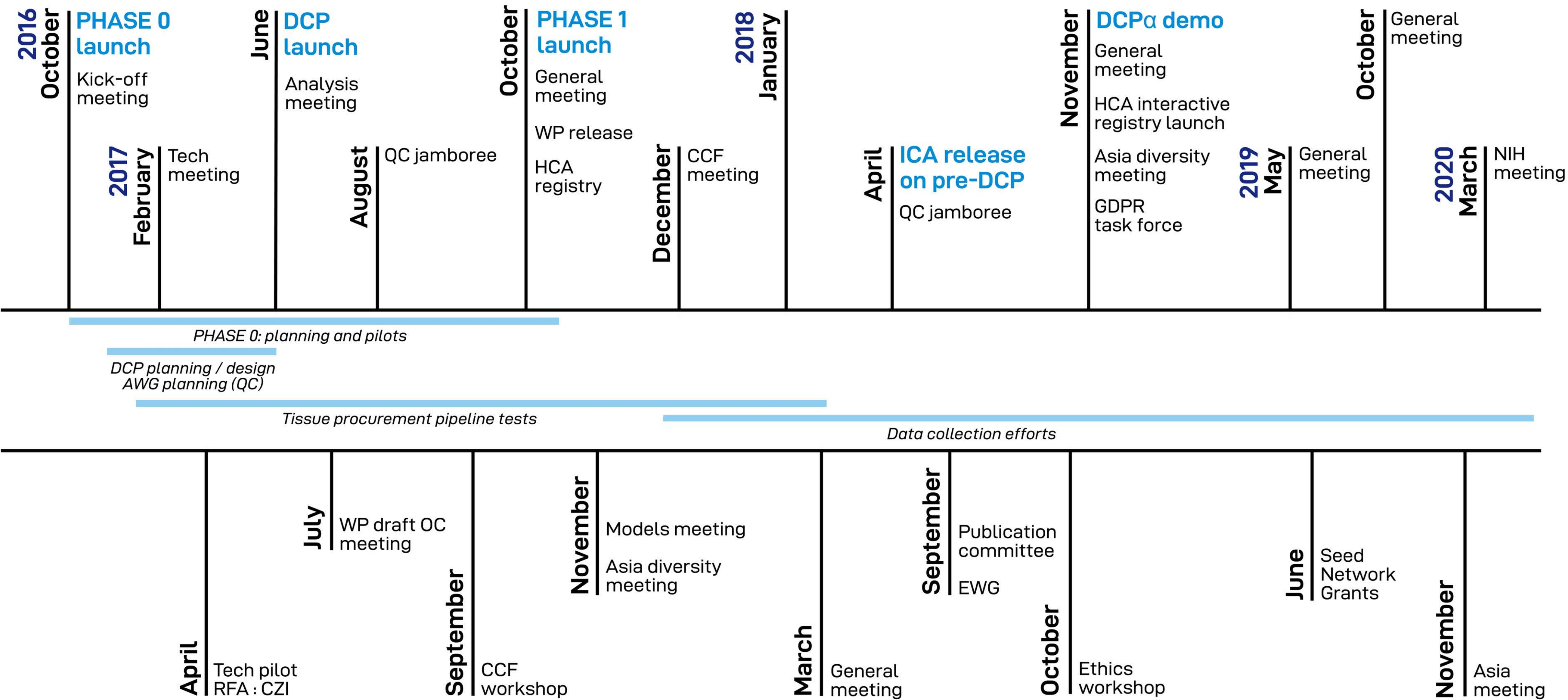
# HCA principles and values



- Quality
- Flexibility
- Community
- Equity: Diversity and inclusion: subjects and scientists
- Transparency and open sharing: data, code, protocols
- Privacy and ethics
- Technological innovation and excellence
- Computational innovation and excellence

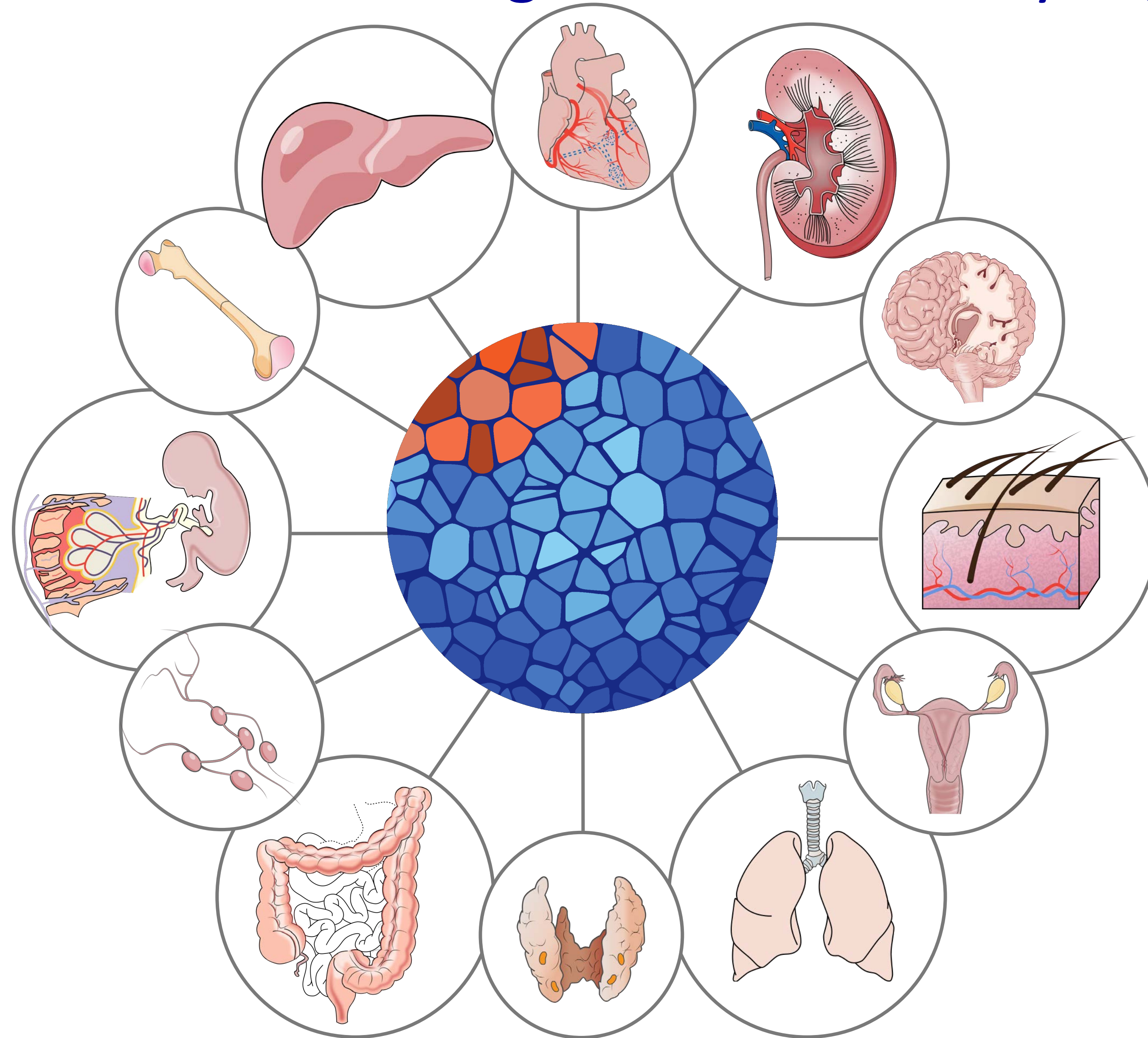


# HCA Timeline: 2016 - present





Atlases are now being made from many organs

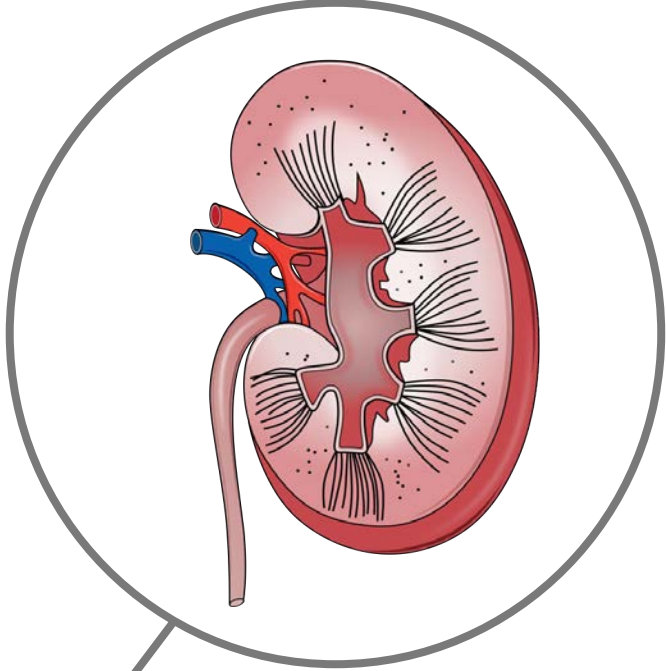
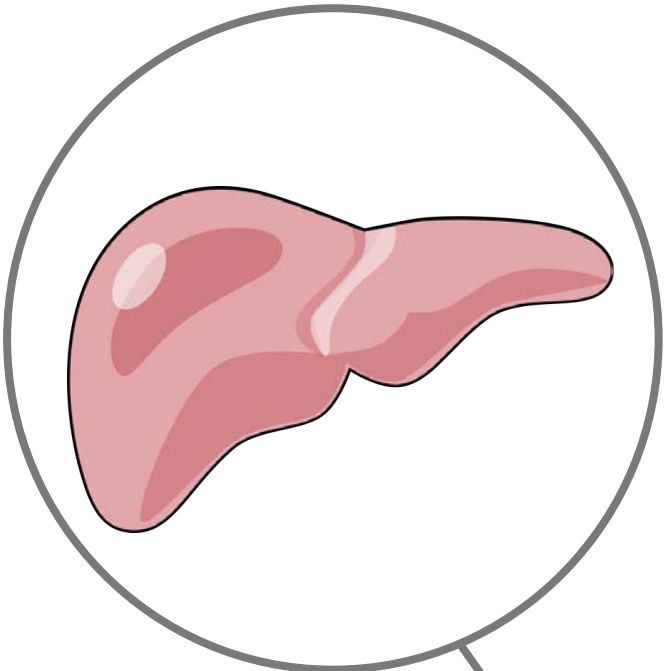




# Views from the Atlases



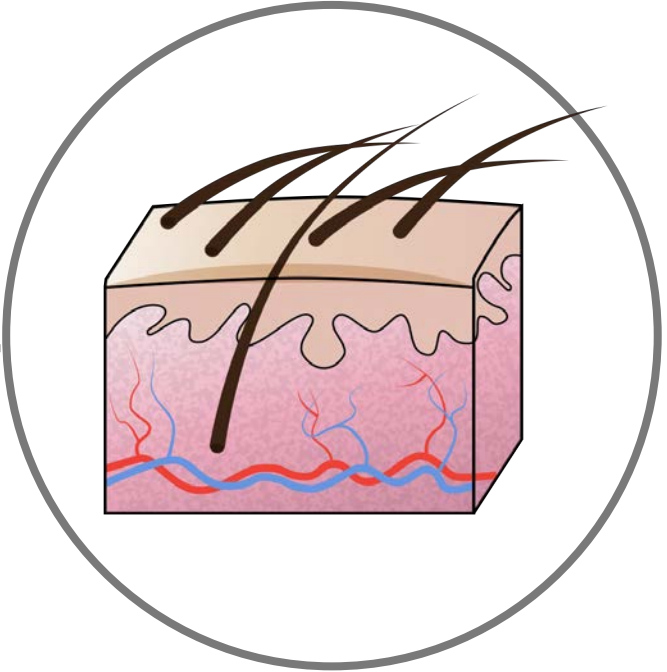
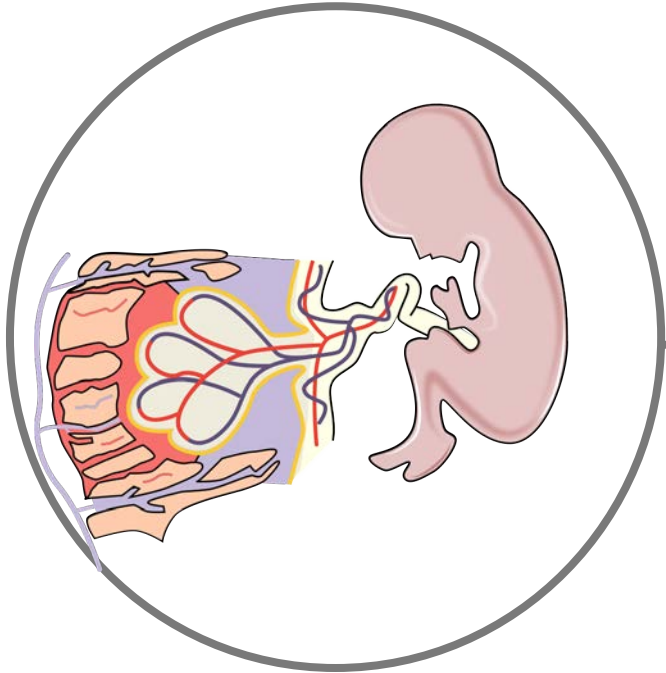
INDIVIDUALS	43
SAMPLES	50
CELLS	111,628




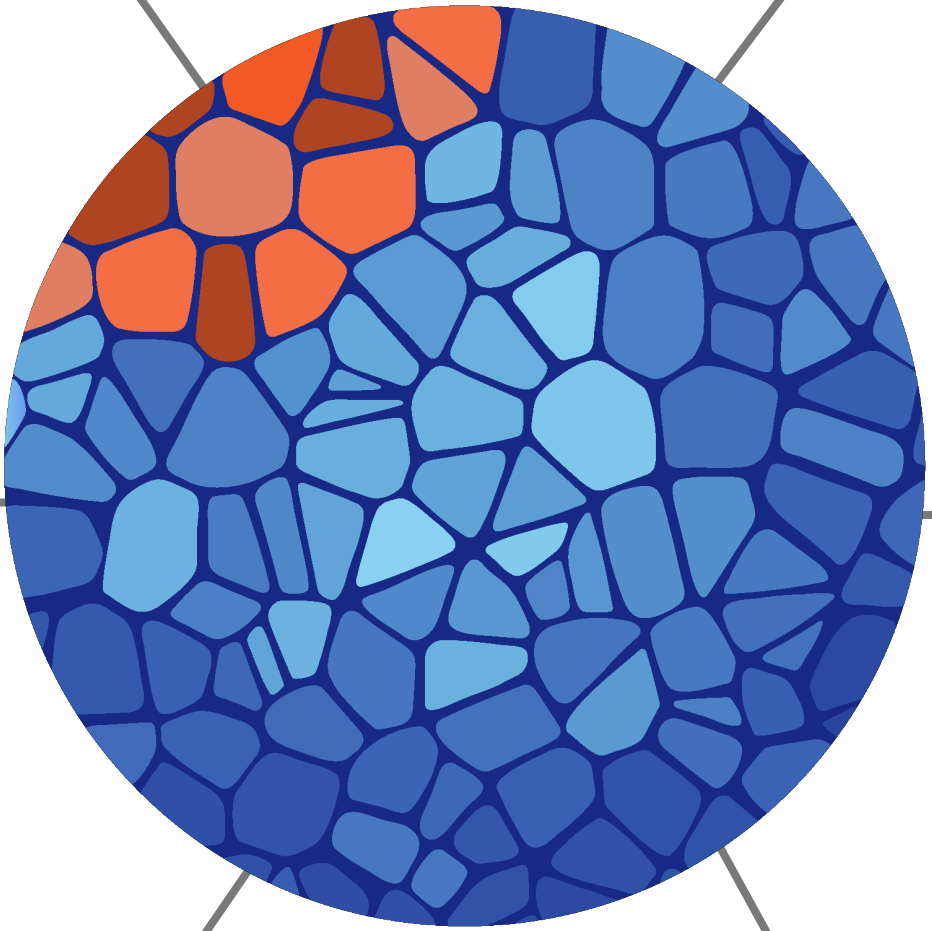
INDIVIDUALS	62
SAMPLES	144
CELLS	759,978



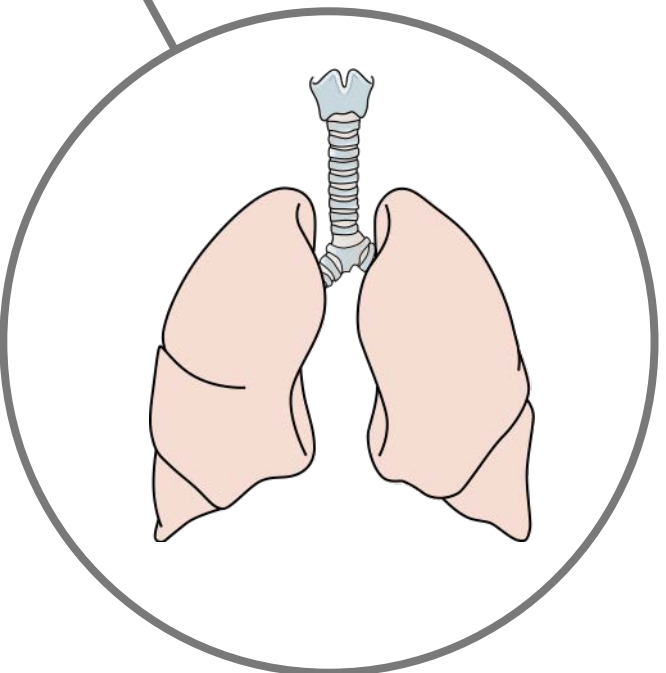
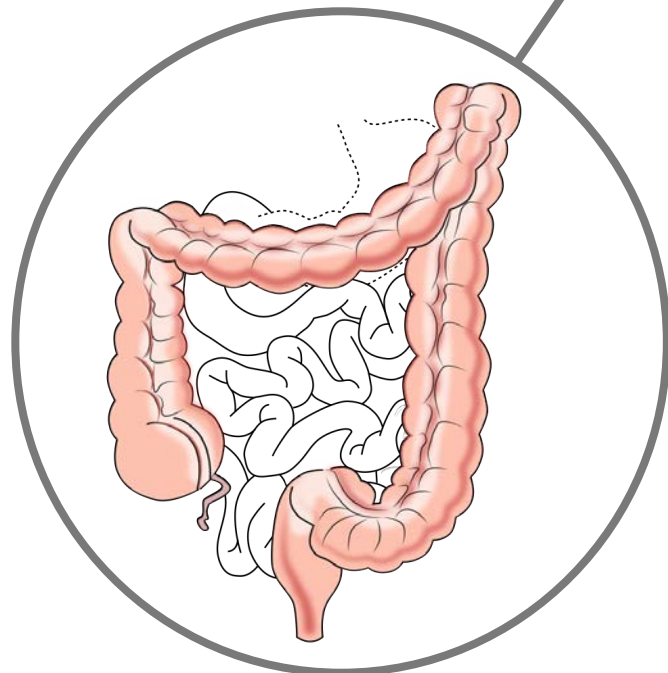
INDIVIDUALS	70
SAMPLES	697
CELLS	4,243,865



INDIVIDUALS	34
CELLS	914,590



INDIVIDUALS	131
SAMPLES	216
CELLS	1,347,864



INDIVIDUALS	160
SAMPLES	308
CELLS	1,148,245

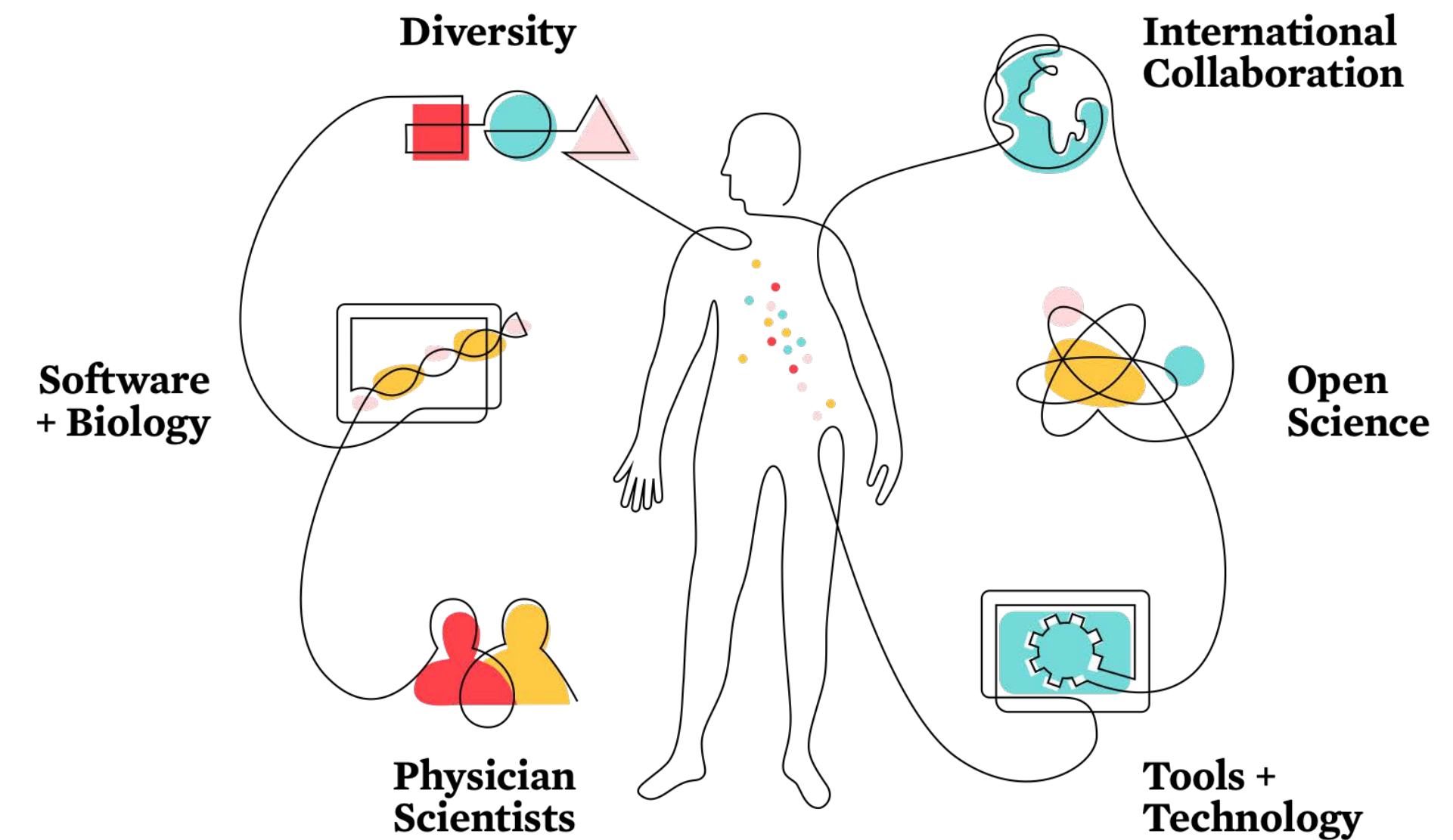


# Building our Biological Networks

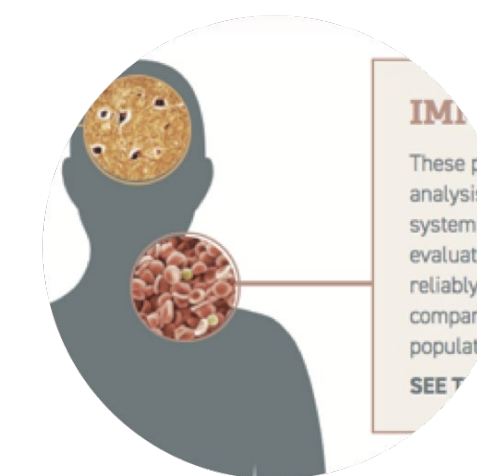
## CZI Seed Networks for the Human Cell Atlas



200+labs in 20 countries



38 projects on 10 organ systems

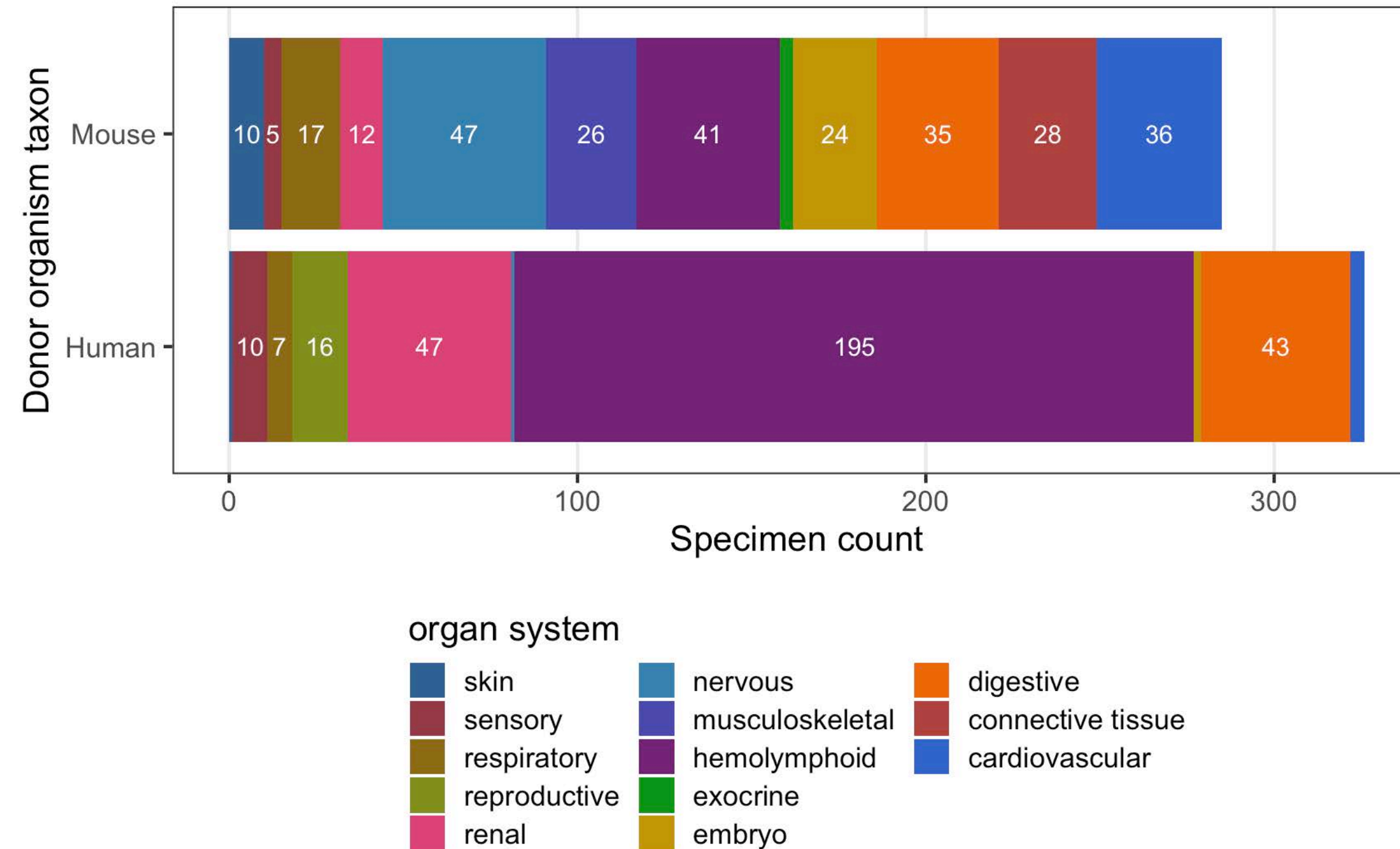


pilot projects

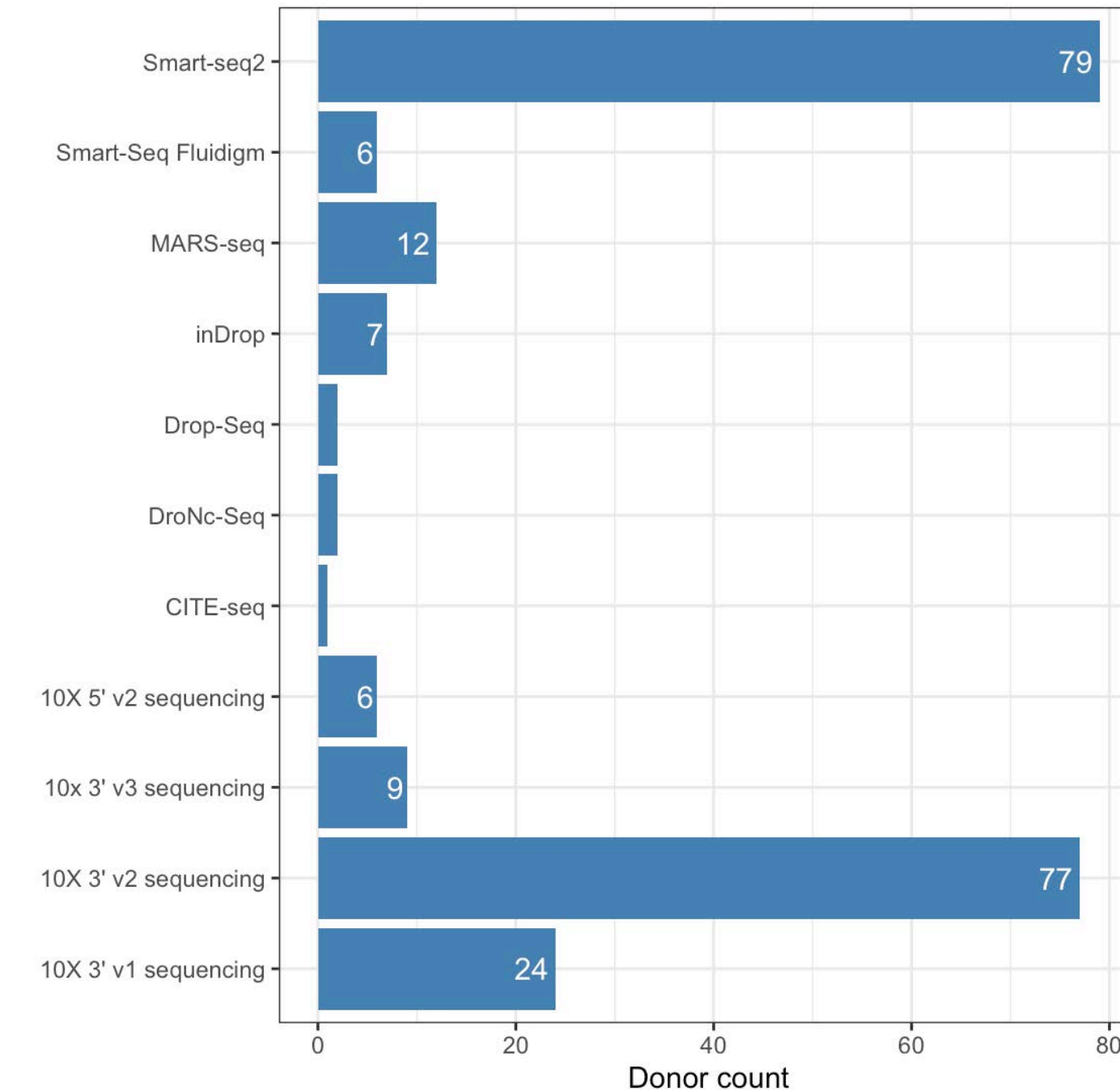


# Building a data coordination platform for HCA

Data from 13 organ systems



Donors per Assay



Pipeline



In progress

In progress



In progress



2,809,355 cells processed

8 projects in last month - 6 pre-publication projects for travel awardees

Next up - Single nucleus RNA-Seq pipelines and analysis portals

Data at [data.humancellatlas.org](https://data.humancellatlas.org)



Slide courtesy of: HCA DCP

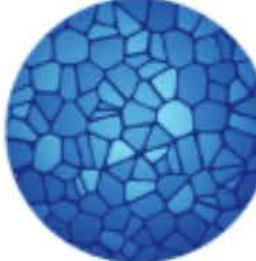
# Sharing our protocols

105 protocols, 338 members, 131 Discussions

protocols.io

SEARCHEXPLOREPLANS

Groups / Human Cell Atlas Method Development Community / Publications

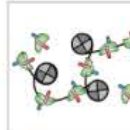


**Human Cell Atlas Method Development Community**

INTERESTS  
single-cell genomics, reference maps, molecules, cells, tissues, organs, systems

TimelineResearchPublications105Members338Discussions131Resources19News3

Category: All Publications  
SORT BY: Date




Isolation of nuclei from frozen tissue for ATAC-seq and other epigenomic assays

Aug 26, 2019

[Ryan Corces](#)<sup>1</sup>, William J. Greenleaf<sup>1</sup>, Howard Y. Chang<sup>1</sup>  
<sup>1</sup>Stanford University

[Human Cell Atlas Method Development Community](#)

CONTACT  
[Ryan Corces](#)




CGAP Human Lung Dissociation - Tissue Stability Study

Jun 17, 2019

[Anna Wilbrey-Clark](#)<sup>1</sup>, [Adam Hunter](#)<sup>2</sup>  
<sup>1</sup>Sanger Institute, Cambridge, HCA, <sup>2</sup>CGAP

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CONTACT  
[Anna Wilbrey-Clark](#)




CGAP Human Spleen Dissociation, Tissue Stability Study

Jun 11, 2019

[Anna Wilbrey-Clark](#)<sup>1</sup>, [Adam Hunter](#)<sup>2</sup>  
<sup>1</sup>Wellcome Trust Sanger Institute, HCA, <sup>2</sup>CGAP

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[Anna Wilbrey-Clark](#)



CODEX Oligo-labeled Antibody Conjugation


Aug 07, 2019

[Yury Goltsev](#)<sup>1</sup>, [Nikolay Samusik](#)<sup>1</sup>, Julia Kennedy-Darling<sup>1</sup>, [Sali Photo](#)<sup>1</sup>, [Matthew Stanford University](#)

[Version 2](#)

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[Gustavo Vazquez](#)




Human adult generic tissue dissociation \*in development\*

Jun 17, 2019

[Anna Wilbrey-Clark](#)<sup>1</sup>, [Adam Hunter](#)<sup>2</sup>  
<sup>1</sup>Sanger Institute, Cambridge, HCA, <sup>2</sup>Sanger Institute, Cambridge, CGAP

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'Frankenstein' protocol for nuclei isolation from fresh and frozen tissue for snRNAseq


Jun 01, 2019

[Luciano Martelotto](#)<sup>1</sup>  
<sup>1</sup>University of Melbourne

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[Luciano Martelotto](#)




Instructional tutorial for using *demuxlet*

Jun 26, 2019

[Hyun Min Kang](#)<sup>1</sup>, [Meena Subramaniam](#)<sup>2</sup>, [Sasha Tarr](#)<sup>2</sup>, [Michelle Nguyen](#)<sup>2</sup>, [Lanka Department of Biostatistics and Center for Statistical Genetics University of Michigan](#)

[Human Cell Atlas Method Development Community](#)

CONTACT  
[Anton Ogorodnikov](#)




CGAP Human Oesophagus Epithelium Dissociation - Tissue Stability

Jun 12, 2019

[Anna Wilbrey-Clark](#)<sup>1</sup>, [Adam Hunter](#)<sup>2</sup>  
<sup>1</sup>Wellcome Trust Sanger Institute, HCA, <sup>2</sup>CGAP

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[Anna Wilbrey-Clark](#)



scNMT-seq

May 24, 2019

[Stephen Clark](#)<sup>1</sup>  
<sup>1</sup>Babraham Institute, Cambridge

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[Stephen Clark](#)

469 views 2 bookmarks

<https://www.protocols.io/groups/hca>

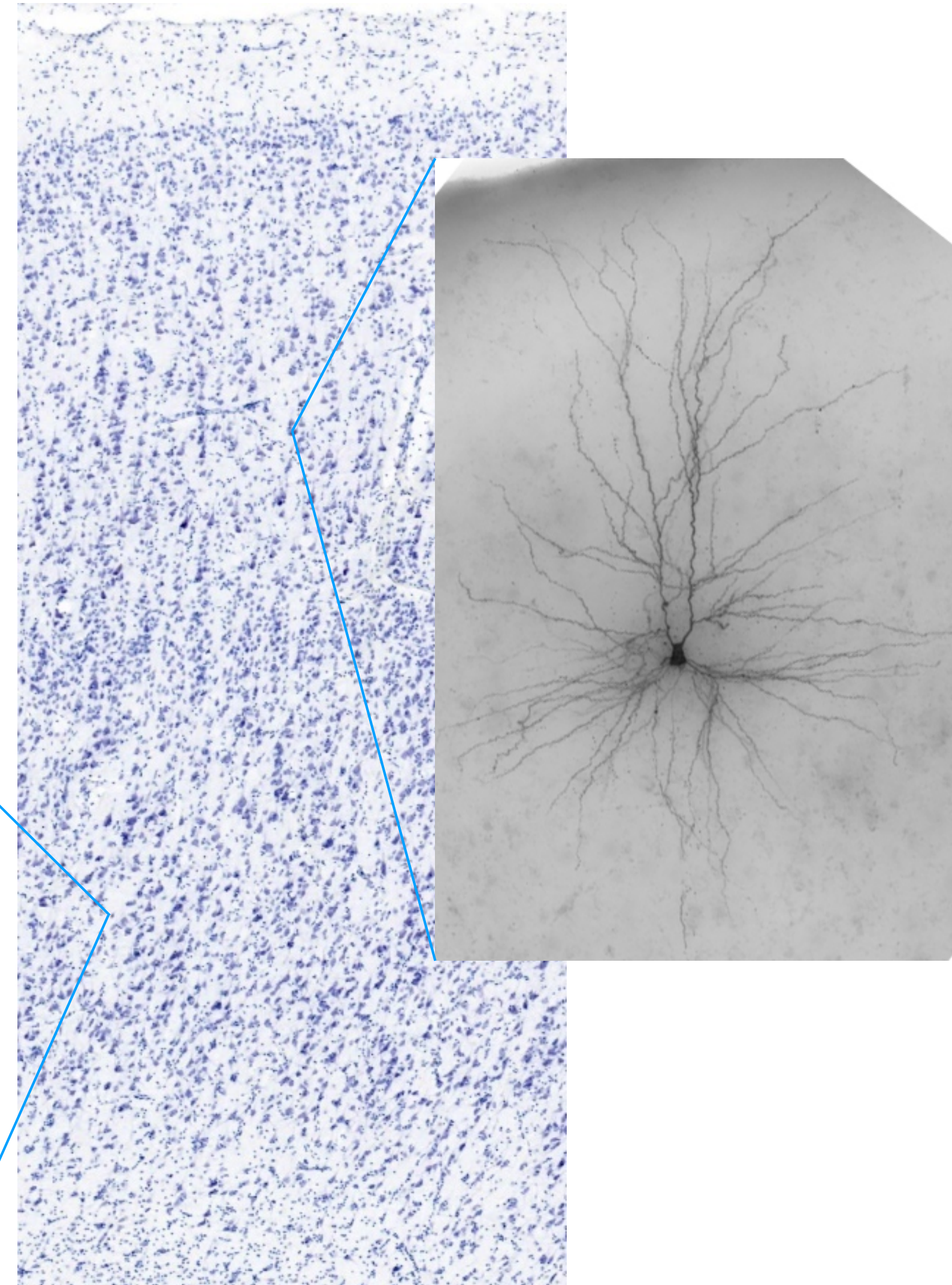


# Defining the census of brain cell types

1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18
1 H Hydrogen 1.008	2 He Helium 4.0026																
3 Li Lithium 6.94	4 Be Beryllium 9.0122	5 B Boron 10.81	6 C Carbon 12.011	7 N Nitrogen 14.007	8 O Oxygen 15.999	9 F Fluorine 18.998	10 Ne Neon 20.180										
11 Na Sodium 22.990	12 Mg Magnesium 24.305	13 Al Aluminum 26.982	14 Si Silicon 28.086	15 P Phosphorus 30.974	16 S Sulfur 32.06	17 Cl Chlorine 35.45	18 Ar Argon 39.948										
19 K Potassium 39.098	20 Ca Calcium 40.078	21 Sc Scandium 44.956	22 Ti Titanium 47.867	23 V Vanadium 50.942	24 Cr Chromium 51.996	25 Mn Manganese 54.938	26 Fe Iron 55.845	27 Co Cobalt 58.933	28 Ni Nickel 58.693	29 Cu Copper 63.546	30 Zn Zinc 65.38	31 Ga Gallium 69.723	32 Ge Germanium 72.630	33 As Arsenic 74.922	34 Se Selenium 78.971	35 Br Bromine 79.904	36 Kr Krypton 83.798
37 Rb Rubidium 85.468	38 Sr Strontium 87.62	39 Y Yttrium 88.906	40 Zr Zirconium 91.224	41 Nb Niobium 92.906	42 Mo Molybdenum 95.94	43 Tc Technetium 98.906	44 Ru Ruthenium 101.07	45 Rh Rhodium 102.91	46 Pd Palladium 106.90	47 Ag Silver 107.87	48 Cd Cadmium 112.41	49 In Indium 114.82	50 Sn Tin 118.71	51 Sb Antimony 121.76	52 Te Tellurium 127.60	53 I Iodine 126.90	54 Xe Xenon 131.29
55 Cs Cesium 132.91	56 Ba Barium 137.33	57-71 La-Lu Lanthanides	72 Hf Hafnium 178.49	73 Ta Tantalum 180.95	74 W Tungsten 183.84	75 Re Rhenium 186.21	76 Os Osmium 190.23	77 Ir Iridium 192.22	78 Pt Platinum 195.08	79 Au Gold 196.97	80 Hg Mercury 200.59	81 Tl Thallium 204.38	82 Pb Lead 207.2	83 Bi Bismuth 208.98	84 Po Polonium 209	85 At Astatine 210	86 Rn Radon 222
87 Fr Francium [223]	88 Ra Radium [226]	89-103 Ac-Lr Actinides	104 Db Dubnium [261]	105 Sg Seaborgium [266]	106 Bh Bohrium [264]	107 Hs Hassium [277]	108 Mt Meitnerium [268]	109 Ds Darmstadtium [271]	110 Nh Nihonium [286]	111 Rg Roentgenium [289]	112 Cn Copernicium [285]	113 Nh Nihonium [284]	114 Fl Flerovium [289]	115 Lv Livermorium [293]	116 Ts Tennessine [294]	117 Og Oganesson [294]	118 Lr Lawrencium [260]

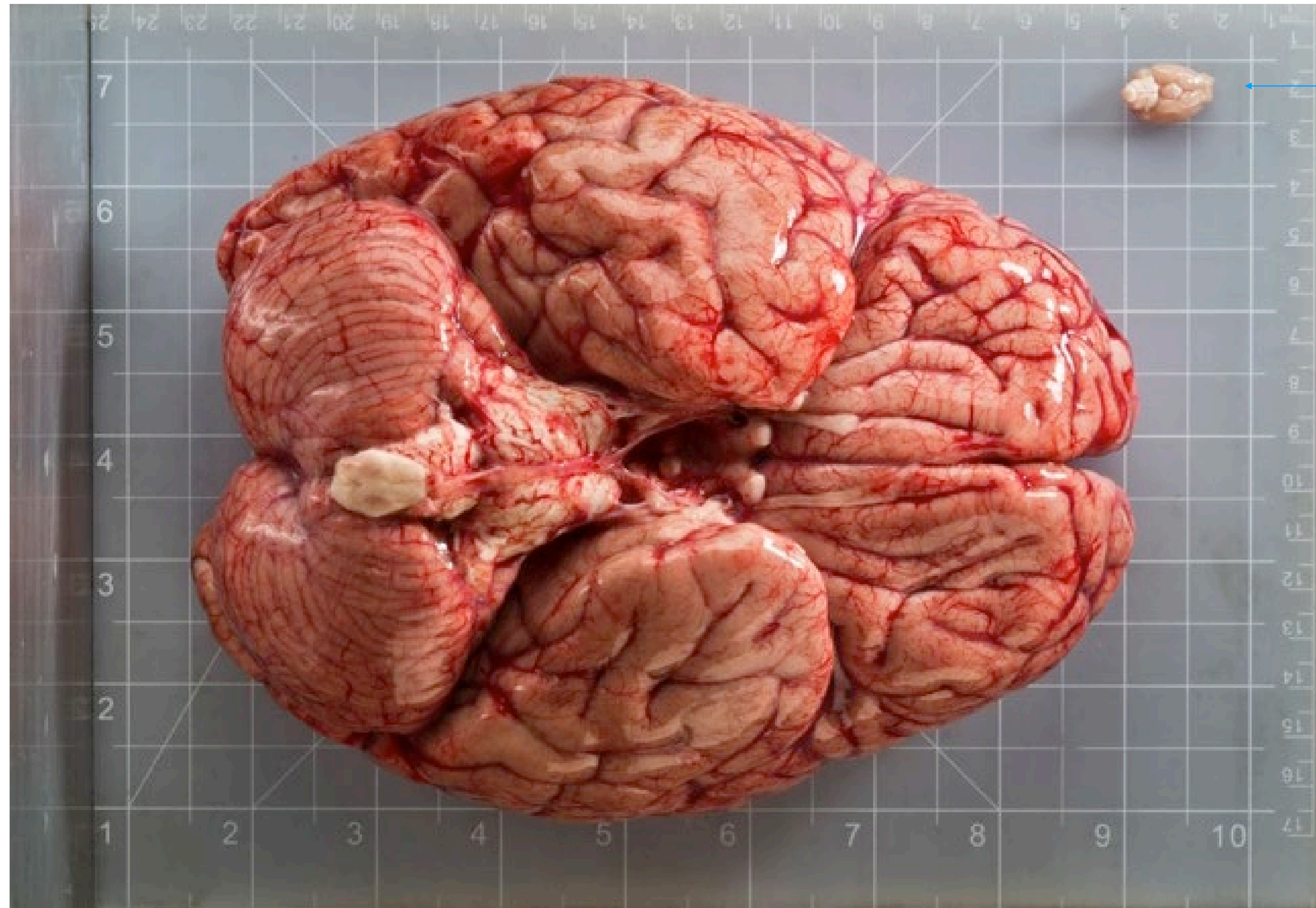
For elements with no stable isotopes, the mass number of the isotope with the longest half-life is in parentheses.

57 La Lanthanum 138.91	58 Ce Cerium 140.12	59 Pr Praseodymium 140.91	60 Nd Neodymium 144.24	61 Pm Promethium [145]	62 Sm Samarium 150.36	63 Eu Europium 151.96	64 Gd Gadolinium 157.25	65 Tb Terbium 158.93	66 Dy Dysprosium 162.50	67 Ho Holmium 164.93	68 Er Erbium 167.26	69 Tm Thulium 168.93	70 Yb Ytterbium 173.05	71 Lu Lutetium 174.97
89 Ac Actinium [227]	90 Th Thorium 232.04	91 Pa Protactinium 231.04	92 U Uranium 238.03	93 Np Neptunium [237]	94 Pu Plutonium [244]	95 Am Americium [243]	96 Cm Curium [247]	97 Bk Berkelium [247]	98 Cf Californium [251]	99 Es Einsteinium [252]	100 Fm Fermium [257]	101 Md Mendelevium [258]	102 No Nobelium [259]	103 Lr Lawrencium [260]





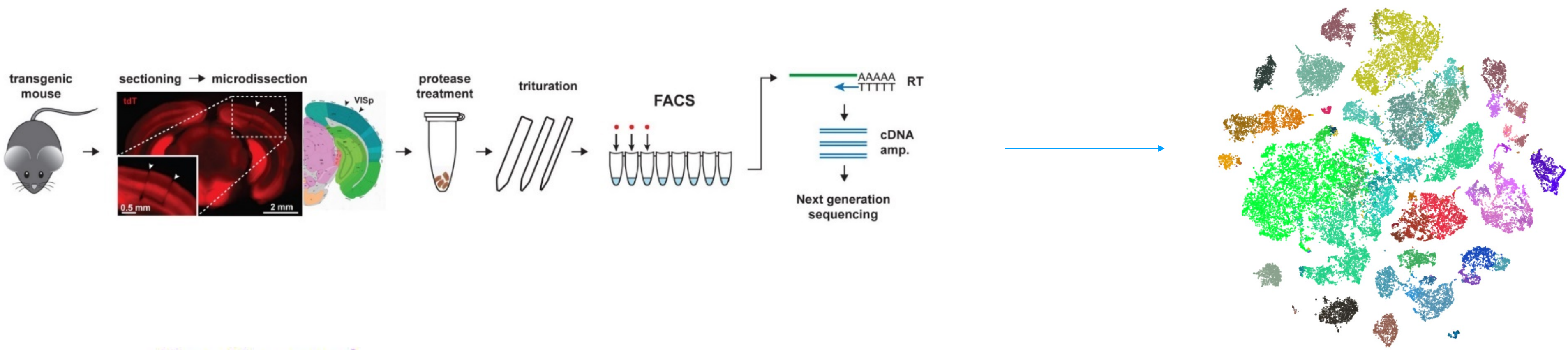
# Major challenges in characterizing human brain compared to model organisms



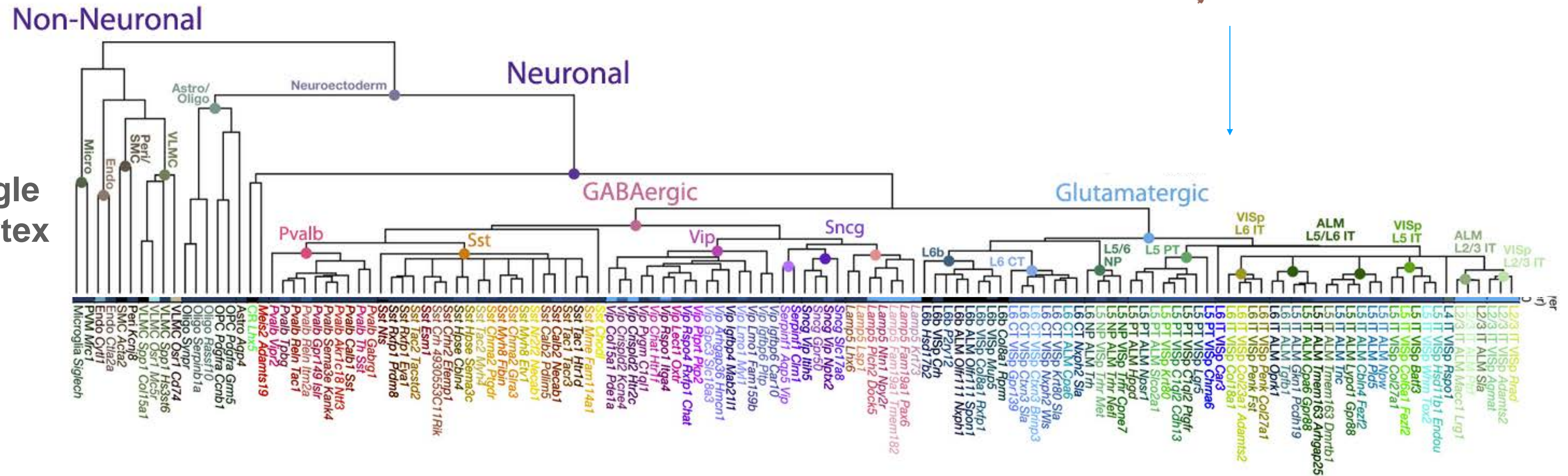
**Mouse**



# Single cell transcriptomics provides an unbiased and essentially complete molecular classification of mouse cortical cell types



~100 cell types in a single region of the mouse cortex

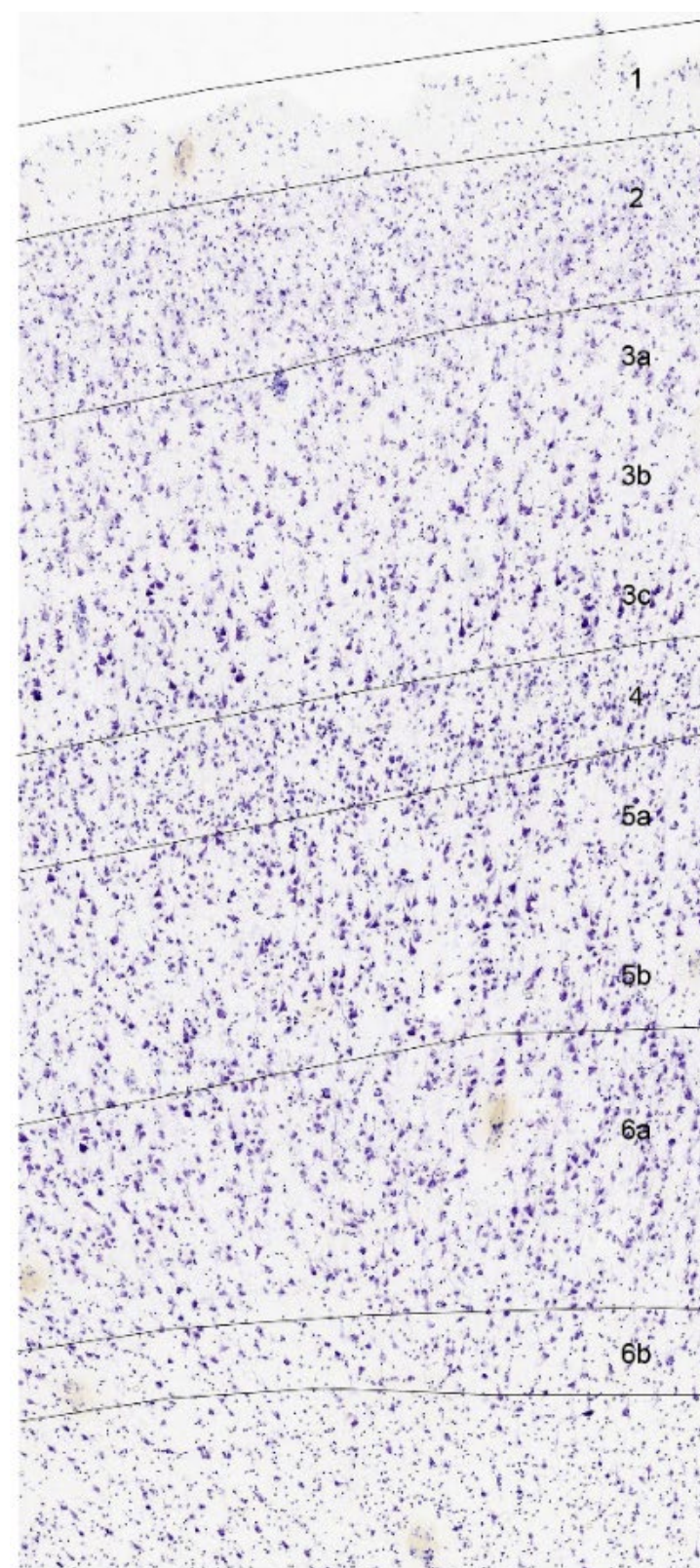


Tasic, Yao, Smith, Graybuck...Koch, Zeng (2018) Nature

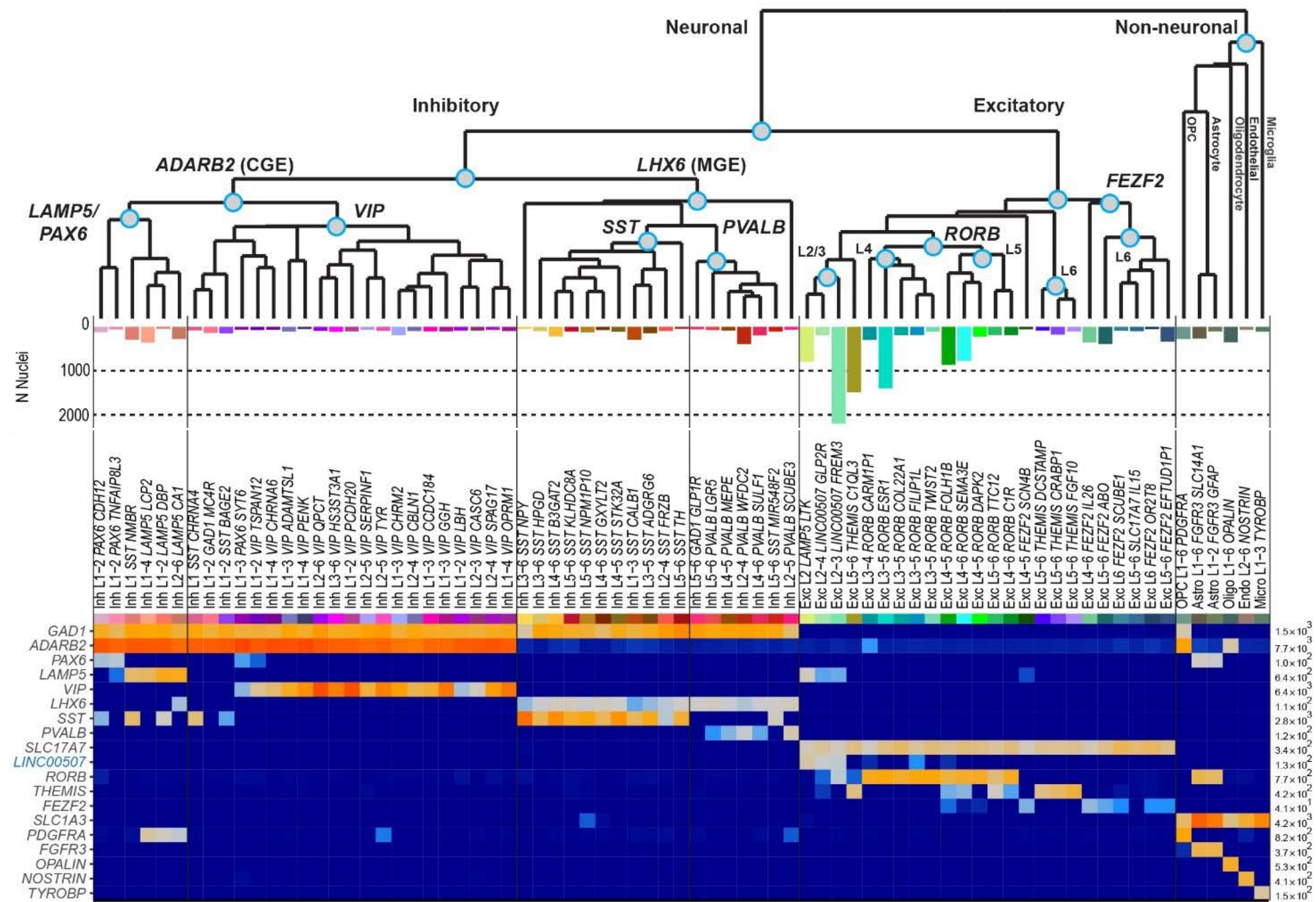


# Similarly detailed cellular classifications can be generated in human cortex using single nucleus transcriptomics

Most cell types are rare



Human MTG

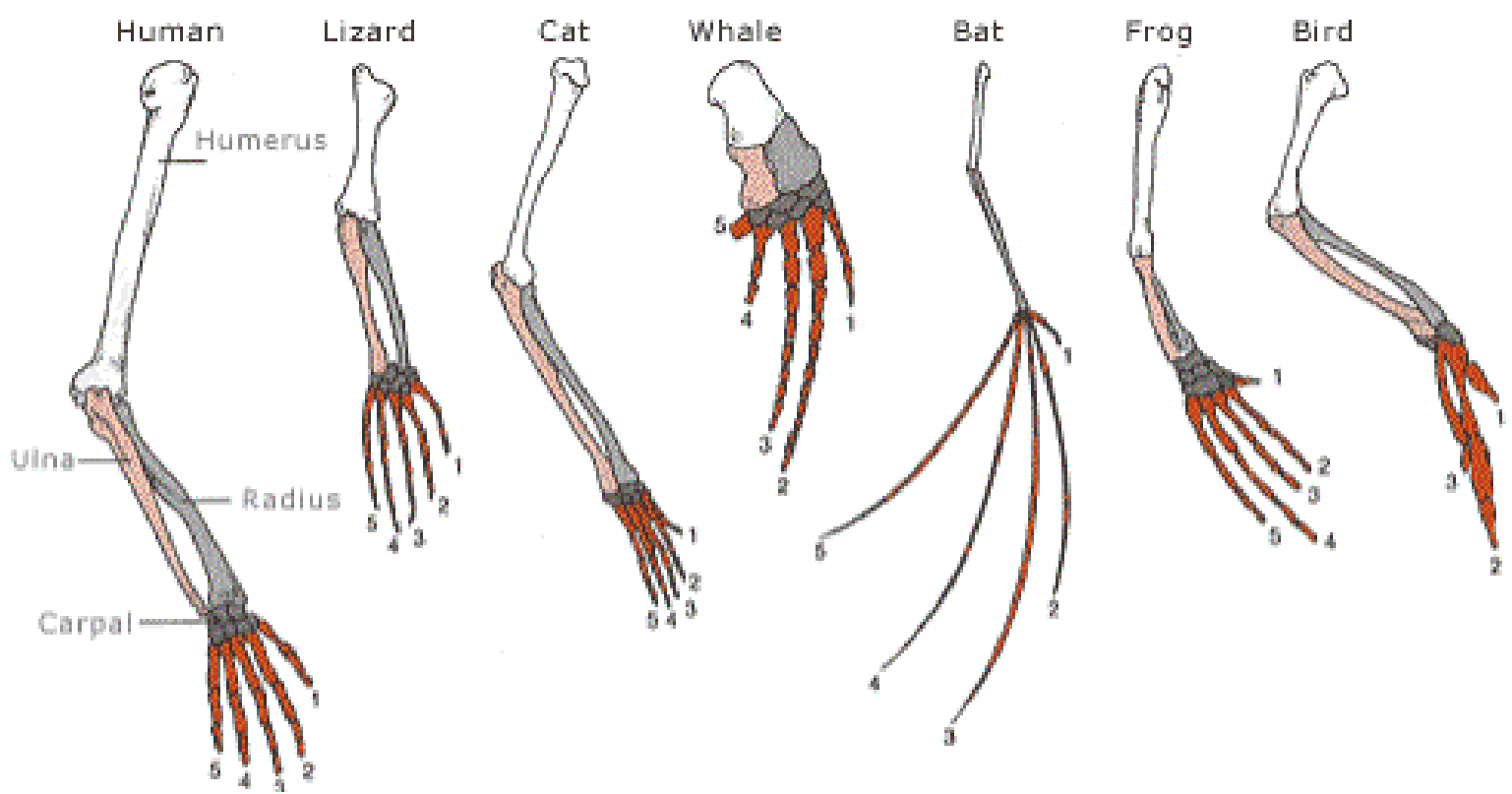


Hodge, Bakken, Miller...Lein (2019) Nature

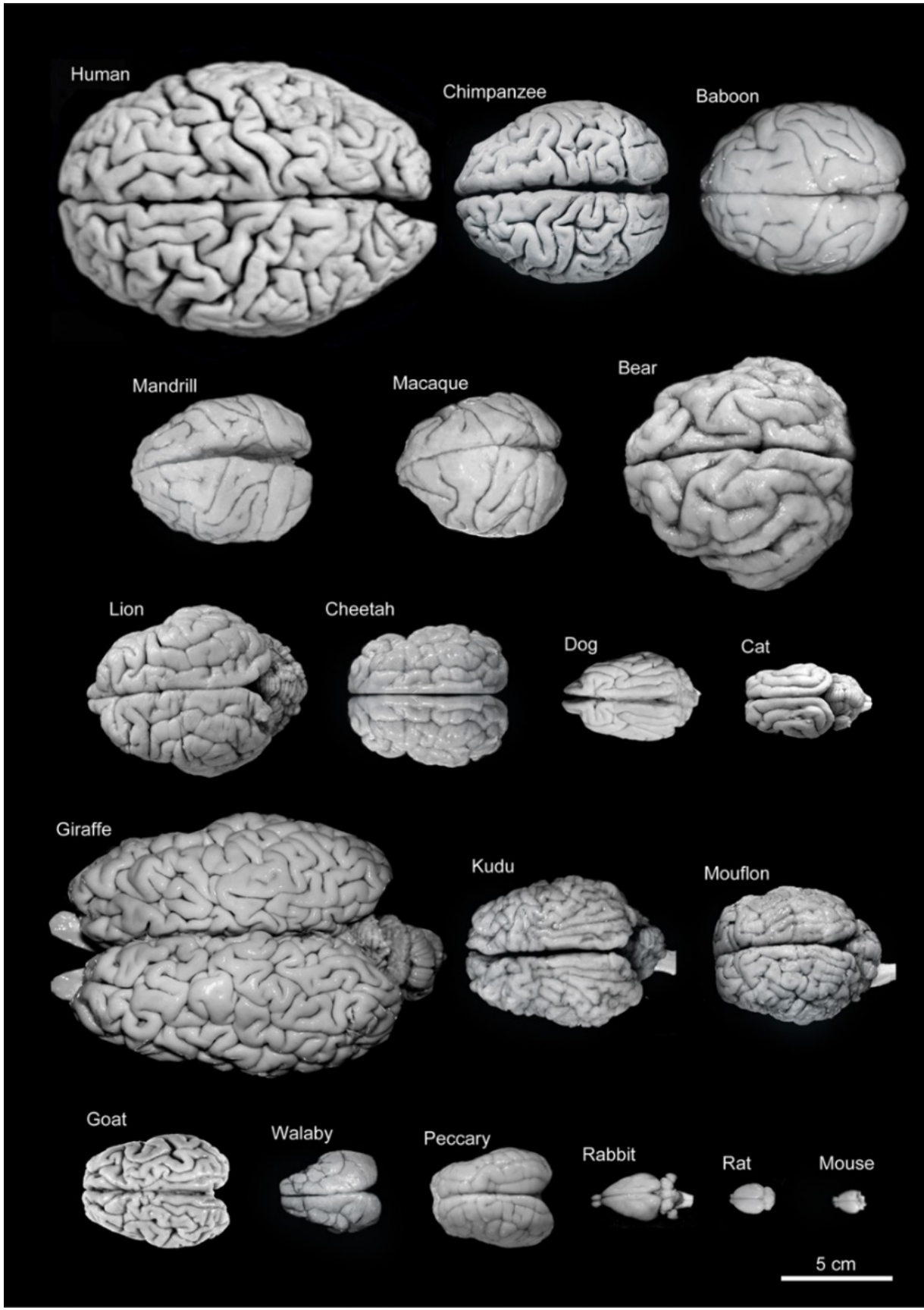


# Evolutionary principles explain similarities and differences between species

## Limb homology

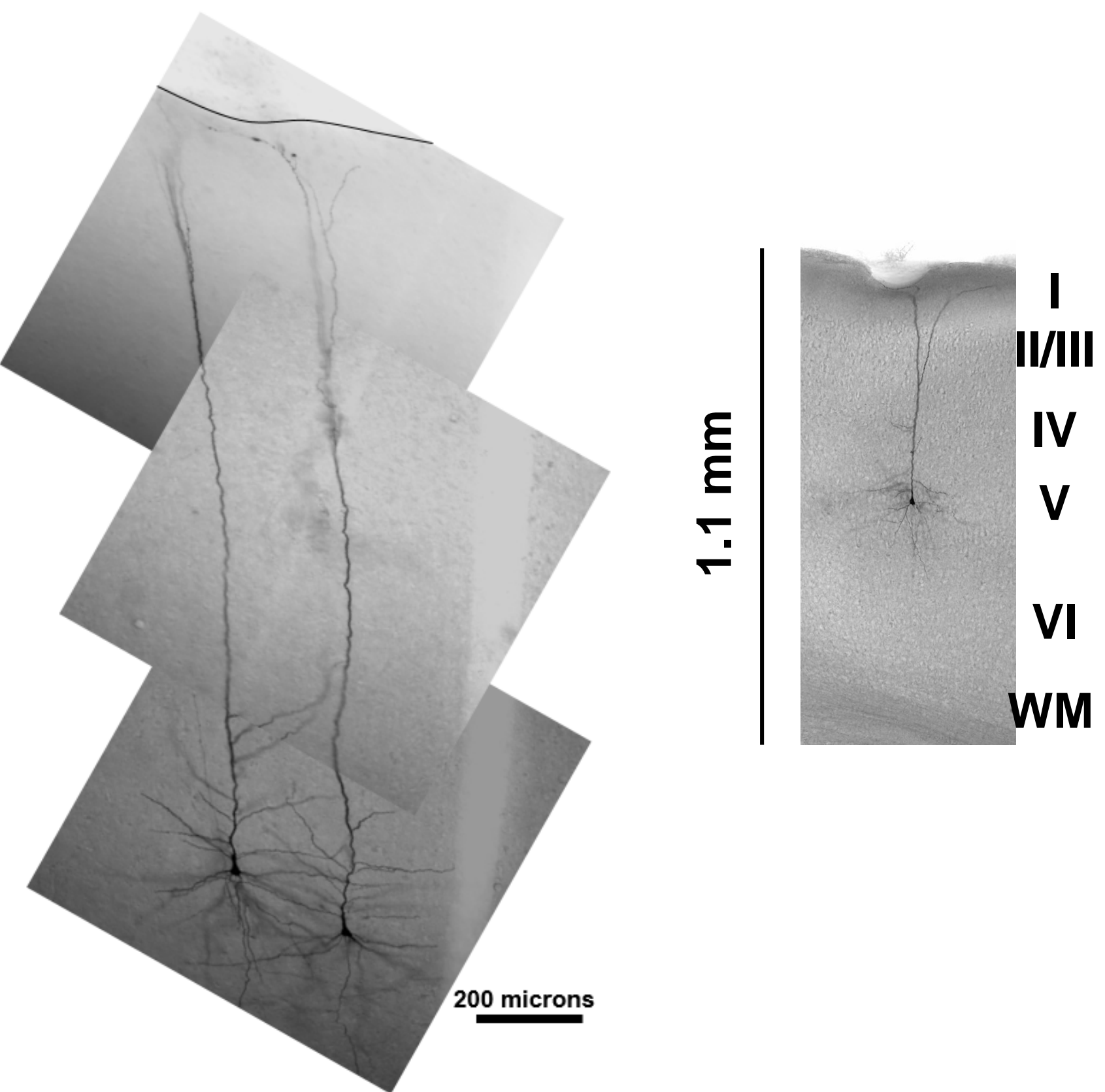


## Cortical size and gyrification



DeFelipe 2011

## Cell Types



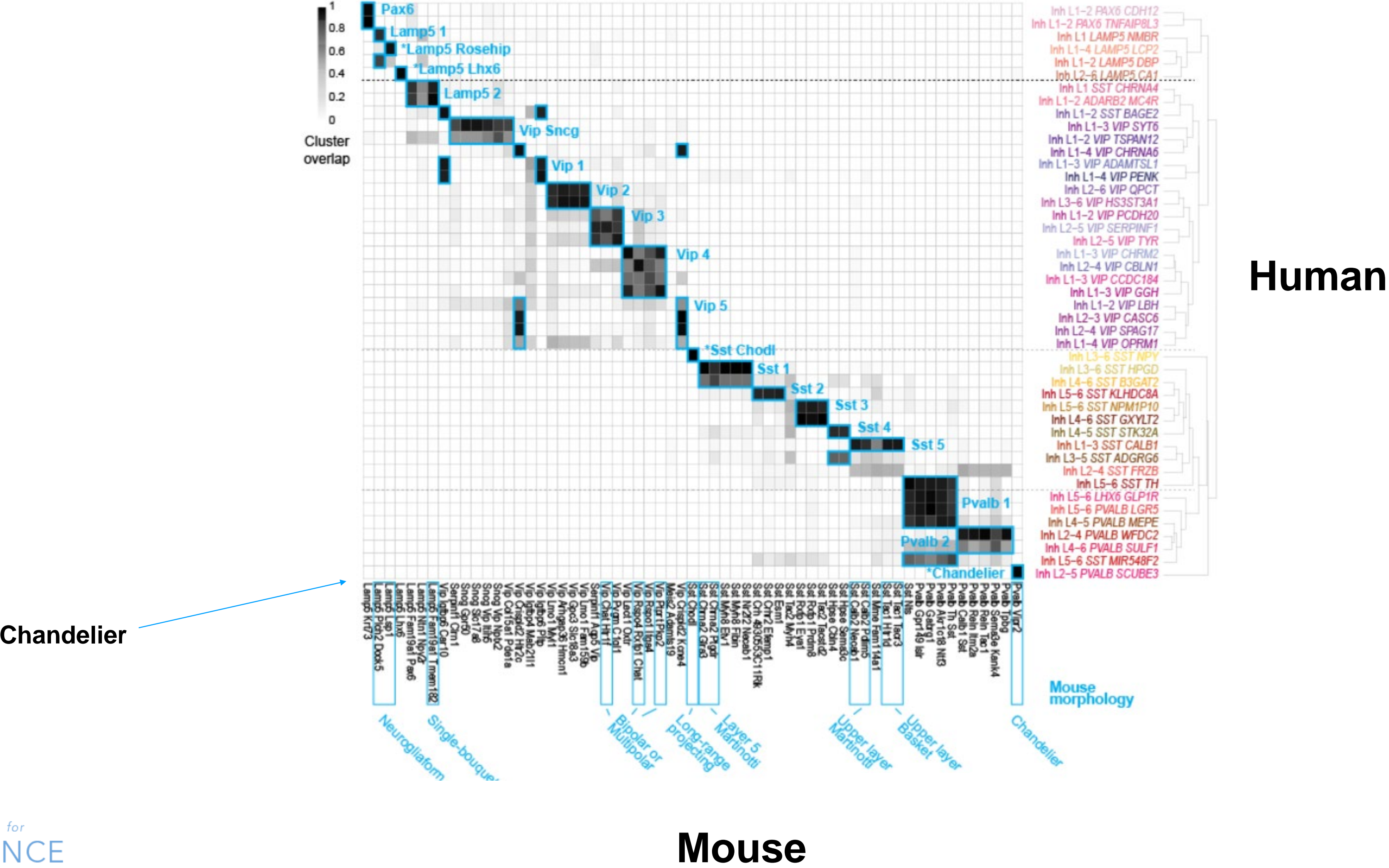
Human

Mouse



# The makeup of cortical cell types is very similar between human and mouse

“Homology mapping” to align cell types across species

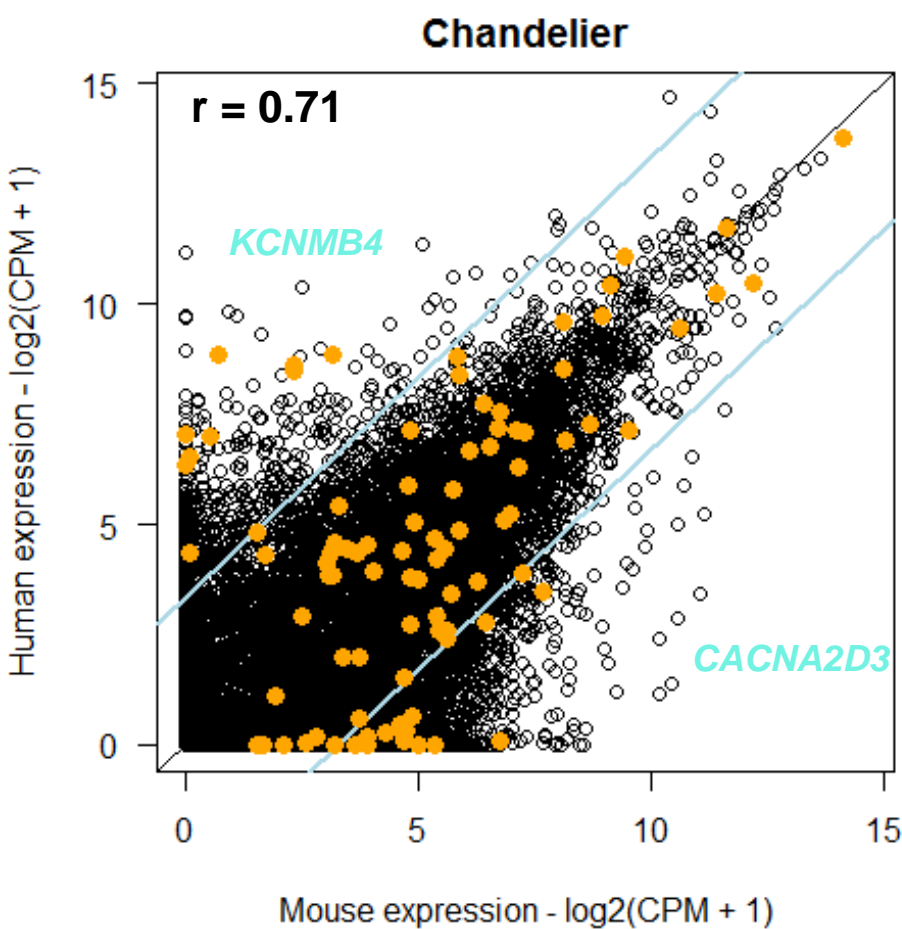




# Homologous cell types have many differences in gene usage across species

The genes are highly conserved but their cellular usage often changes

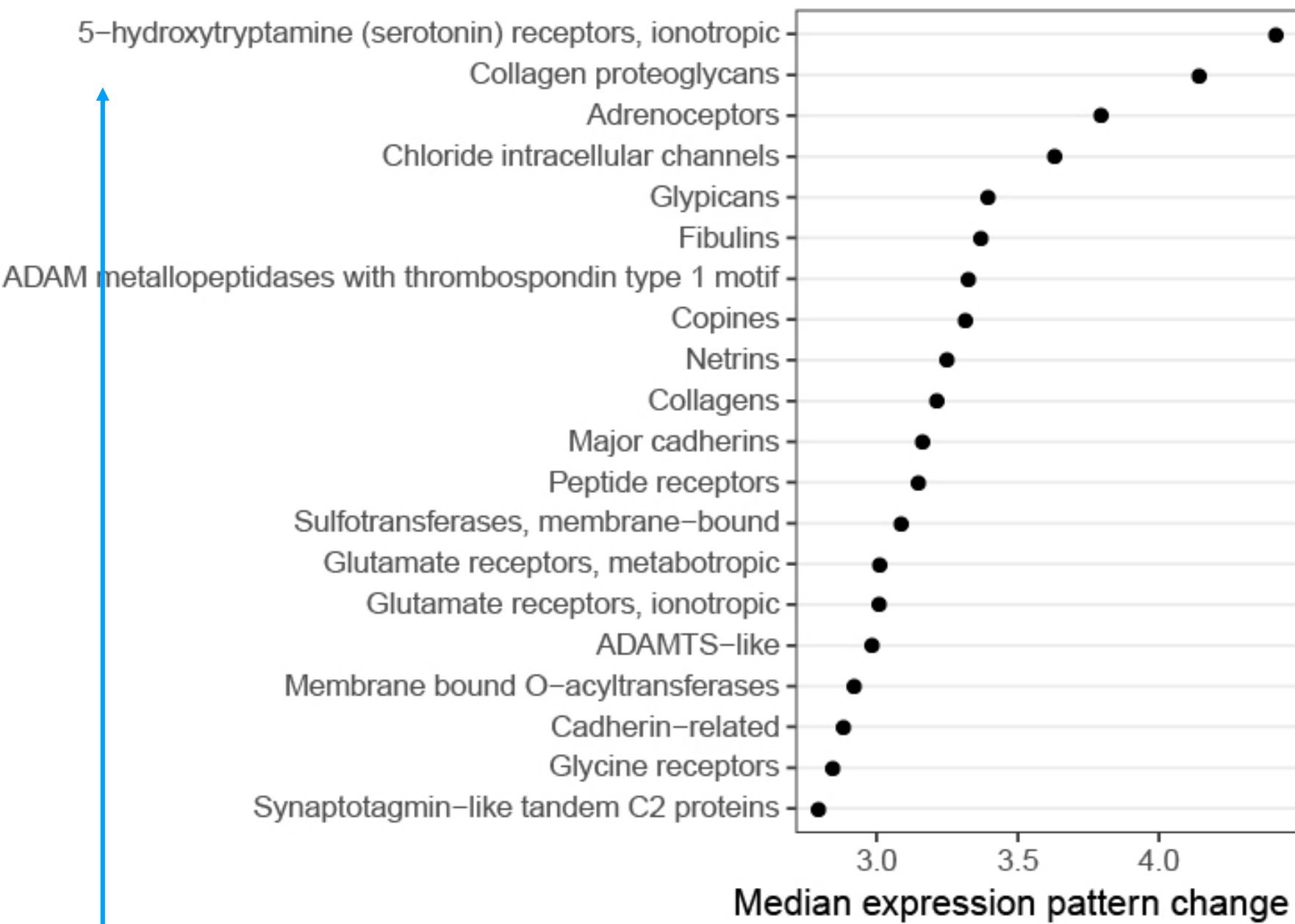
Gene expression in cell types is highly conserved overall



But 12-20% of the ~8000 genes detected have >10x difference

Genes with different are functionally important

Top 20 most divergent functional gene classes



The gene family showing the most differences between human and mouse is involved in serotonin signaling, which is associated with major depression, mood disorders, schizophrenia, addiction, ADHD and autism.



# The genetically-based Cell Atlas is only the beginning

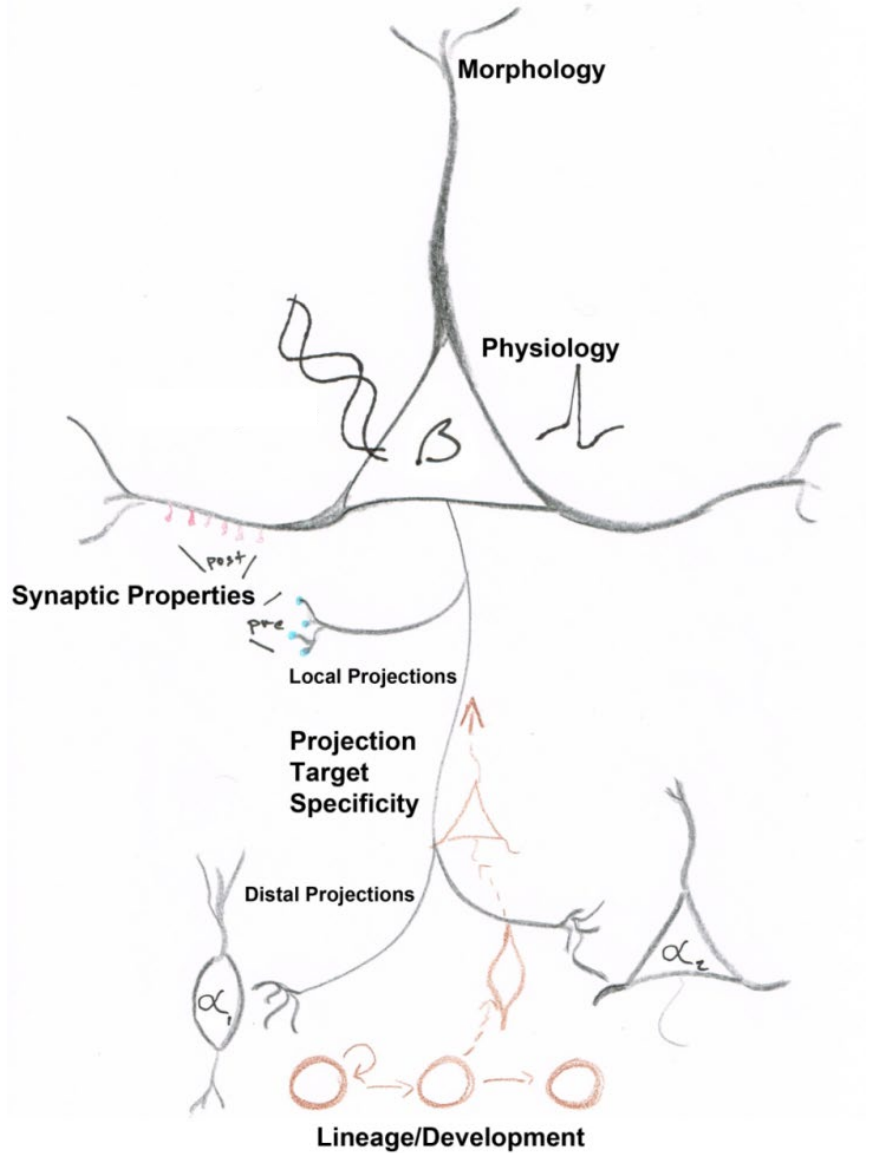
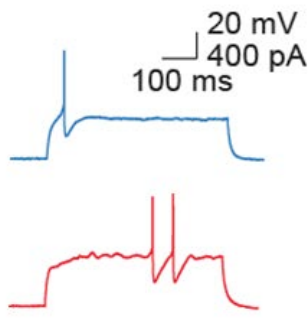
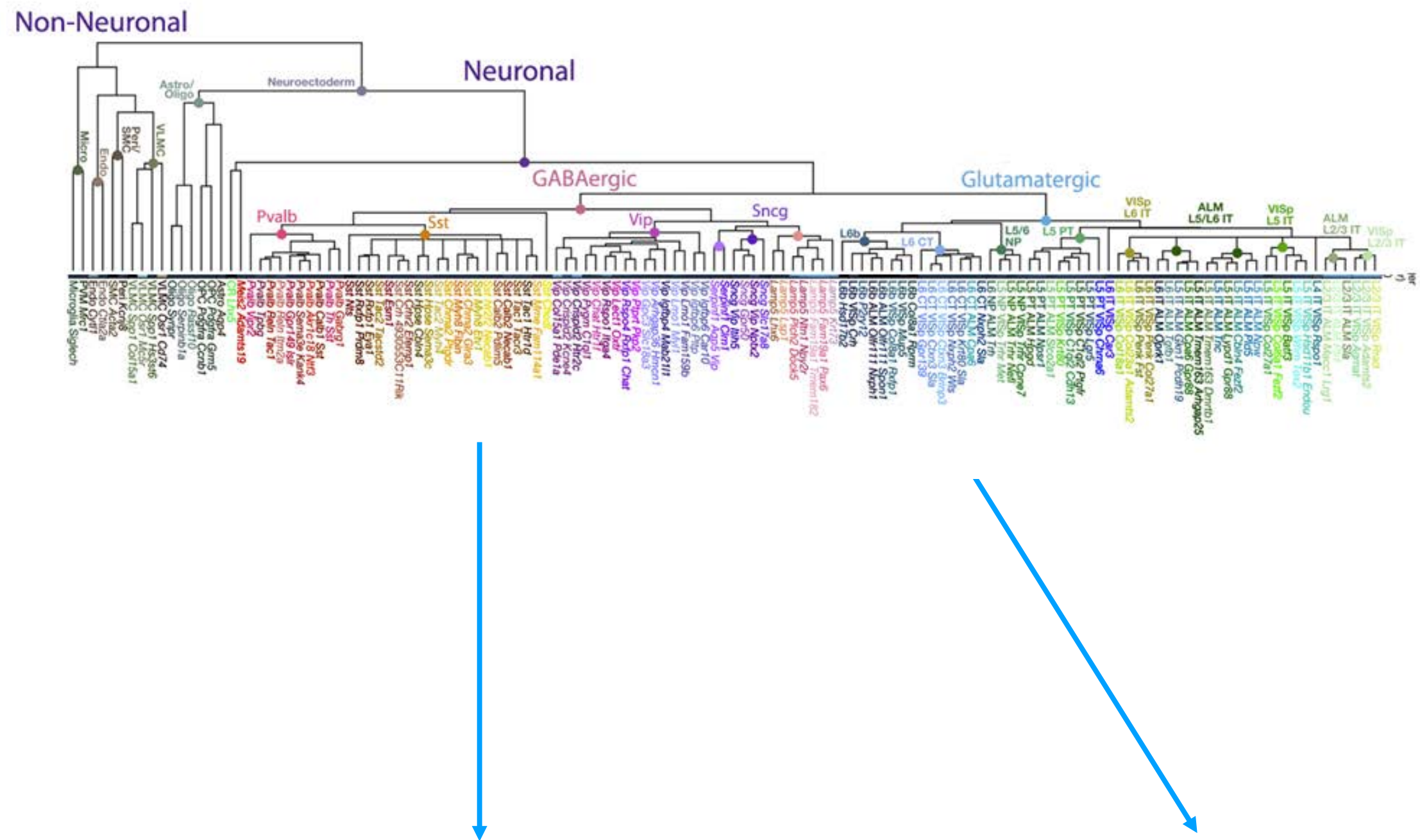
Understand the properties of cell types

Cell type classification based on Single Cell Transcriptomics

Electrophysiology

Connectivity,  
function

Anatomy



Understand the cellular basis of  
brain diseases

Develop viral genetic tools for  
selective genetic manipulation and  
gene therapy



### A Unique Microglia Type Associated with Restricting Development of Alzheimer’s Disease

Hadas Keren-Shaul,<sup>1,6</sup> Amit Spinrad,<sup>1,2,6</sup> Assaf Weiner,<sup>1,3,6,\*</sup> Orit Matcovitch-Natan,<sup>1,2,6</sup> Raz Dvir-Szternfeld,<sup>2</sup> Tyler K. Ulland,<sup>4</sup> Eyal David,<sup>1</sup> Kuti Baruch,<sup>2</sup> David Lara-Astaiso,<sup>1</sup> Beata Toth,<sup>5</sup> Shalev Itzkovitz,<sup>6</sup> Marco Colonna,<sup>4</sup> Michal Schwartz,<sup>2,7,\*</sup> and Ido Amit<sup>1,7,8,\*</sup>

<sup>1</sup>Department of Immunology, Weizmann Institute of Science, Rehovot 7610001, Israel  
<sup>2</sup>Department of Neurobiology, Weizmann Institute of Science, Rehovot 7610001, Israel  
<sup>3</sup>Hubrecht Institute-KNAW (Royal Netherlands Academy of Arts and Sciences), and University Medical Center, Cancer Genomics Netherlands, 3584 CG Utrecht, the Netherlands  
<sup>4</sup>Department of Pathology and Immunology, Washington University School of Medicine, St. Louis, MO 63110, USA  
<sup>5</sup>Department of Molecular Cell Biology, Weizmann Institute of Science, Rehovot 7610001, Israel  
<sup>6</sup>These authors contributed equally  
<sup>7</sup>Senior author  
<sup>8</sup>Lead Contact  
\*Correspondence: assaf.weiner@weizmann.ac.il (A.W.), michal.schwartz@weizmann.ac.il (M.S.), ido.amit@weizmann.ac.il (I.A.)  
<http://dx.doi.org/10.1016/j.cell.2017.05.018>

#### SUMMARY

Alzheimer’s disease (AD) is a detrimental neurodegenerative disease with no effective treatments. Due to cellular heterogeneity, defining the roles of immune cell subsets in AD onset and progression has been challenging. Using transcriptional single-cell sorting, we comprehensively map all immune populations in wild-type and AD-transgenic (Tg-AD) mouse brains. We describe a novel microglia type associated with neurodegenerative diseases (DAM) and identify markers, spatial localization, and pathways associated with these cells. Immunohistochemical staining of mice and human brain slices shows DAM with intracellular/phagocytic Aβ particles. Single-cell analysis of DAM in Tg-AD and triggering receptor expressed on myeloid cells 2 (Trem2)<sup>−/−</sup> Tg-AD reveals that the DAM program is activated in a two-step process. Activation is initiated in a Trem2-independent manner that involves downregulation of microglia checkpoints, followed by activation of a Trem2-dependent program. This unique microglia-type has the potential to restrict neurodegeneration, which may have important implications for future treatment of AD and other neurodegenerative diseases.

*et al., 2011; Ziv et al., 2006*). In the mouse embryo, microglia migrate from the yolk sac to the CNS at embryonic days 8–9, undergo a stepwise program of development that is synchronized with the brain developmental process, and subsequently acquire a stable phenotype essential for the brain protection and homeostasis (Ginhoux and Prinz, 2015; Matcovitch-Natan et al., 2016). Microglia immune activity is restrained by dedicated immune inhibitory pathways that suppress unwanted inflammatory responses and tissue destruction that are often associated with immune activation (Hanisch and Kettenmann, 2007). These checkpoint mechanisms include direct inhibitory interactions of microglia with neurons through the receptor-ligand pairs CX3CL1-CX3CR1 and CD200-CD200R, soluble molecules present in the CNS milieu (e.g., transforming growth factor β [TGF-β]), and intracellular regulators such as the transcription factor MafB (Butovsky et al., 2015; Kierdorf and Prinz, 2013; Lauro et al., 2015; Matcovitch-Natan et al., 2016; Ransohoff and Cardona, 2010). Nevertheless, these mechanisms may be disadvantageous under extreme conditions when reparative microglial activity is needed.










Alzheimer’s disease (AD) is an age-related neurodegenerative disease characterized by progressive memory decline and cognitive dysfunction, often manifested histologically by the parenchymal deposition of amyloid-beta (Aβ) plaques, the formation of neurofibrillary tangles and neuroinflammation (Hardy and Selkoe, 2002; Holtzman et al., 2011). Numerous studies reported conflicting results regarding the contribution of systemic immunity, recruited monocytes, and tissue-resident microglia to AD onset and disease progression (Baruch et al., 2016; Deardorff and Greenham, 2017; Guillot-Saïrier et al., 2015; Ibar et al., 2015;

## Single-cell transcriptomic analysis of Alzheimer’s disease

Hansruedi Mathys<sup>1,2,10</sup>, Jose Davila-Velderrain<sup>3,4,10</sup>, Zhuyu Peng<sup>1,2</sup>, Fan Gao<sup>1,2</sup>, Shahin Mohammadi<sup>3,4</sup>, Jennie Z. Young<sup>1,2</sup>, Madhvi Menon<sup>4,5,6</sup>, Liang He<sup>3,4</sup>, Fatema Abdurrob<sup>1,2</sup>, Xueqiao Jiang<sup>1,2</sup>, Anthony J. Martorell<sup>1,2</sup>, Richard M. Ransohoff<sup>7</sup>, Brian P. Hafler<sup>4,5,6,8</sup>, David A. Bennett<sup>9</sup>, Manolis Kellis<sup>3,4,11\*</sup> & Li-Huei Tsai<sup>1,2,4,11\*</sup>

Alzheimer’s disease is a pervasive neurodegenerative disorder, the molecular complexity of which remains poorly understood. Here, we analysed 80,660 single-nucleus transcriptomes from the prefrontal cortex of 48 individuals with varying degrees of Alzheimer’s disease pathology. Across six major brain cell types, we identified transcriptionally distinct subpopulations, including those associated with pathology and characterized by regulators of myelination, inflammation, and neuron survival. The strongest disease-associated changes appeared early in pathological progression and were highly cell-type specific, whereas genes upregulated at late stages were common across cell types and primarily involved in the global stress response. Notably, we found that female cells were overrepresented in disease-associated subpopulations, and that transcriptional responses were substantially different between sexes in several cell types, including oligodendrocytes. Overall, myelination-related processes were recurrently perturbed in multiple cell types, suggesting that myelination has a key role in Alzheimer’s disease pathophysiology. Our single-cell transcriptomic resource provides a blueprint for interrogating the molecular and cellular basis of Alzheimer’s disease.

## A single-cell atlas of entorhinal cortex from individuals with Alzheimer’s disease reveals cell-type-specific gene expression regulation

Alexandra Grubman<sup><sup>1,2,3,9</sup></sup>, Gabriel Chew<sup>4,9</sup>, John F. Ouyang<sup><sup>4,9</sup></sup>, Guizhi Sun<sup>1,2,3</sup>, Xin Yi Choo<sup><sup>1,2,3,5</sup></sup>, Catriona McLean<sup><sup>6</sup></sup>, Rebecca K. Simmons<sup>7,8</sup>, Sam Buckberry<sup>7,8</sup>, Dulce B. Vargas-Landin<sup><sup>7,8</sup></sup>, Daniel Poppe<sup>7,8</sup>, Jahnvi Pflueger<sup>7,8</sup>, Ryan Lister<sup><sup>7,8</sup></sup>, Owen J. L. Rackham<sup><sup>4\*</sup></sup>, Enrico Petretto<sup><sup>4\*</sup></sup> and Jose M. Polo<sup><sup>1,2,3\*</sup></sup>

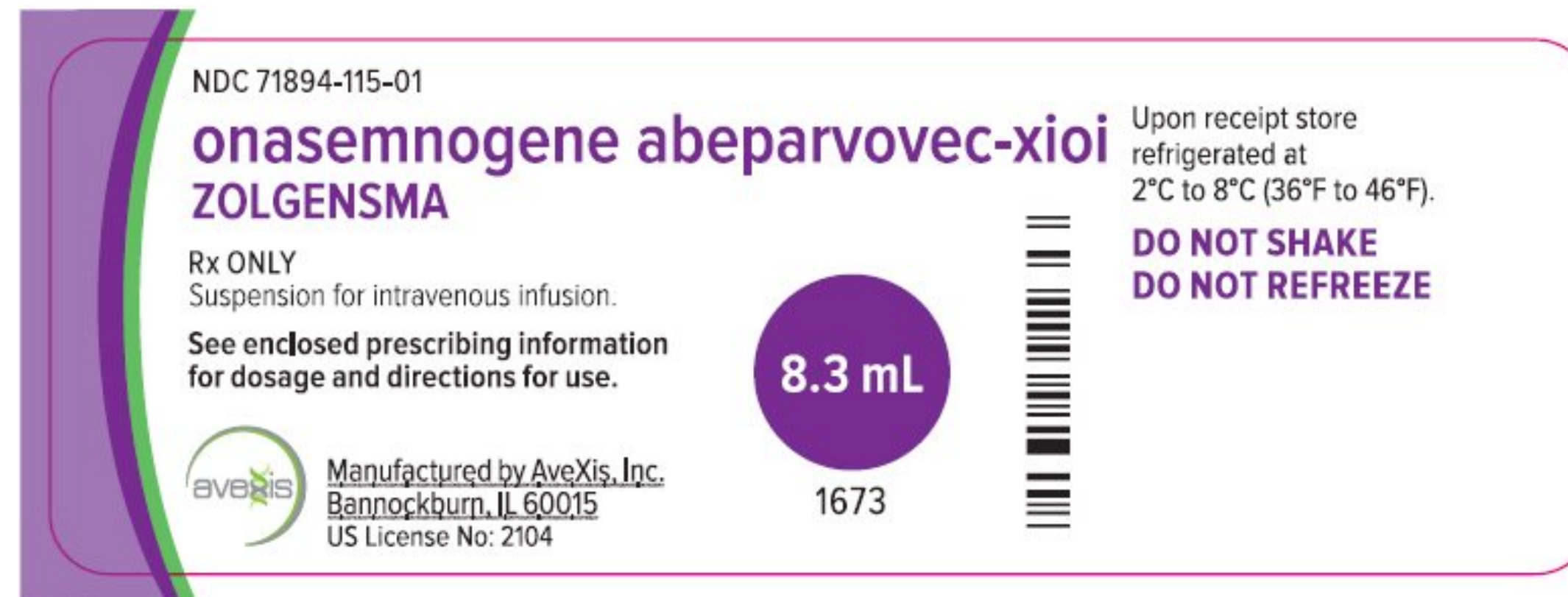
There is currently little information available about how individual cell types contribute to Alzheimer’s disease. Here we applied single-nucleus RNA sequencing to entorhinal cortex samples from control and Alzheimer’s disease brains (*n* = 6 per group), yielding a total of 13,214 high-quality nuclei. We detail cell-type-specific gene expression patterns, unveiling how transcriptional changes in specific cell subpopulations are associated with Alzheimer’s disease. We report that the Alzheimer’s disease risk gene *APOE* is specifically repressed in Alzheimer’s disease oligodendrocyte progenitor cells and astrocyte subpopulations and upregulated in an Alzheimer’s disease-specific microglial subpopulation. Integrating transcription factor regulatory modules with Alzheimer’s disease risk loci revealed drivers of cell-type-specific state transitions towards Alzheimer’s disease. For example, transcription factor EB, a master regulator of lysosomal function, regulates multiple disease genes in a specific Alzheimer’s disease astrocyte subpopulation. These results provide insights into the coordinated control of Alzheimer’s disease risk genes and their cell-type-specific contribution to disease susceptibility. These results are available at <http://adsn.ddnetbio.com>.



# AAV-based gene therapy for the nervous system is becoming a reality



**Biallelic RPE65 mutation-associated retinal dystrophy**



**SMN1 mutation-associated Spinal Muscular Atrophy**

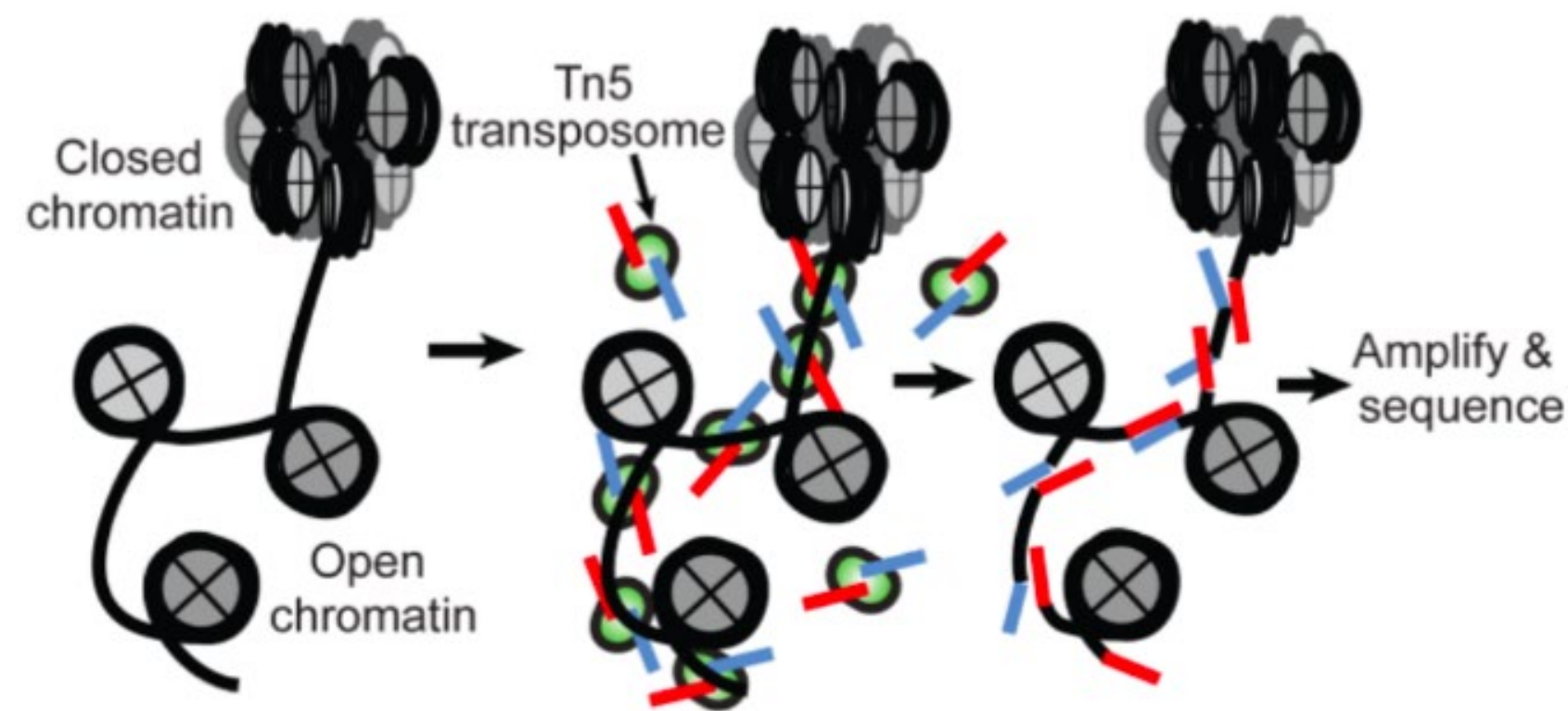


# A genetic cell classification leads directly to genetic tools to target cell types

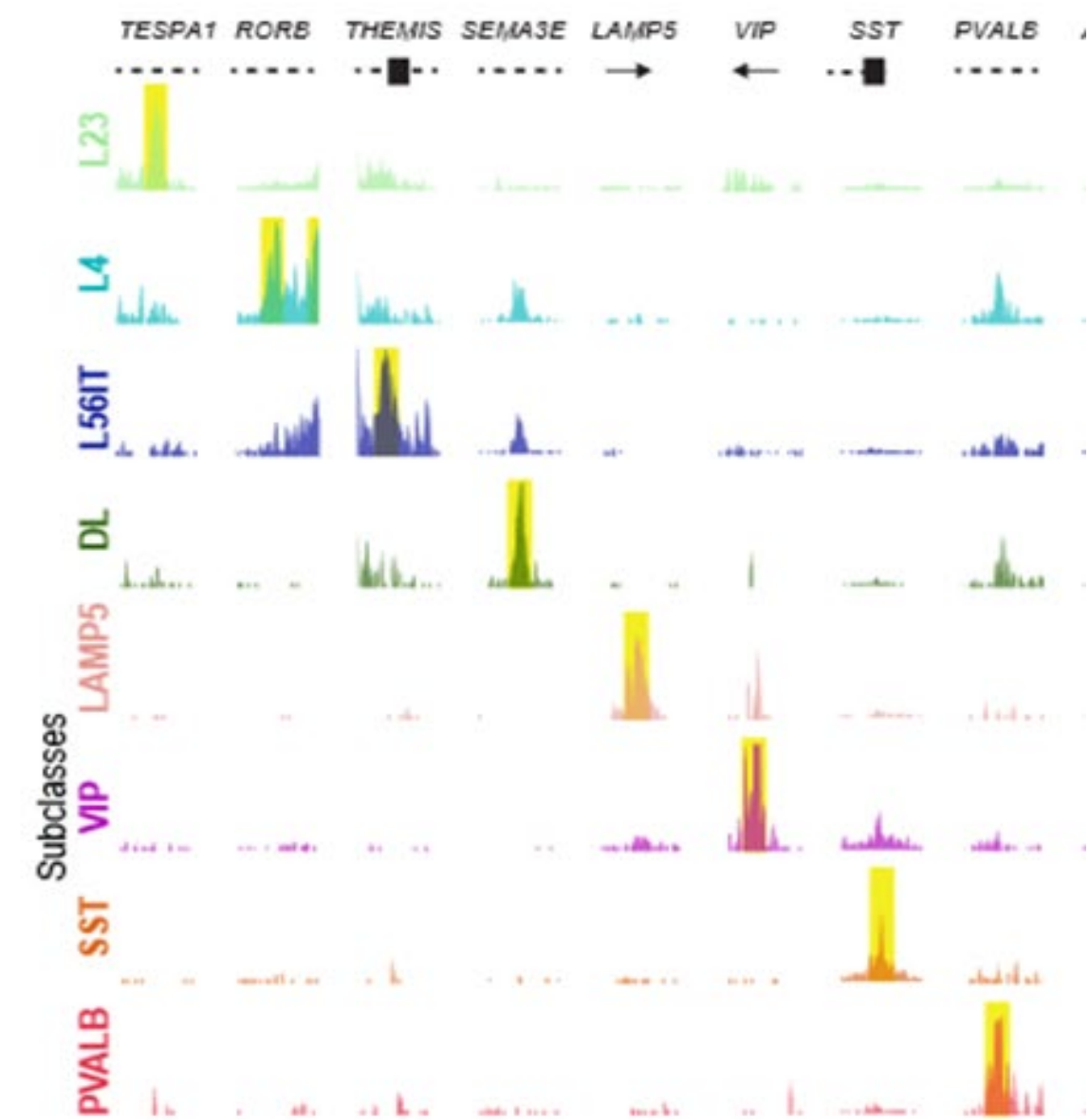
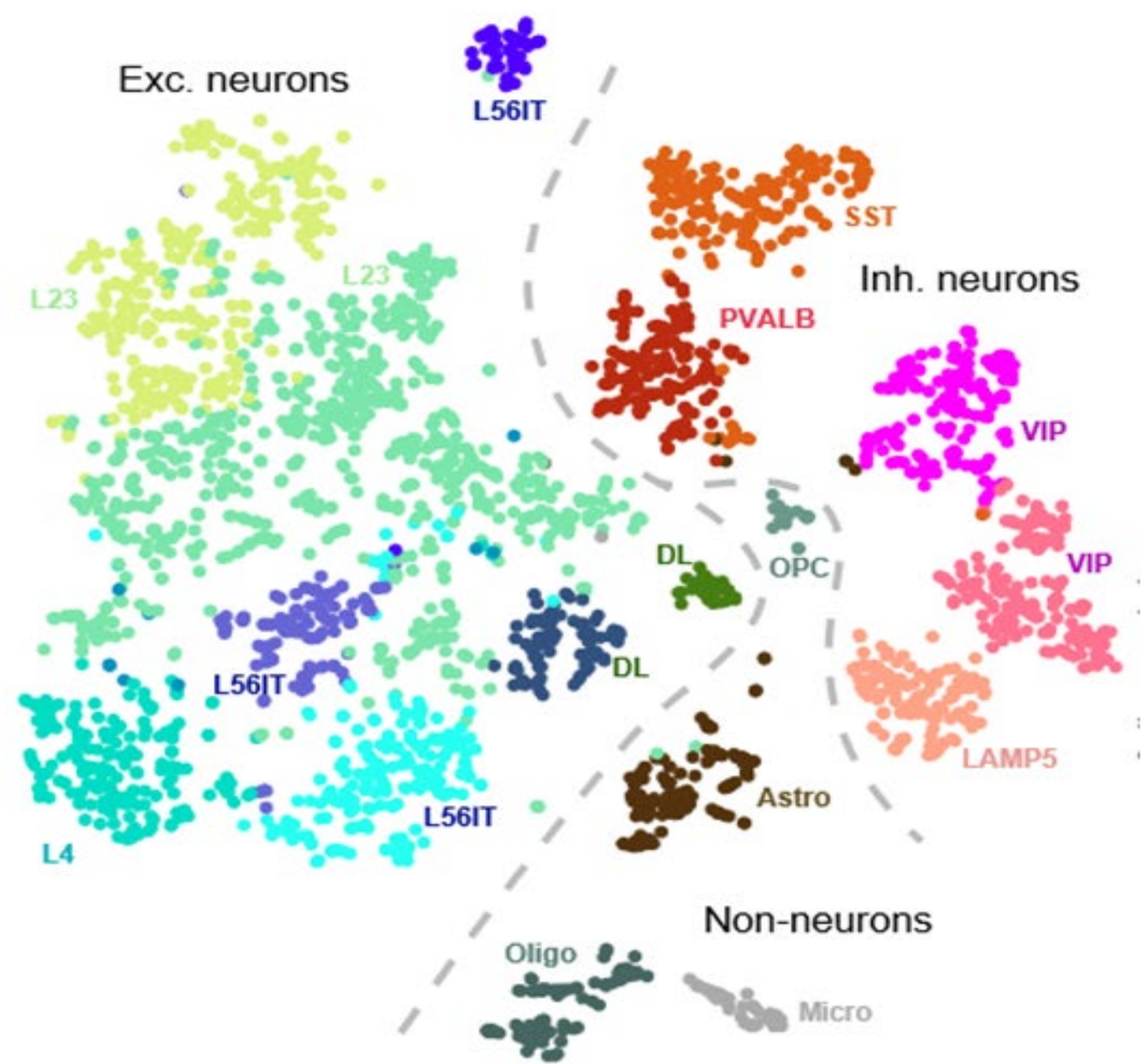
Gene regulatory regions are found in open regions of the genome that are potential “enhancers” of gene expression

Different cell types have different potential enhancers

Single cell “ATAC-Seq”

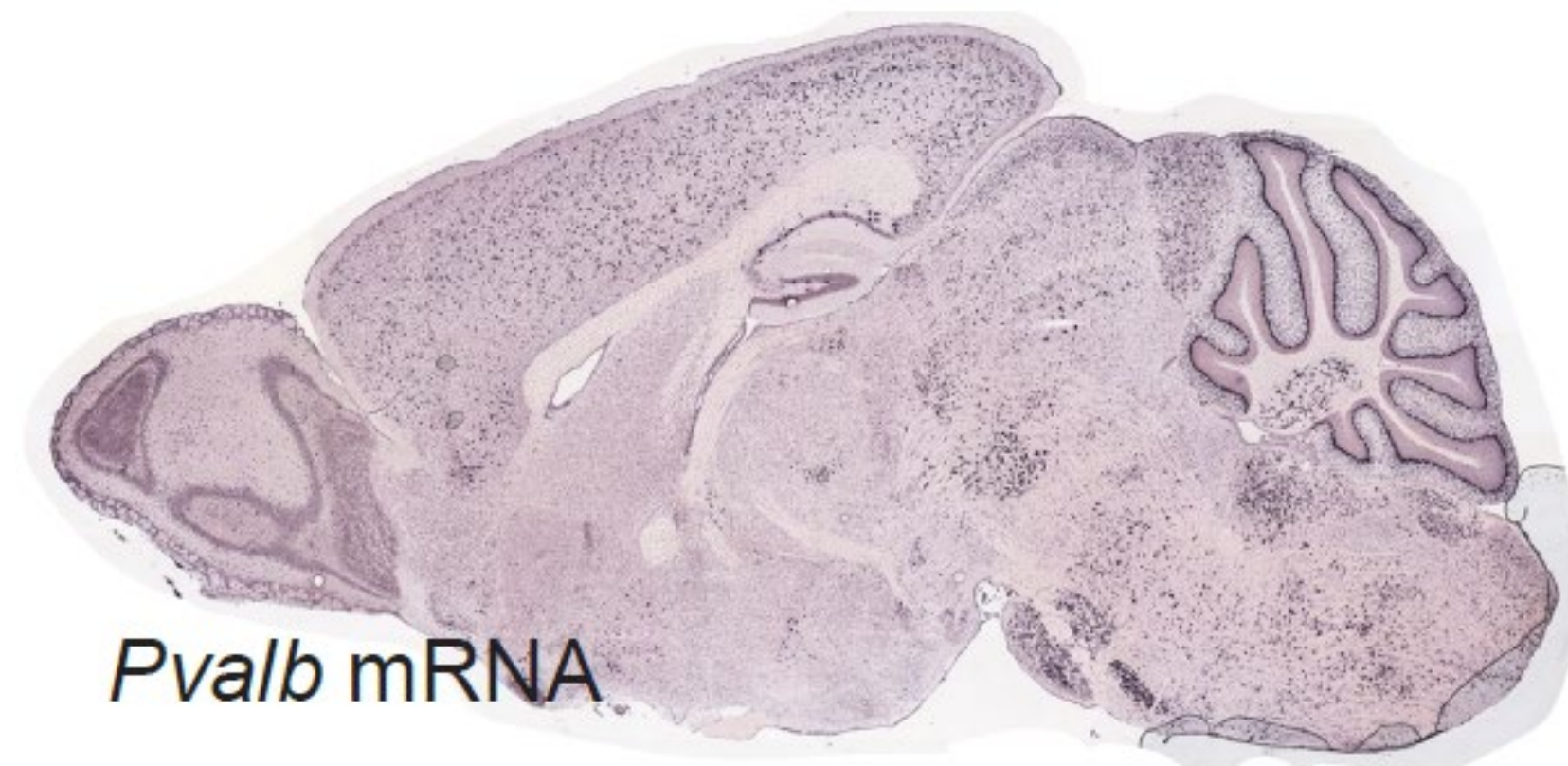


Buenrostro et al., (2013) Nat. Methods, (2015) Nature





# Future of precision medicine: Gene therapy targeted to specific cell types affected in disease

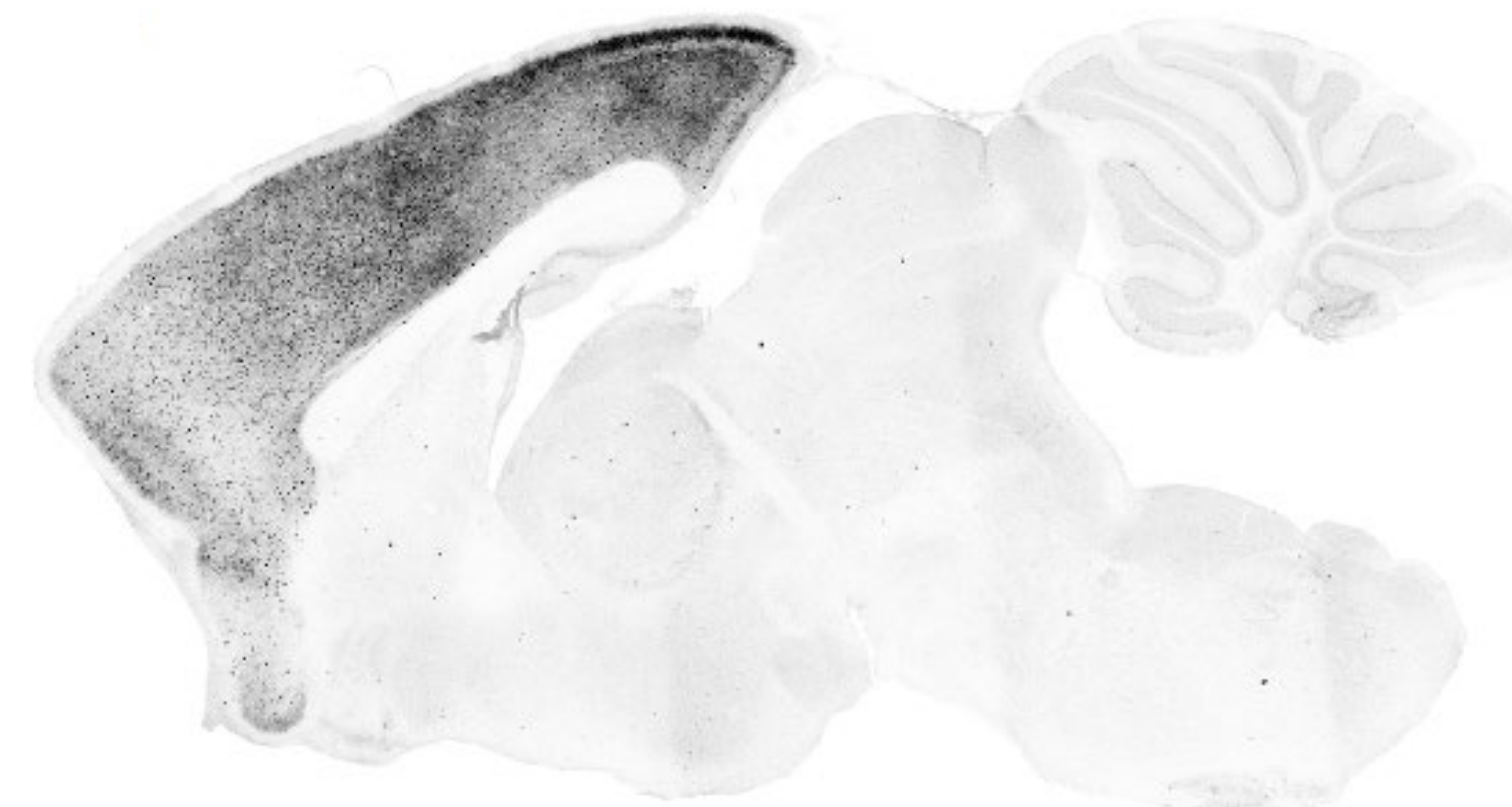


Short enhancers drive gene expression in different cell types following viral infection

eHGT\_023h



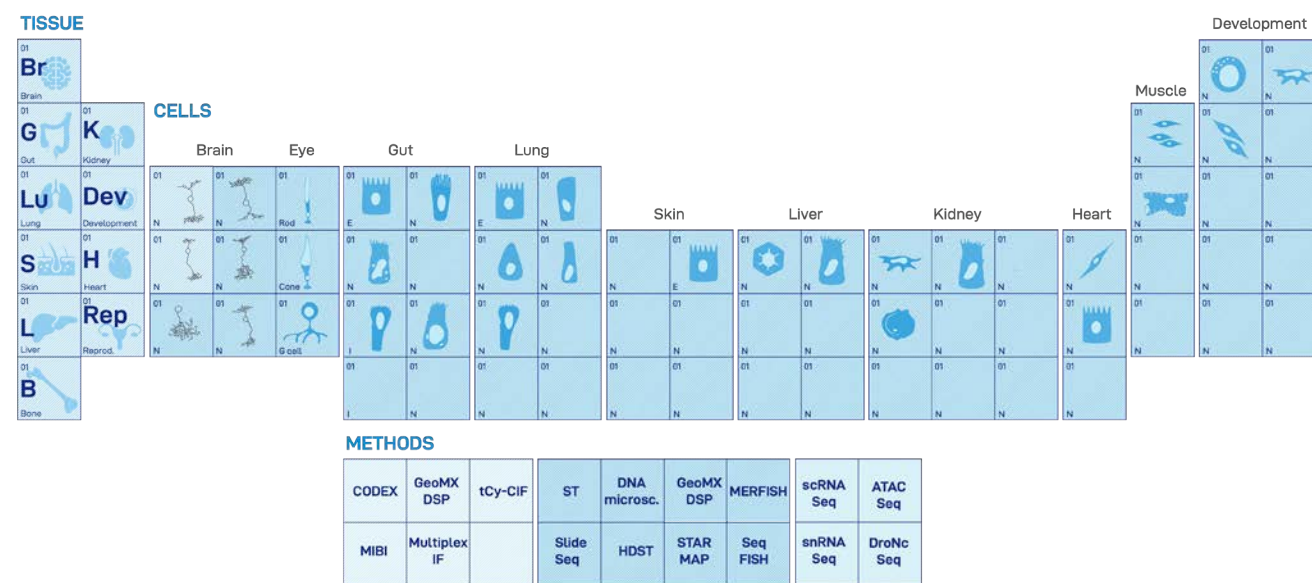
eHGT\_140h





# The brain atlas is the foundation for a new era in understanding and treating brain disease

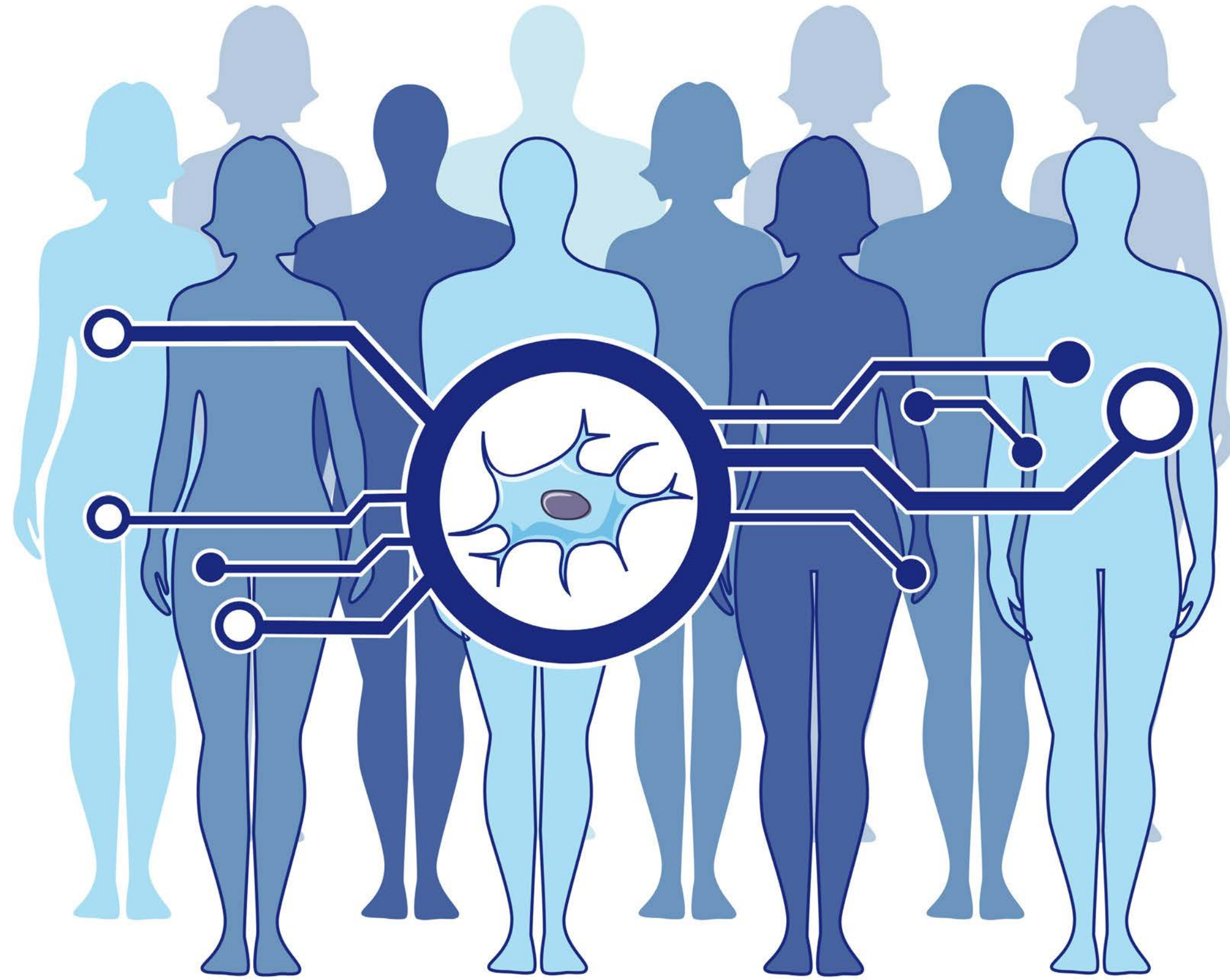
Do neurological, neuropsychiatric, or neurodegenerative diseases involve pathology of specific cell types?



The molecular tools are available now to probe this question by building on the baseline “periodic table”:

- Are some cell types selectively vulnerable or resistant?
- What molecular pathways are perturbed in which cell types?
- Where is the best cellular and molecular target for intervention, potentially using gene therapy applications?



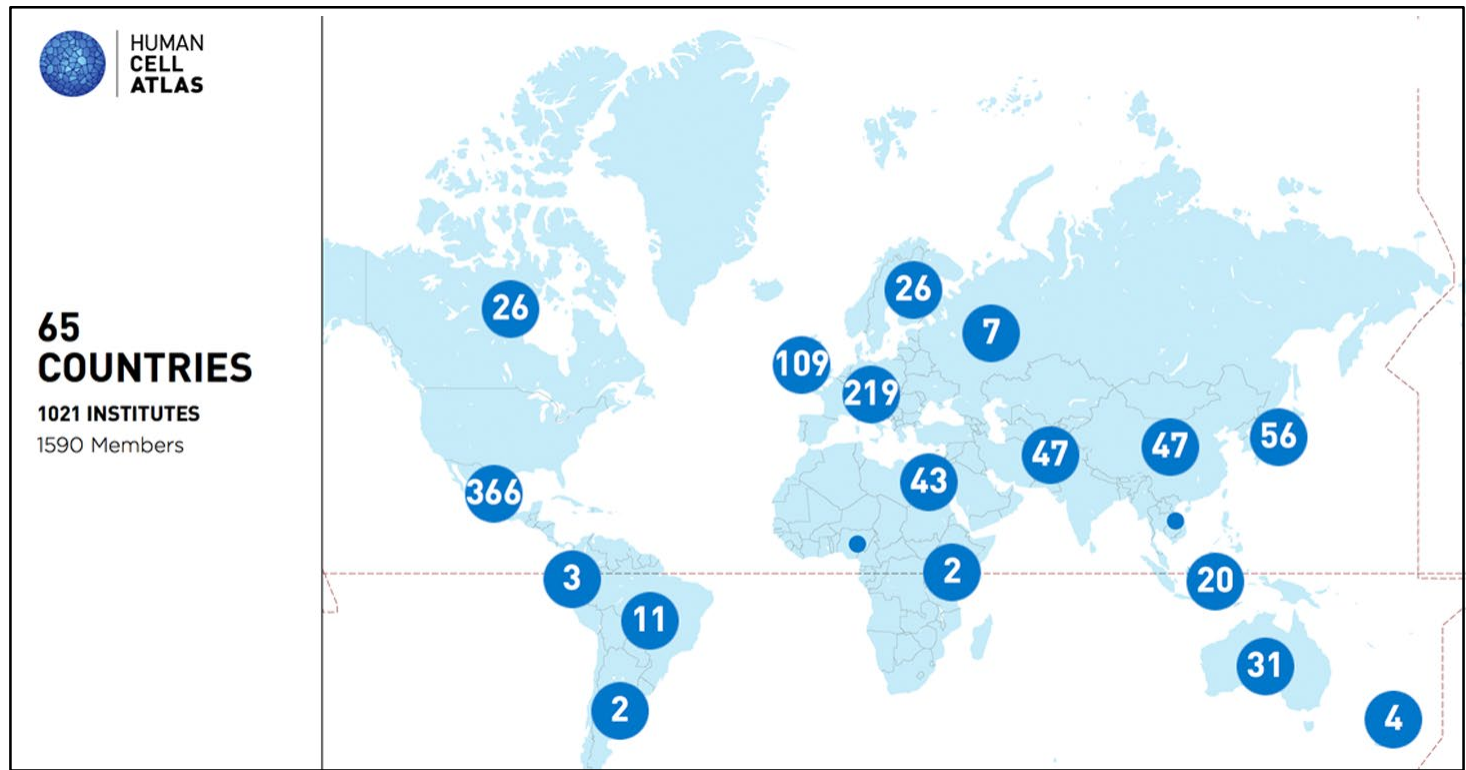


Humans are Diverse.  
So Are their Cells.  
No One Lab or Country  
Should Build the Atlas  
Alone.

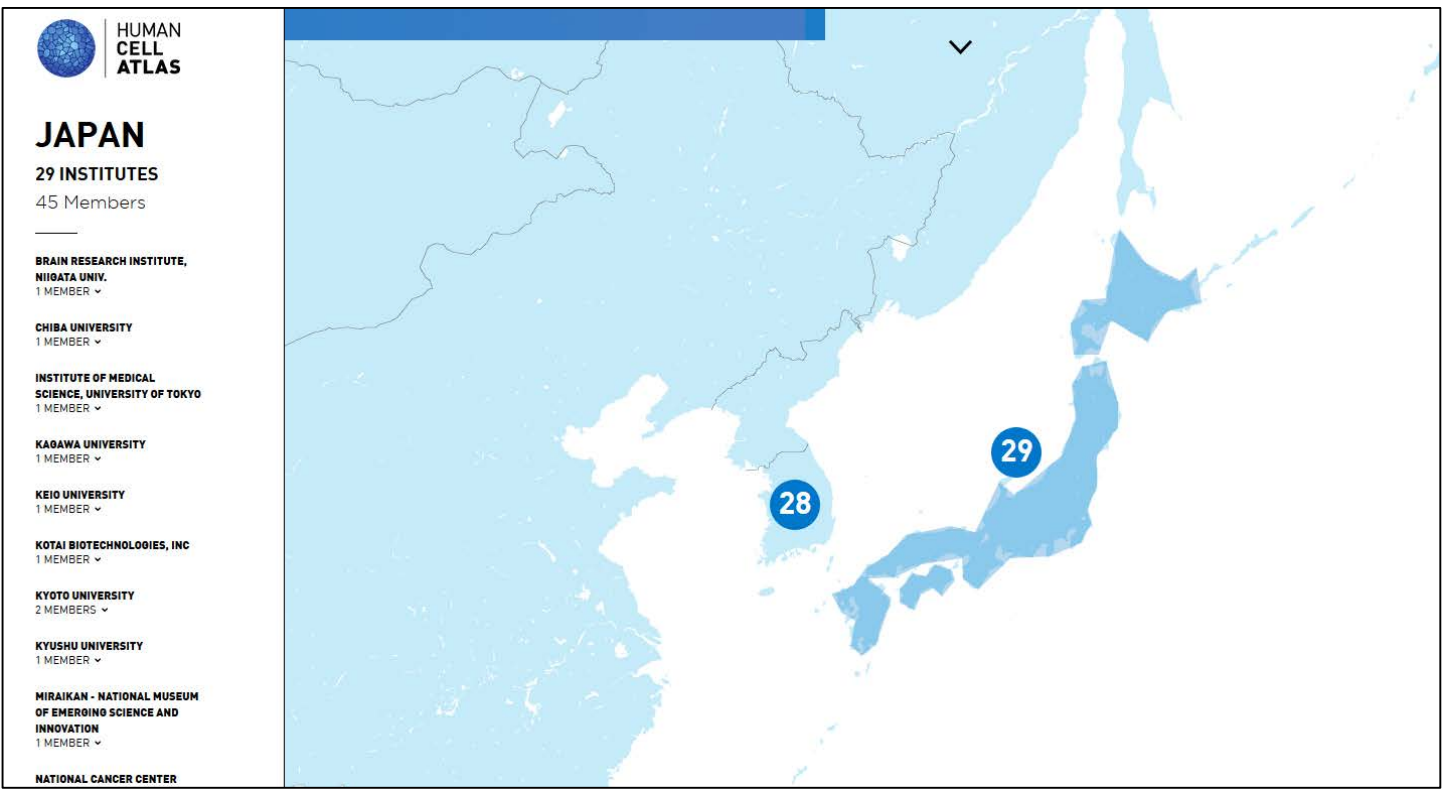


# HCA membership: global and rapidly growing

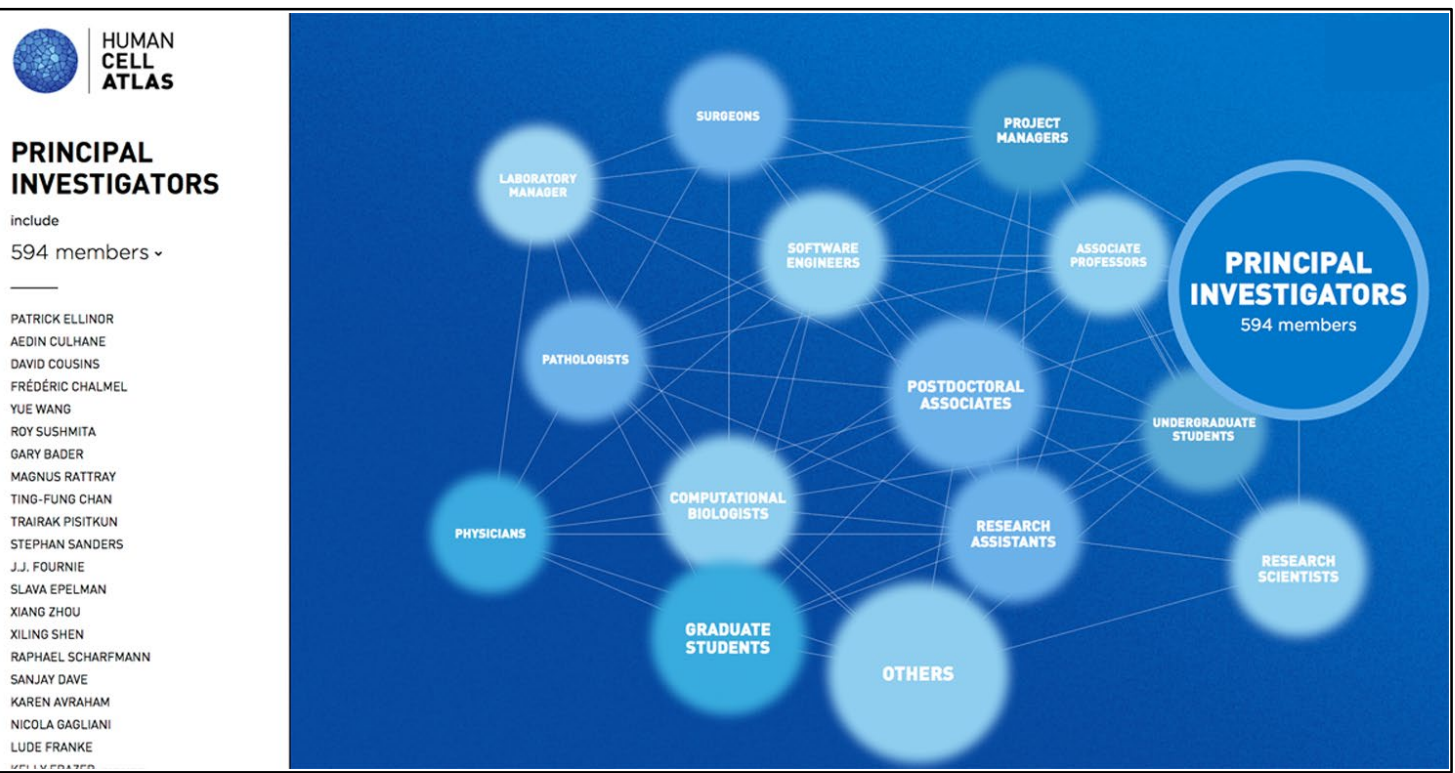
1756 Members, 70 Countries, 1030 Institutes



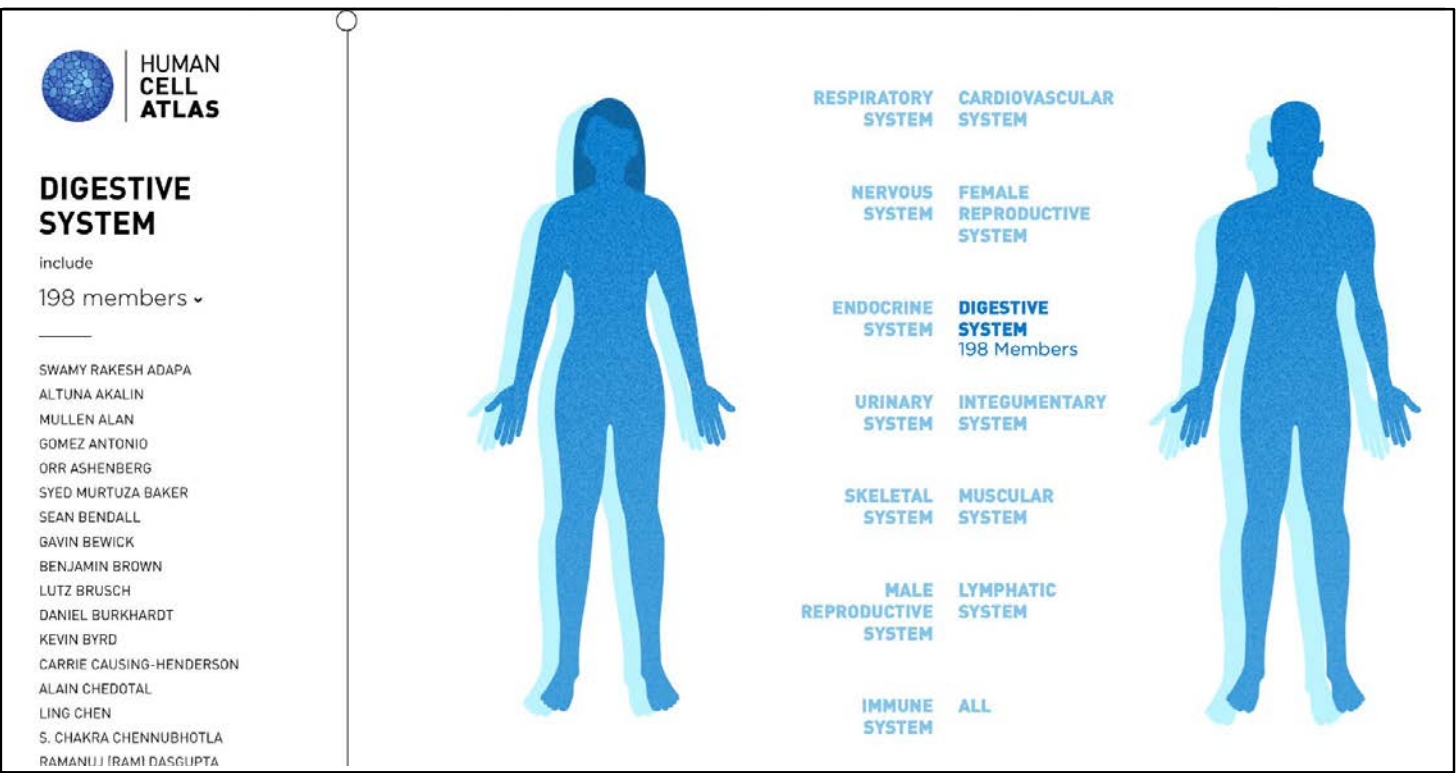
Japan: 45 Members, 29 Institutes



1756 members, 641 PI



14 organs/systems: e.g. digestive system; 198 members



<https://www.humancellatlas.org/join-hca/>



# Learning more about the HCA

<https://www.humancellatlas.org/>



## The Human Cell Atlas

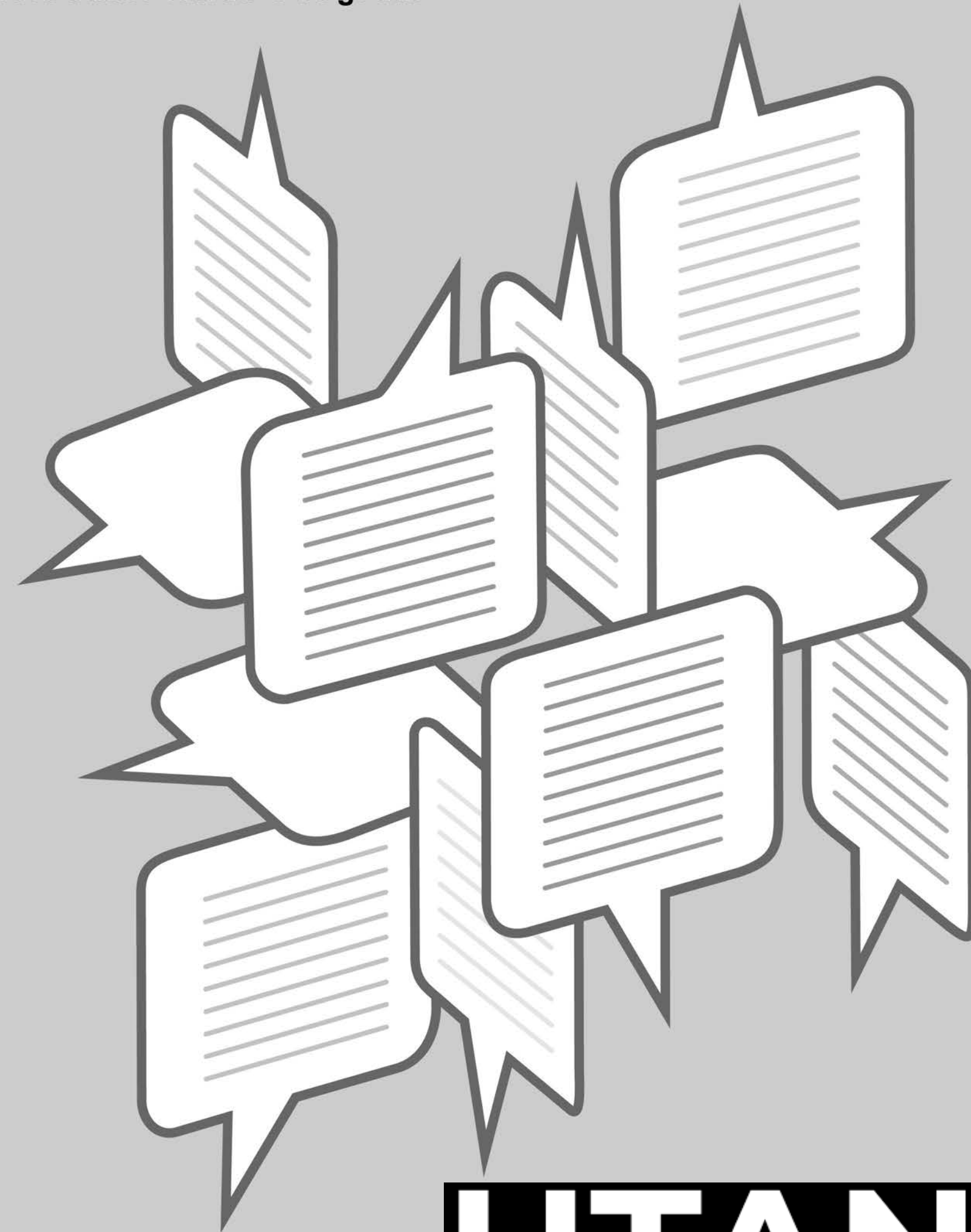
**Abstract** The recent advent of methods for high-throughput single-cell molecular profiling has catalyzed a growing sense in the scientific community that the time is ripe to complete the 150-year-old effort to identify all cell types in the human body. The Human Cell Atlas Project is an international collaborative effort that aims to define all human cell types in terms of distinctive molecular profiles (such as gene expression profiles) and to connect this information with classical cellular descriptions (such as location and morphology). An open comprehensive reference map of the molecular state of cells in healthy human tissues would propel the systematic study of physiological states, developmental trajectories, regulatory circuitry and interactions of cells, and also provide a framework for understanding cellular dysregulation in human disease. Here we describe the idea, its potential utility, early proofs-of-concept, and some design considerations for the Human Cell Atlas, including a commitment to open data, code, and community.

DOI: <https://doi.org/10.7554/eLife.27041.001>

AVIV REGEV\*, SARAH A TEICHMANN\*, ERIC S LANDER\*, IDO AMIT, CHRISTOPHE BENOIST, EWAN BIRNEY, BERND BODENMILLER, PETER CAMPBELL, PIERO CARNINCI, MENNA CLATWORTHY, HANS CLEVERS, BART DEPLANCKE, IAN DUNHAM, JAMES EBERWINE, ROLAND EILS, WOLFGANG ENARD, ANDREW FARMER, LARS FUGGER, BERTHOLD GÖTTGENS, NIR HACHOEN, MUZLIFAH HANIFFA, MARTIN HEMBERG, SEUNG KIM, PAUL KLENERMAN, ARNOLD KRIEGSTEIN, ED LEIN, STEN LINNARSSON, EMMA LUNDBERG, JOAKIM LUNDEBERG, PARTHA MAJUMDER, JOHN C MARIONI, MIRIAM MERAD, MUSA MHLANGA, MARTIJN NAWIJN, MIHAI NETEA, GARRY NOLAN, DANA PE'ER, ANTHONY PHILLIPAKIS, CHRIS P PONTING, STEPHEN QUAKE, WOLF REIK, ORIT ROZENBLATT-ROSEN, JOSHUA SANES, RAHUL SATIJA, TON N SCHUMACHER, ALEX SHALEK, EHUD SHAPIRO, PADMANEE SHARMA, JAY W SHIN, OLIVER STEGLE, MICHAEL STRATTON, MICHAEL J T STUBBINGTON, FABIAN J THEIS, MATTHIAS UHLEN, ALEXANDER VAN OUDENAARDEN, ALLON WAGNER, FIONA WATT, JONATHAN WEISSMAN, BARBARA WOLD, RAMNIK XAVIER, NIR YOSEF AND HUMAN CELL ATLAS MEETING PARTICIPANTS

<https://www.humancellatlas.org/publications/> and: URL: <https://arxiv.org/abs/1810.05192v1>





## Partnership across programs and funders



- Related and complementary initiatives
- Diverse funded data collected projects across the globe
- Support for central efforts: DCP, meetings, ethics, equity
- Discounts to help enable research:

- Biolegend: Discount on reagents
- Nanostring: Discount of reagents, instruments
- Takara: Discount on reagents
- 10x Genomics: New discount plan

